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An annotated checklist of Gelechiidae (Lepidoptera) of Israel with description of two new species

OLEKSIY BIDZILYA¹, OLE KARSHOLT², VASILIIY KRAVCHENKO³ & JAN ŠUMPICH⁴

¹*Institute for Evolutionary Ecology of the National Academy of Sciences of Ukraine, 37 Academician Lebedev str., 03143, Kiev, Ukraine. E-mail: olexbid@gmail.com*

²*Zoological Museum, Natural History Museum of Denmark, Universitetsparken 15, DK-2100 Copenhagen, Denmark. E-mail: okarsholt@snm.ku.dk*

³*Department of Zoology, The George S. Wise Faculty of Life Sciences, Tel Aviv University, Ramat Aviv, 69978, Israel. E-mail: vasiliiy1953@yandex.com*

⁴*National Museum, Natural History Museum, Department of Entomology, Cirkusová 1470, CZ-193 00 Praha 9 - Horní Počernice, Czech Republic. E-mail: jansumpich@seznam.cz*



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OLEKSIY BIDZILYA, OLE KARSHOLT, VASILIJ KRAVCHENKO & JAN ŠUMPICH

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Abstract

One hundred forty-six species of Gelechiidae including 36 new records are reported from Israel. *Anarsia balioneura* Meyrick, 1921 and *Polyhymno chionarcha* Meyrick, 1913 are recorded for the first time in the Palaearctic region. Two new species are described: *Metzneria freidbergi* sp. nov., and *Scrobipalpa aravensis* sp. nov. Six new synonyms are established: *Stygmatoptera* Hartig, 1936 **syn. nov.** of *Polyhymno* Chambers, 1874; *Eulamprotes* Bradley, 1971 **syn. nov.** of *Oxypteryx* Rebel, 1911; *Polyhymno abaiella* Amsel, 1974 **syn. nov.** of *Polyhymno chionarcha*, Meyrick, 1913; *Gelechia haifella* Amsel, 1935 **syn. nov.** of *Athrips rancidella* (Herrich-Schäffer, 1854); *Sophronia catharurga* Meyrick, 1923 and *Sophronia parahumerella* Amsel, 1935 **syn. nov.** of *Pseudosophronia exustellus* (Zeller, 1847). The following new combinations are proposed: *Anacampsis karmeliella* (Amsel, 1935) **comb. nov.**, *Stomopteryx tesseraepunctella*

(Amsel, 1935) **comb. nov.**, *Aproaerema languidella* (Amsel, 1936) **comb. nov.**, *Aproaerema telaviviella* (Amsel, 1935) **comb. nov.**, *Acompsia (Telephila) ballotellus* (Amsel, 1935) **comb. nov.**, *Polyhymno dumonti* (Hartig, 1936) **comb. nov.**, *Oxypteryx atrella* (Denis & Schiffermüller, 1775) **comb. nov.**, *Oxypteryx immaculatella* (Douglas, 1850) **comb. nov.** and *Chrysoesthia amseli* (Bidzilya, 2008) **comb. nov.** A lectotype is designated for *Lita rhamnifoliae* Amsel & Hering, 1931. The genitalia of both sexes of *Sophronia sagittans* Meyrick, 1923, *Anacampsis karmeliella* (Amsel, 1935), *Stomopteryx tesserapunctella* Amsel, 1935 as well as male genitalia of *Stomopteryx lacteolella* Caradja, 1924, *Aproaerema telaviviella* (Amsel, 1935), *Acompsia ballotellus* (Amsel, 1935), *Polyhymno dumonti* (Hartig, 1936) and *Chrysoesthia amseli* (Bidzilya, 2008) are illustrated and described for the first time. New or additional host plants are recorded for *Metzneria aspretella* Lederer, 1869, *M. agraphella* (Ragonot, 1895), *M. ehikeella* Gozmány, 1954 and *Scrobipalpa suaedivorella* (Chrétien, 1915). Photographs of the type specimens of most taxa described from Israel and Palestine are presented. The following species are removed from the list of Gelechiidae of Israel: *Nothris sulcella* Staudinger, 1879, *N. skyvai* Karsholt & Šumpich, 2015, *Anarsia spartiella* Schrank, 1802, *Megacraspedus cerussatellus* Rebel, 1930, *Oxypteryx atrella* (Denis & Schiffermüller, 1775), *Isophrictis anthemidella* (Wocke, 1871), *Metzneria metzneriella* (Stainton, 1851), *Scrobipalpa otregata* Povolný, 1972, *Scrobipalpa nitentella* (Fuchs, 1902), *Scrobipalpa remota* Povolný, 1972, *Scrobipalpa salinella* (Zeller, 1847) and *Ephysteris diminutella* (Zeller, 1847). Moreover, *Stomopteryx remissella* (Zeller, 1847) is recorded as new to the Altai Mountains of Russia, *Anarsia balioneura* Meyrick, 1921 is new to Cyprus and Libya, *Polyhymno dumonti* (Hartig, 1936) is new to Libya and Sudan, *Scrobipalpa superstes* is new to Greece, *Stenolechia gemmella* (Linnaeus, 1758) is new to Jordan and *Polyhymno chionarcha* is new to Saudi Arabia.

Key words: New species, new synonyms, new records, new combinations, host-plants, distribution, Palaearctic region

Introduction

Israel is located in the eastern part of the Mediterranean basin which lies in the northern part of the East African Rift Valley. The geography of the country is mainly determined by its position within the Mediterranean zone. Israel is a “crossroad” between three continents: Europe, Asia and Africa. In contrast to the more uniform and monotonous landscapes of the Levant, Israel is topographically distinctive with a wide variety of different zones and habitats. The northern and Mediterranean parts of the country include the southern tip of the Anti-Lebanon ridge with elevations up to 2200 m above sea-level, bearing the forested Mediterranean north, while in the south extreme deserts are situated around the Dead Sea and Arava valley, with pockets rich in Afrotropical fauna and flora (Zohary & Orhansky 1949).

The first data on Gelechiidae from territories of modern Palestine and Israel were published by Stainton (1867) including six species from Jaffa and the “plains of Jordan”. Two of these, *Gelechia nitidula* and *Gelechia subdiminutella* were the first taxa he described. Later two new species were described by Rebel (1902, 1926) and four by Meyrick (1923). The next contribution to the Gelechiidae from Palestine was made by Bodenheimer (1930a). The most considerable contribution to Gelechiidae from Palestine was made by Hans Georg Amsel. It included descriptions of three new taxa (Amsel & Hering 1931) followed by a list of 78 gelechiid species published in a monograph “Die Lepidopteren Palästinas” (Amsel 1933). Most species of Gelechiidae named by Amsel in this paper for the first time were formally described in a series of subsequent publications (Amsel 1935a,b,c). All these data were summarized in “Prodromus Faunae Palaestinae” (Bodenheimer 1937) which comprises 88 species of Gelechiidae (including a few species listed twice). Later two additional species from Israel and Palestine were described by Amsel (1942, 1952). The systematic status of several taxa described from this region was changed, some records were corrected and additional new species were described in the revisions of corresponding genera (Sattler 1967, 1976, 1979, 1992; Karsholt & Rutten 2005). The tribe Gnorimoschemini was revised and completed by description of new species and new records by Povolný (1972, 2002) and later by Huemer & Karsholt (2010). An additional new species of *Istrianis* was described recently (Bidzilya & Karsholt 2015). Pests of crops, stored products, forests and ornamental trees were reviewed in a number of publications (Bytinski-Salz 1966; Yathom & Meisner 1966; Bytinski-Salz & Sternlicht 1967; Donahaye & Calderon 1969; Halperin 1986; Halperin & Sauter 1992 and others). The tomato leafminer *Tuta absoluta* (Meyrick, 1917) was recorded for the first time from Israel in 2010 (Seplyarsky *et al.* 2010).

The specimens on which Stainton (1867) and Meyrick (1923, 1931) based their new species are kept in the Natural History Museum in London, and most specimens collected and described by Amsel are now in the Staatliches Museum für Naturkunde Karlsruhe. However, type specimens of some of Amsel’s Gelechiidae are deposited

in several other museums (e.g. Deutsches Kolonial- und Übersee-Museum in Bremen and Zoologisches Museum, Humboldt University, Berlin), and some could not be located. Most recently collected material of Israeli Gelechiidae is kept in the University of Tel Aviv.

Despite considerable progress in the study of Gelechiidae of Israel and Palestine, knowledge of the fauna of this region remains rather unsatisfactory both taxonomically and faunistically. Some genera remain unrevised, which considerably hinders biodiversity studies. It is especially true for such genera as *Aristotelia* Hübner, [1825], *Isophrictis* Meyrick, 1917, *Metzneria* Zeller, 1839 and *Aproaerema* Durrant, 1897. Altogether 155 species of Gelechiidae are recorded from Israel and Palestine. This number seems to be far from complete when one considers the rich diversity of landscapes of the region. Beside the material listed below, we examined several specimens which could not be identified—either because they were in bad condition or because they belong to unrevised genera. Therefore the specimens cannot be identified at the moment. Such specimens are not dealt with here, but are left for future studies.

This contribution aims to revise and summarise all data on Gelechiidae from Israel, provide new records, clarify the taxonomic state of some taxa and describe new species. Most publications of Lepidoptera from before World War II referred to the area of present-day Israel as “Palestine” or “Palestina”, and although Amsel (1935c: 228) gave a list of some of the localities where he had collected, it is not always possible to refer data from earlier publications, including data of type specimens, to exact places within (or outside of) Israel, in as much as the borders are not internationally agreed on. In using the term Palestine we are referring to data from literature or specimen data, and we are not taking part of any disagreements about the political borders in the area. The disputed area of the Golan Heights, as well as Gaza Strip and West Bank are included within the broad definition of “Israel” used in the paper.

Material and methods

The paper is based on materials deposited in the collection of the Steinhardt Museum of Natural History in Tel Aviv University, Israel (TAU), Staatliches Museum für Naturkunde Karlsruhe, Germany (SMNK), National Museum Prague, Czech Republic (NMPC), Natural History Museum, London, U.K. (NHMUK), Naturhistorisches Museum, Vienna, Austria (NHMW), Museum für Naturkunde, Berlin (MfN), Finnish Museum of Natural History, Helsinki, Finland (MZH), Übersee-Museum Bremen, Germany (UMB), Zoological Museum Kiev Taras Shevchenko National University, Ukraine (ZMKU) and Zoological Museum, Natural History Museum of Denmark, Copenhagen, Denmark (ZMUC).

The adults were collected by light trap or netting. We list all material of Gelechiidae examined by us from Israel and Palestine. In most cases we state sexes of the specimens, but for a large material collected by light traps and pinned, but not spread, we only give the number of specimens (abbreviated as ‘ex’), as it is not possible to determine the sex without damaging the specimens. Male and female genitalia were dissected and prepared using standard methods (Huemer and Karsholt 1999). Pinned specimens were photographed with an Olympus E-410 digital camera attached to an Olympus SZX12 microscope or with a Canon 750D in the combination of a Canon MP-E-65 mm lens. Slide-mounted genitalia were photographed with a Canon EOS 600D digital camera mounted on an Olympus U-CTR30-2 trinocular head combined with a Carl Zeiss microscope body or with a Canon EOS 1100D camera mounted on an Olympus BX41 stereomicroscope. Sets of 10–60 images were taken for each specimen and assembled to deep-focused images using Helicon Focus 6 and edited in Adobe Photoshop CS5.

The system of Gelechiidae follows Karsholt *et al.* (2013). The order of species within genera follows their revisions (Sattler 1967, 1976; Bidzilya & Karsholt 2015; Karsholt & Šumpich 2015 and others). For unrevised genera the species are arranged tentatively based on similarity of the genitalia characters. The nomenclature of the host-plant follows Euro+Med Plantbase (<http://www.emplantbase.org>). Incorrect and doubtful records for Israel are listed separately.

Results

Subfamily Anacampsinae Bruand, 1851

Tribe Anacampsini Bruand, 1851

Mesophleps trinitella Herrich-Schäffer, 1856

Mesophleps trinitella Herrich-Schäffer, 1856—Neue Schmett. (1): 6, fig. 46.
Xystophora aurantiella Rebel, 1915—Rovart. Lapok 22: 178.
Crossobela barysphenata Meyrick, 1923—Exot. Microlep. 3 (1–2): 34.
Batrachedra subtilipennis Turati, 1924—Atti Soc. ital. Sci. nat. 63: 180, pl. 6, fig. 18.

Records. Amsel 1933: 126; Amsel, 1935c: 264; Bodenheimer 1937: 101.

Material examined. 2 ♂, Keafar Shemal, 15.iv.1968 (collector unknown); 1 ♂, Holom, 19.iv.1968 (collector unknown) (all TAU); 2 ♀, East bank of Kenneret lake, Mevo Hama, 18.iv.2018, light (Kravchenko & Bidzilya) (TAU, ZMKU); 1 ♀, Israel, Golan Heights, Masd al Shams, oak forest, 26.viii.2000 (Pavliček) (NMPC).

Distribution. South and Central Europe to Lower Volga and East Caucasus, Turkey, Cyprus, Morocco, Algeria, Libya, Turkmenistan (Ivinskis & Piskunovas 1994: 39; Karsholt & Riedl 1996: 120; Ponomarenko 2008: 103; Li & Sattler 2012: 39), Israel.

Mesophleps ochracella (Turati, 1926)

Nothris ochracella Turati, 1926—Atti Soc. ital. Sci. nat. 65: 69, fig. 32.

Material examined. 1 ♀, 1 ♂, Nahal Oren, Mt. Carmel, light trap 24.vi.1997 (Pavliček & Kravchenko) (gen. prep. 19648 and 19655, J. Šumpich); 1 ♀, ditto but 1.viii.1997 (gen. prep. 19654, J. Šumpich) (all NMPC).

Distribution. Spain, South France, Malta, Greece, Morocco, Algeria, Libya and perhaps Turkmenistan (Li & Sattler 2012: 40), Israel (new record).

Remarks. The specimens from Israel are here tentatively referred to *M. ochracella*. They differ in small details from the photographs of this species by Li & Sattler (2012). We find it likely that *M. ochracella* includes cryptic diversity, which is beyond the scope of the present paper to revise.

Mesophleps silacella (Hübner, 1796)

Tinea silacella Hübner, 1796—Samml. eur. Schmett. [8]: 37, pl. 17, fig. 117.
Tinea pyropella Hübner, 1793—Vögel und Schmett.: fig. 95.
Mesophleps pudicellus var. *apicellus* Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 113.
Mesophleps silacellus calaritanus Amsel, 1939—Mem. Soc. ent. ital. 17 (1938): 73.

Records. Amsel 1933: 126; Amsel 1935c: 264; (partim as *corsicella*); Bodenheimer 1937: 101.

Material examined. 1 ♂, Ramallah, Jerusalem. 21.iv.1930 (Amsel); 2 ♀, Kirjat Anawim, Jerusalem, 22.iv.1930 (Amsel) (SMNK); 2 ♂, Tel Aviv vic., Cesaria, Dunes, 14.iv.2018, day collecting (Bidzilya) (TAU, ZMKU).

Distribution. Europe eastwards to Middle Volga region, Turkey, Algeria, Turkmenistan (Ivinskis & Piskunovas 1994: 38; Ponomarenko 2008: 103; Li & Sattler 2012: 34), Israel.

Mesophleps corsicella Herrich-Schäffer, 1856

Mesophleps corsicella Herrich-Schäffer, 1856—Neue Schmett. (1): 7.
Mesophleps lala Agenjo, 1960—Verh. XI. int. Kongr. Ent. Wien 1: 150.

Records. Amsel 1933: 126; Amsel 1935c: 264; Bodenheimer 1937: 101; Halperin & Sauter 1992: 134.

Material examined. 2 ♀, Kirjat Anawim, Jerusalem, 22.iv.????, 30.v.1930 (Amsel) (SMNK); 1 ♂, 1 ♀, Tel-Aviv, Botanical garden, 14.iv.2018 (Bidzilya & Kravchenko) (ZMKU); 1 ♂, 28 exx, Jerusalem, 6.vi.2004 (Kravchenko)

(all TAU, ZMUC); 1 ♂, Nahal Oren, Mt. Carmel, light trap 17.v.1997 (Pavliček & Kravchenko); 1 ♀, ditto but 28.iii.1998 (gen. prep. 19647, J. Šumpich); 1 ♂, 1 ♀, ditto but 23.iv.1998; 1 ♀, ditto but 21.viii.1998; 1 ♂, ditto but 5.vii.1999 (all NMPC).

Host plant. In Israel the larvae have been recorded feeding in fruits of *Cistus creticus* L. (Halperin & Sauter 1992: 134).

Distribution. Spain (including Balearic Islands), Portugal, France (including Corsica), Italy (including Sardinia and Sicily), Greece, Morocco, Lebanon (Karsholt & Riedl 1996: 120; Li & Sattler 2012: 35), Israel.

Pseudosphronia exustellus (Zeller, 1847)

Ypsolophus exustellus Zeller, 1847—*Isis* von Oken 1847 (11): 820.

Sophronia catharurga Meyrick, 1923, **syn. nov.**—*Exot. Microlep.* 3 (1–2): 34.

Sophronia parahumerella Amsel, 1935, **syn. nov.**—*Mitt. zool. Mus. Berl.* 20 (2): 297.

Sophronia buvati Nel, 1998—*Alexanor* 20(4): 228, figs 17–23.

Records. Amsel 1933: 126; Amsel 135a: 297, pl. 11, fig. 95; Amsel 1935c: 265; Bodenheimer 1937: 102 (*parahumerella*; *catharurga*); Bytinski-Salz & Sternlicht 1967: 130; Halperin & Sauter 1992: 122.

Material examined. Holotype of *S. parahumerella* ♀, Tel Aviv, Paläs., Düne, 17.ix.1930 (Amsel) (SMNK); 2 ♂, 1 ♀ Kefar Shemuel, 15.iv.1968 (gen. slide 128/18♂, 129/18♀, O. Bidzilya (TAU); 4 ♂, Jericho, Kalia, v.2004 (Kravchenko) (TAU, ZMUC).

Remarks. Specimens from Israel match both externally and genitally (Figs 1–4) *Pseudosphronia exustellus* (Nel 1998; Corley 2001). They also agree in all details with the female holotype of *Sophronia catharurga* collected in Akka, Israel (Clarke 1969: 376, pl. 188, figs 1–1b). The female holotype of *Sophronia parahumerella* described from the vicinity of Tel Aviv is identical to *P. exustellus* too (Fig. 2). Hence, we propose the following synonymy: *Sophronia catharurga* Meyrick, 1923 and *Sophronia parahumerella* Amsel, 1935 **syn. nov.** of *Pseudosphronia exustellus* (Zeller, 1847).

Host plant. Unknown. The record of *Quercus ithaburensis* Decne. as a host plant for larvae of *P. exustellus* in Israel (Bytinski-Salz & Sternlicht 1967: 130; Halperin & Sauter 1992: 122) requires verification.

Distribution. Spain, Portugal, France, Italy (including Sicily) (Karsholt & Riedl 1996: 118), Israel.

Sophronia sagittans Meyrick, 1923

Sophronia sagittans Meyrick, 1923—*Exot. Microlep.* 3 (1–2): 33.

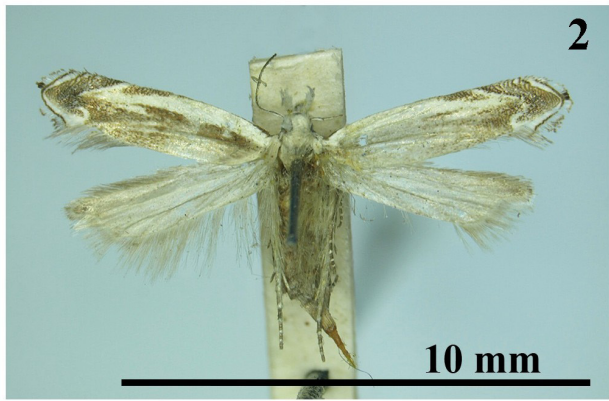
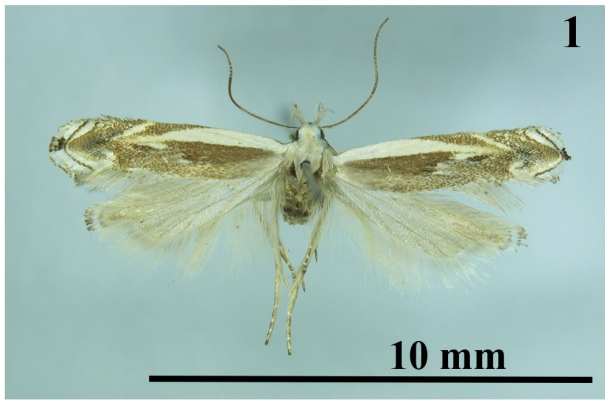
Sophronia karmeliella Amsel, 1933—*Zoogeograph.* 2 (1): 126 (nomen nudum).

Records. Amsel, 1933: 126; Amsel, 1935c: 264, 265; Bodenheimer 1937: 102; Clarke 1969: 376, pl. 188, fig. 3.

Material examined. 2 ♂, 3 ♀, Karmel, Haifa, 7.v.1930 (Amsel) (SMNK); 1 ♀, Kefar Shemuel, 15.iv.1968; 1 ♂, 1 ♀, Latraca, 25, 27.iv.1968 (TAU); 3 ♂, 10 ♀, East bank of Kenneret lake, Mevo Hama, 18.iv.2018 (gen. slide 117/18♀, 118/18♂, O. Bidzilya) (Bidzilya & Kravchenko) (TAU, ZMKU).

Remarks. *Sophronia sagittans* was described from three specimens (male and female syntypes) collected by P. A. Buxton in Haifa in May. Our material agrees in all details with the photograph of the “Type” of *S. sagittans* in Clarke (1969: 376) who wrote that both male and female lack abdomen. Amsel (1933) listed the species as *Sophronia karmeliella*, without giving a description, and later (1935: 297) he stated that it belonged to *S. sagittans*. Here we provide the first description of the genitalia of both sexes.

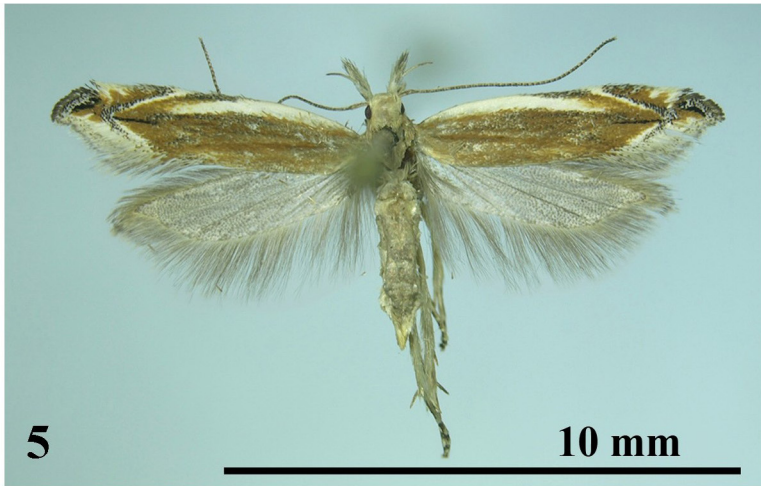
Male genitalia (Fig. 6). Uncus sub-rectangular, longer than broad, weakly rounded posteriorly; gnathos long, stout, sickle-shaped; tegumen elongated, anteromedial emargination extending to 1/3 its length; valva slender, strongly broadened distally, apex rounded, extending to half length of uncus; sacculus absent; juxta lobes projecting to 2/3 length of valva, dorsal surface densely covered with microspines in distal portion; saccus short, rectangular; phallus as long as tegumen and uncus, gradually broadened to mid length then narrowed distally, apex weakly broadened with three short rounded humps.



FIGS 1–4. *Pseudosphronia exustellus* Z. 1. Adult, Israel, ♀ (gen. slide 129/18, O. Bidzilya). 2. Adult, Israel, ♀ (holotype of *S. parahumerella*). 3. Male genitalia (gen. slide 128/18, O. Bidzilya). 4. Female genitalia (gen. slide 129/18, O. Bidzilya).

Female genitalia (Fig. 7). Papillae anales sub-ovate; ovipositor long; apophyses posteriores very long and narrow, extending far beyond the bottom of corpus bursae; segment VIII three times as long as broad, evenly sclerotized, antrum subrectangular, half length of segment VIII, proximal 2/3 laterally sclerotized, with distinct transition to ductus bursae; ductus bursae narrow, gradually broadened towards rounded corpus bursae; signum narrow, straight, pointed, slightly less than half length of ductus bursae, basal plate prolonged.

Distribution. Israel.



FIGS 5–7. *Sophronia sagittans* Meyr. 5. Adult, ♀. 6. Male genitalia (gen. slide 118/18, O. Bidzilya). 7. Female genitalia (gen. slide 117/19, O. Bidzilya).

Remarks. *Sophronia sagittans* (Fig. 5) superficially resembles *S. sicariellus* (Zeller, 1839) but the white pattern in *S. sagittans* arises nearly from base rather than from middle of costal margin in *S. sicariellus*. The slender valva in combination with the spindle-shaped phallus distinguish *S. sagittans* from its congeners (valva of *S. sicariellus* is thicker, phallus of *S. chilonella* (Treitschke, 1833) is not narrowed on basal portion). The female genitalia are defined by the long and narrow signum and long, laterally sclerotized antrum.

Anacamptis karmeliella (Amsel, 1935) comb. nov.

Tachyptilia karmeliella Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 299, Pl. 10, fig. 70.

Records. Amsel 1933: 125; Amsel 1935a: 299; Bodenheimer 1937: 101.

Material examined. Syntypes of *Tachyptilia karmeliella*: 3 ♂, 2 ♀, Ain Karim, Jerusalem, Z. 333, 18.v.1930, *Crataegus* (Amsel) (gen. slide 32/18♂; 33/18♀, O. Bidzilya; 1331♂ Hendriksen); 1 ♀, Karmel, Haifa, 7.v.1930 (Amsel) (GU 657♂) (all SMNK).

Remarks. *Tachyptilia karmeliella* was described from an unstated number of specimens (“Karmel, 7.5., Ain Karim, 18. 5, ex l. *Crataegus*”) from Ain Karim, Jerusalem and Karmel mountain near Haifa. A holotype was not selected in the original description and the five specimens in SMNK labelled as “Typus” and “Paratypus” should be considered as syntypes.

Here we provide the first description of the genitalia of both sexes.

Male genitalia (Fig. 10). Uncus subovate, posterior margin strongly sclerotized, covered with long setae; gnathos broad, hook-shaped; tegumen moderately broad, with short anteromedial emargination; valva nearly straight, weakly narrowed at base, apex rounded extending to the top of uncus; posterior margin of vinculum with triangular projections that extend nearly to 2/3 length of valva; saccus long, narrow, triangular; phallus stout, weakly curved, equalling combined length of sacculus and valva, apex weakly pointed, caecum weakly swollen, half length of phallus.

Female genitalia (Fig. 11). Papillae anales broad, subtriangular; segment VIII broader than long, weakly sclerotized along anterior margin and at the base of apophyses anteriores, anterior margin straight, ostium small, funnel-shaped, posterior projection large, subrectangular, 2.5 times as long as broad, rounded posteriorly, about ¼ width of segment VIII, extending the posterior margin of segment VIII; apophyses posteriores slightly longer than apophyses anteriores; ductus bursae narrow, as long as the apophyses anteriores; corpus bursae subovate, longer than ductus bursae, densely covered with short spines; signum a narrow, weakly sclerotized plate, anterior margin gradually broadened, posterior margin straight, covered with triangular teeth.

Host plant. The type series was bred from larvae found feeding on *Crataegus* sp. (Amsel 1935a: 299).

Distribution. Israel.

Remarks. *Anacamptis karmeliella* resembles externally (Figs 8–9) *A. obscurella* (Denis & Schiffermüller, 1775) but the forewing is more unicolorous, and the black subapical fascia is narrow and less distinct. The weakly curved phallus in combination with the long and narrow valva are characteristic for the male genitalia. The female genitalia resembles those of *A. obscurella*, but differ in the longer posterior projection and shorter ductus bursae. *Anacamptis malella* Amsel, 1959 from Iraq is externally similar to *A. karmeliella*, but differs in details of the male genitalia, especially by having a distinctly longer phallus.

Anacamptis timidella (Wocke, 1887)

Tachyptilia timidella Wocke, 1887—Bresl. ent. Z. 12: 63.

Tachyptilia disquei Meess, 1907—Mitt. Badische Zool. Ver. 18: 125.

Tachyptilia quercella Chrétien, 1907—Le Naturaliste 29 (496): 250.

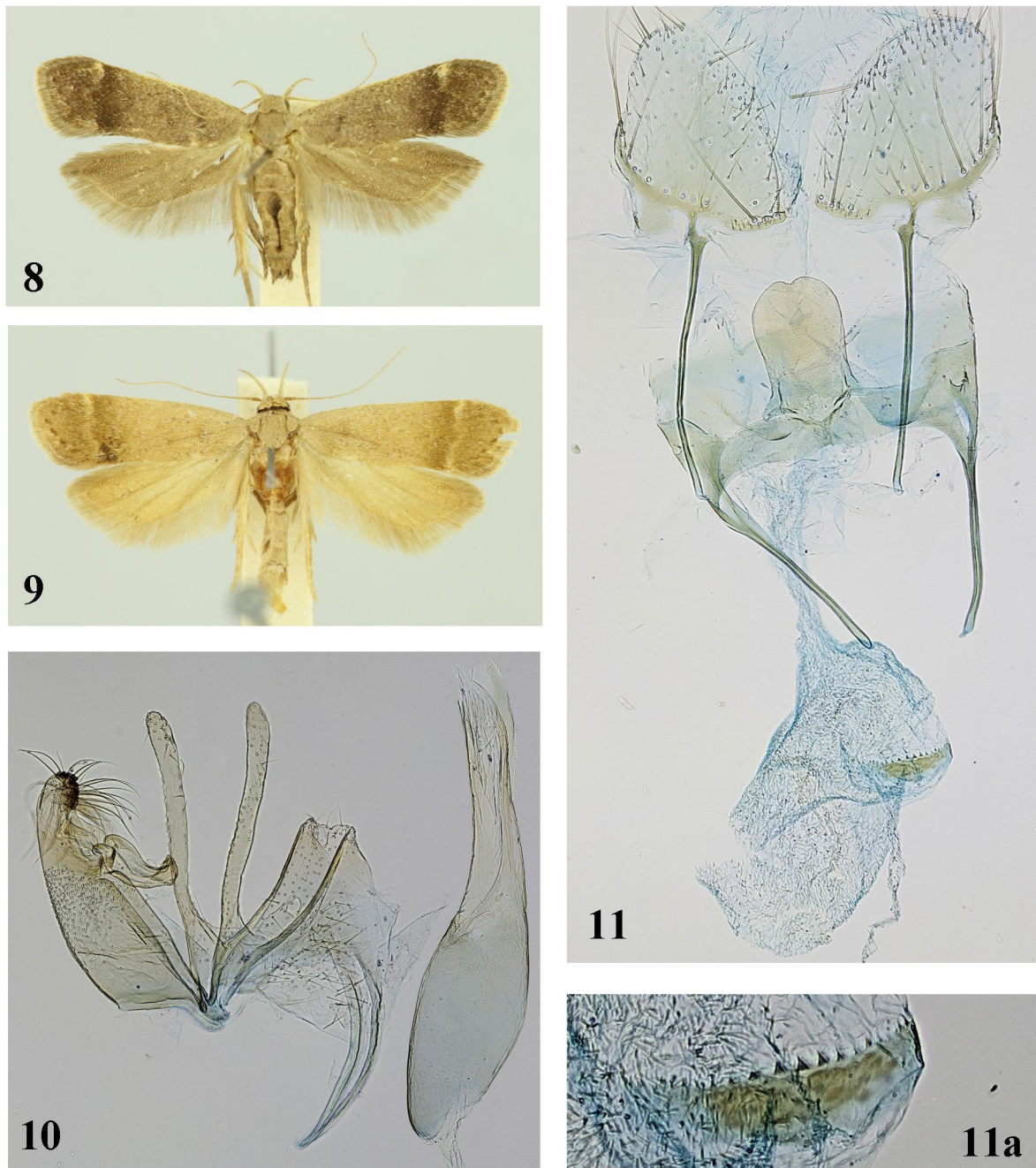
Tachyptilia suberiella Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 105.

Records. Amsel 1933: 125 (*quercella*); Amsel 1935c: 264 (*quercella*); Bodenheimer 1937: 101 (*quercella*); Bytinski-Salz & Sternlicht 1967: 130 (*quercella*); Halperin 1986: 125, 129, 131 (*quercella*).

Material examined. 3 ♂, 1 ♀, Waldheim, Haifa, 9.v.1930 (Amsel) (SMNK); 2 ex, 2 ♂, 3 ♀, Jogneam, pupa 6, 13, 24, 29.iv.1974, *Q. ithaburensis*, em. 6, 7, 10, 12, 19, 27.v.1974 (Halperin) (TAU).

Host plant. In Israel the larvae feed on *Quercus ithaburensis* Decne. (Amsel 1935c: 264; Bytinski-Salz & Sternlicht 1967: 130; Halperin 1986: 125, 129,131) and *Q. boissieri* (now *Quercus infectoria* subsp. *veneris* (AS. Kern.) Meikle) (Halperin & Sauter 1992: 134).

Distribution. Europe (except for Scandinavia) eastwards to Ural Mountains, Turkey, Israel (Karsholt & Riedl 1996: 118; Junnilainen *et al.* 2010: 61), Khabarovskiy krai of Russia (Ponomarenko 2008: 102).



FIGS 8–11. *Anacamptis karmeliella* Ams. 8. Adult, syntype, ♂. 9. Adult, syntype, ♀. 10. Male genitalia (gen. slide 32/18, O. Bidzilya). 11. Female genitalia (gen. slide 33/18, O. Bidzilya). 11a. Signum (enlarged).

Stomopteryx remissella (Zeller, 1847)

Gelechia remissella Zeller, 1847—*Isis von Oken* 1847 (11): 854.

Anacampsis vetustella Herrich-Schäffer, 1855—Syst. Bearb. Schmett. Europ. 5 (65): 191, (58) (ii), fig. 526.
Gelechia submissella Frey, 1880—Lep. Schweiz: 367 (preocc. by *Gelechia submissella* Stainton, 1859).
Xystophora bicolorella Rebel, 1903—Ann. nat. Mus. Wien 18: 329, pl. 3, fig. 14.
Aristotelia remissella rufobasella Rebel, 1916—Verh. zool.-bot. Ges. Wien 66: 10.
Stomopteryx yunusemrei Koçak, 1986—Priamus 4 (1–2): 58 (repl. for *Gelechia submissella* Frey, 1880).

Records. Amsel 1933: 125; Amsel 1935c: 264; Bodenheimer 1937: 101.

Material examined. 1 ♂, Karmel, Haifa, 7.v.1930 (Amsel); 1 ♂, Ain Karem, Jerusalem, 20.vi.1930 (Einsler) (all SMNK); 1 ♀, Michmoret, 26.viii.1968 (TAU); 2 ♂, slopes by Yflach, 31.v.2004 (Kravchenko); 1 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (TAU, ZMUC); 2 ♂, Nahal Oren, Mt. Carmel, light trap, 19.vi.1998 (Pavliček & Kravchenko) (gen. prep. 19650, J. Šumpich); 1 ♂, ditto but 21.viii.1998 (all NMPC).

Additional material. 1 ♂, Russia, Altai Republic, Belyashi (Dzhazator) env. (25 km NW), confluence of Argut and Karagem rivers, 49°51'56"N, 87°10'22"E, rocky steppe, 1400 m, 27.-28.vii.2017 (Šumpich) (DNA Barcode NMPC-Lep-0393) (NMPC).

Distribution. Europe to South Ural (Junnilainen *et al.* 2010: 55), Altai Mts. (new record), Lebanon (Zerny 1934: 20), Israel.

Stomopteryx tesserapunctella (Amsel, 1935) comb. nov.

Gelechia tesserapunctella Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 300, pl. 11, fig. 85.

Records. Amsel 1933: 125; Amsel 1935a: 300; Amsel 1935c: 263; Bodenheimer 1937: 100.

Material examined. Syntype of *Gelechia tesserapunctella* ♂, “Waldheim, Haifa, 9.v.1930, H. Amsel | Typus, ♂, *Gelechia tesserapunctella*“ GU.710 (SMNK); 1 ♂, Kefar Shemal, 22.viii.1968 (TAU); 14 ♂, slopes by Yflach, 31.v.2004 (Kravchenko) (TAU, ZMUC); 3 ♂, 1 ♀, Golan Heights, Masd al Shams, oak forest, 26.viii.2000 (Pavliček) (gen. slide 19028, J. Šumpich); 1 ♀, Nahal Orn, Mt. Karmel, 29.x.1998 (Pavliček & Kravchenko) (gen. slide 19019, J. Šumpich) (NMPC).

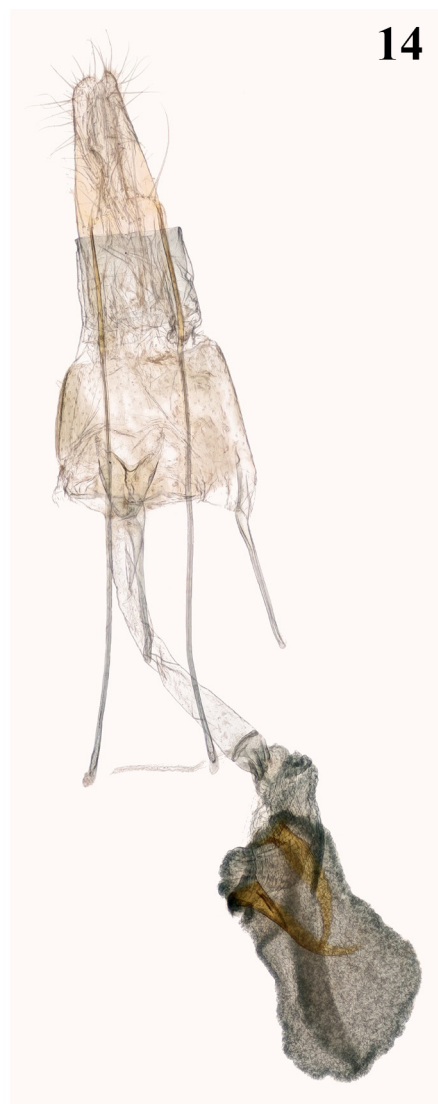
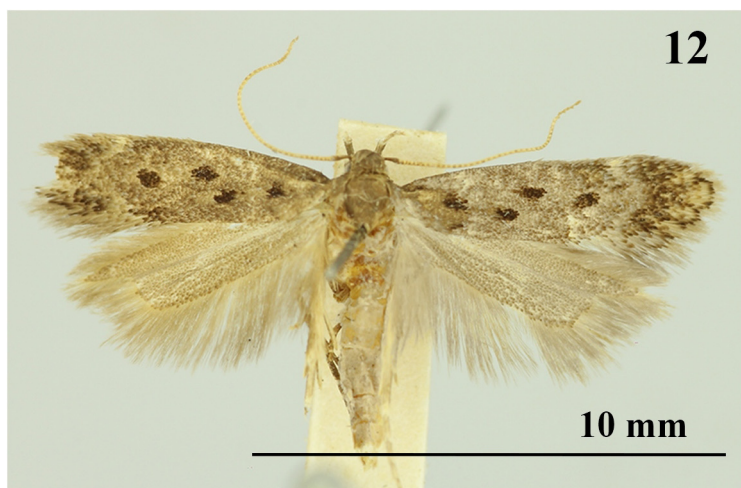
Remarks. *Gelechia tesserapunctella* was described from an unstated number of specimens collected 9.v.1930 in Waldheim near Haifa. A holotype was not selected in the original description. We examined a single male in SMNK (see label above) which should be considered as a syntype (Fig. 12). This specimen bears a label “GU. 710”, but the abdomen is intact. Here we provide the first description of the genitalia of both sexes.

Male genitalia (Fig. 13). Uncus elongate, lateral folds densely covered with long hair-like setae extending anteriorly to middle of tegumen; gnathos narrow, ring-shaped; tegumen subovate; valva gradually curved, broadest in the middle, apex rounded, extending to mid-length of the uncus; vinculum with a pair of rounded lobes surrounding the phallus; phallus swollen in basal third, distal portion narrow, straight, pointed apically, ventral process of even width, stout, separated from the phallus by a wide gap, broader than distal half of the phallus, apex with pointed hook.

Female genitalia (Fig. 14). Papillae anales elongated, subtriangular; apophyses posteriores three times as long as the length of apophyses anteriores, extending anteriorly to the entrance of ductus bursae; segment VIII as long as broad, evenly sclerotized, antrum funnel-shaped, posterior margin with V-shaped emargination; apophyses anteriores subequal in length with segment VIII; ductus bursae moderately broad, of even width; bursae copulatrix slightly shorter than ductus bursae, elongated, with a pair of narrow, weakly curved, pointed apically hook-shaped signa extending to middle of corpus bursae.

Remarks. *Stomopteryx tesserapunctella* slightly resembles externally *S. nugatricella* Rebel, 1893 but it has broader forewings, with darker light brown, but less distinct markings, the labial palpi are grey rather than white and antennae are light brown (blackish grey in *S. nugatricella*). A stout ventral process of equal width is the most characteristic feature of male genitalia for separating *S. tesserapunctella* from other *Stomopteryx* species. Two long hook-shaped signa in combination with the funnel-shaped antrum are characteristic for the female genitalia.

Distribution. Israel.



FIGS 12–14. *Stomopteryx tesserapunctella* Ams. 12. Adult, syntype, ♂. 13. Male genitalia (gen. slide 19028, J. Šumpich). 14. Female genitalia (gen. slide 19019, J.Šumpich).

Stomopteryx basalis (Staudinger, 1876)

Anacamptis (?) *basalis* Staudinger, 1876—Stettin. ent. Ztg 37: 147.

Material examined. 1 ♂, Ein Gedi, 17.viii.1968 (TAU).

Distribution. France, Italy (including Sicilia), Malta (Karsholt & Riedl 1996: 119), Spain (Requena 2009: 50), Croatia (Šumpich & Skyva 2012: 173), Greece (Karsholt 2011), Israel (new record).

Stomopteryx lacteolella Turati, 1924

Stomopteryx lacteolella Turati, 1924—Atti Mus. Civ. Milano 63: 166.

Stomopteryx lacteolella f. *unipunctella* Turati, 1924—Atti Mus. Civ. Milano 63: 166, pl. 6, fig. 10.

Material examined. 3 ♂, Negev, Nahal (Wadi) Zin, 8.x.1999 (Pavliček) (gen. slide 19007, J. Šumpich) (NMPC).

Diagnosis. *Stomopteryx lacteolella* can easily be recognized externally by its light, yellowish brown forewing with a brown dot in the cell corner and dark brown apex. The male genitalia are rather similar to those of *S. deter-*

sella (Zeller, 1847), but *S. lacteolella* has broader valvae bulging anteriorly whereas they are boomerang-shaped in *S. detersella*. Vinculum surrounding phallus has a triangular sclerotization in *S. detersella* but it is straight thorn-shaped in *S. lacteolella*. Moreover, the phallus is more robust, with shorter ventral process and larger sclerotized apical hook in *S. lacteolella*.

Distribution. Libya, Israel (new record).

Remarks. *Stomopteryx lacteolella* was described from one specimen from Benghazi (“Berca”) in Libya. Turati (1924: 166) wrote (here translated from Italian): “Of this species recently described by Dumont was collected only one exemplar...”. However, no description of a *lacteolella* Dumont seems to exist, and the name should therefore be ascribed to Turati. Moreover he was of the opinion that his specimen was a form of (the undescribed) *S. lacteolella* Dumont. As Turati had only one specimen the name *unipunctella* becomes an objective synonym of *S. lacteolella*. This species seems to be known only from the holotype. It has not been dealt with in the literature since its description, apart from a few catalogues and websites. The specimen from Israel (Fig. 15) matches the colour figure of the holotype well.

Here we describe the male genitalia for the first time.

Male genitalia (Fig. 16). Uncus subtriangular, slightly longer than broad, lateral folds densely covered with long hair-like setae, extending anteriorly to middle of tegumen; gnathos narrow, ring-shaped; tegumen narrow, parallel-sided to 2/3 length, then weakly narrowed, with indistinct transition to uncus; valva narrow at base, ventral margin distinctly broadened in middle, apex rounded, extending nearly to mid length of the uncus; vinculum with a pair of rounded lobes surrounding the phallus; phallus swollen in basal third, distal portion narrow, weakly curved, pointed apically, ventral process stout, separated from the phallus by wide gap, subtriangular, strongly narrowed towards distinct apical hook.

Approaerema polychromella (Rebel, 1902)

Anacampsis polychromella Rebel, 1902—Dt. ent. Z. Iris 15: 109.

Anacampsis faceta Meyrick, 1914—Ann. Transv. Mus. 4 (4): 192.

Anacampsis argyrolobiella Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 10.

Anacampsis polychromella var. *rebeliella* Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 109.

Records. Bodenheimer 1930a: 174; Amsel 1933: 125, 126; Amsel 1935b: 210; Amsel 1935c: 264; Bodenheimer 1937: 101.

Material examined. 2 ex, Tel-Aviv (Dune), 17.ix, 21.x.1930 (Einsler); 2 ex., Südende d. Toten Meers, 15-27.iii.1933 (SMNK); 6 ♂, 1 ♀, Huliioth, 27.iii.1968 (Shoham); 13 ex., Hazeva, Field School, Hazeva, 16.iii, 20, 16. iv, 4, 9, 11, 18.v.1998 (Ashkenazi) Malaise Trap; 1 ♀, Nahal Hidron, 25km W Arava Hwy (90) 9.iii.1988 (Ashkenasi); 1 ex, same data but 5 km W, 12.iii.1998; 2 ex, Shezaf N.R., Nahal Shahaq, 28.iv, 1.vi.1999 (Yarom) Malaise Trap (all TAU); 11 exx, Jericho, Kalia, v.2004 (Kravchenko); 1 ♀ slopes by Yflach, 31.v.2004 (Kravchenko); 2 exx, Jerusalem, 6.vi.2004 (Kravchenko) (all TAU, ZMUC); 9 ♂, Nahal Oren, Mt. Carmel, light trap 17.v.1997 (Pavliček & Kravchenko); 1 ♂, ditto but 21.v.1998 (all NMPC).

Distribution. Spain (with Canary Islands), Portugal (with Madeira), Gibraltar, Italy (with Sicily), Austria, Malta, Croatia, Macedonia, Ukraine, Greece, Crete, Cyprus, Middle East, Egypt, Sudan, Arabia, Turkey, Iraq, Armenia, Uzbekistan, Turkmenistan, Mongolia, India, Yemen, Kuwait, Cameroon, Sudan, Namibia, South Africa. Recorded as a migrant to Denmark, England and Czech Republic (Amsel 1961; Ivinskis & Piskunovas 1994: 40; Karsholt & Riedl 1996: 119; Vávra *et al.* 2008: 89; Falkovitsh & Bidzilya 2009: 82; Bidzilya & Budashkin 2017: 16; De Prins & De Prins 2018).

Approaerema anthyllidella (Hübner, [1813])

Tinea anthyllidella Hübner, [1813]—Samml. eur. Schmett. [8]: pl. 48, fig. 330.

Gelechia nigritella Stainton, 1854—Ins. Britannica: 133.

Gelechia psoralella Millière, 1865—Iconogr. Descr. Chenilles Lépid. inédits 2: 83, 99, pl. 61.

Gelechia (Anacampsis) lachtensis Erschoff, [1877]—Horae Soc. ent. Ross. 12 (4): 345.

Gelechia (Anacampsis) sparsiciliella Barrett, 1891—Ent. mon. Mag. 27: 7.

Anacampsis infestella Rebel, 1896—Ann. Mus. Wien 11: 128.
Monochroa brundini (Benander, 1945)—Ent. Tidskr. 66: 132.
Aproaerema alfalfella Amsel, 1958—Beitr. naturk. Forsch. Südwdtschl. 17 (1): 80.
Aproaerema aureliana Capuse, 1964—Ent. Tidskr. 85: 15.

Records. Amsel 1933: 125; Amsel 1935c: 264; Bodenheimer 1937: 101.

Material examined. 1 ex., Jericho, 11.iv.1930, Lichtfang; 1 ex., Georgskloster, Wadi et Kelt, Lichtfang, 1.iv.1930; 1 ex., On alfalfa, Rehovot, 16.ii.1936 (SMNK); 1 ♀, 5.iii.1975, Huliioth, 21.vi.1968; 1 ex., Kyriat Haim, Lotus creticus, 3.iv.1975, coll. J. Halperin / *Stomopteryx anthyllidella alfalfella* Ams., ♂, GP 7314, W. Sauter; 1 ♂, Bat-Shlomo, 20.viii.1968 (gen. slide 131/18, O. Bidzilya) (all TAU); 1 ♂, slopes by Yflach, 31.v.2004 (Kravchenko); 1 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (all TAU); 3 ♂, East bank of Kenneret lake, Mevo Hama, 18.iv.2018, light (Kravchenko & Bidzilya) (TAU, ZMKU); 1 ♂, 2 ♀, Keshet, 21.iv.2018, at light (Kravchenko & Bidzilya) (ZMKU).

Host plant. In Israel the larvae have been recorded feeding on *Trifolium* sp. (Bodenheimer 1937: 101), *Medicago sativa* L. and *Lotus creticus* L. (according to label).

Distribution. Palaearctic region.

Aproaerema languidella (Amsel, 1936) comb. nov.

Schützeia languidella Amsel, 1936 – Veröff. dt. Kol. Übersee-Mus. 1: 366, pl. 15, fig. 22.

Records. Amsel 1936: 366; Bodenheimer 1937: 101

Material examined. Syntypes of *Schützeia languidella* ♂, "Tel-Aviv (Düne), 25.v.1932, W. Einsler, H. Amsel | Typus, leg. H. Amsel, *Schützeia languidella*" (gen. slide 20/18, O. Bidzilya). 1 ex (abdomen missing), same data, but "Paratypus" (all SMNK).

Distribution. Israel.

Remarks. *Schützeia languidella* was described based on three males collected in the vicinity of Tel Aviv. We traced two specimens from type series in SMNK, which are considered as syntypes. The species is closely related to *A. anthyllidella* in the male genitalia, but adult differs by distinct nearly connected white tornal and costal spot at the basal $\frac{3}{4}$ on the forewing (Fig. 39). *Aproaerema languidella* looks very similar externally to the next species (Figs 17, 18) too, except for more diffuse subapical fascia. Moreover, three males collected sympatrically with syntypes of *A. languidella* are identical externally to *A. telaviviella*. Hence, both species are most likely conspecific. See also notes for the following species.

Aproaerema telaviviella (Amsel, 1935) comb. nov.

Anacampsis telaviviella Amsel, 1935—Mitt. Zool. Mus. Berl. 20(2): 301, pl. 11, fig. 107.

Records. Amsel 1933: 125; Amsel 1935a: 301, pl. 11, fig. 107; Amsel 1935c: 264; Amsel 1936: 366, pl. 15, fig. 23; Bodenheimer 1937: 101.

Material examined. Syntypes of *Anacampsis telaviviella* 1 ♀, "Tel-Aviv (Düne), 19.v.1930 H. G. Amsel" | "GU. 713" | "Typus. leg. H. Amsel, *Anacampsis telaviviella*", 1 ♂ (abdomen missing), "Tel-Aviv, Lichtf.[ang], Düne, 15.v.30 (Pal), H. Amsel" | "Paratypus. leg. H. Amsel, *Anacampsis telaviviella*"; 3 ♂, Tel-Aviv (Düne), 25.v.1932 (Einsler) (gen. slide 29/18, O. Bidzilya); 1 ♂, same place but 21.x.1932 (all SMNK).

Distribution. Israel.

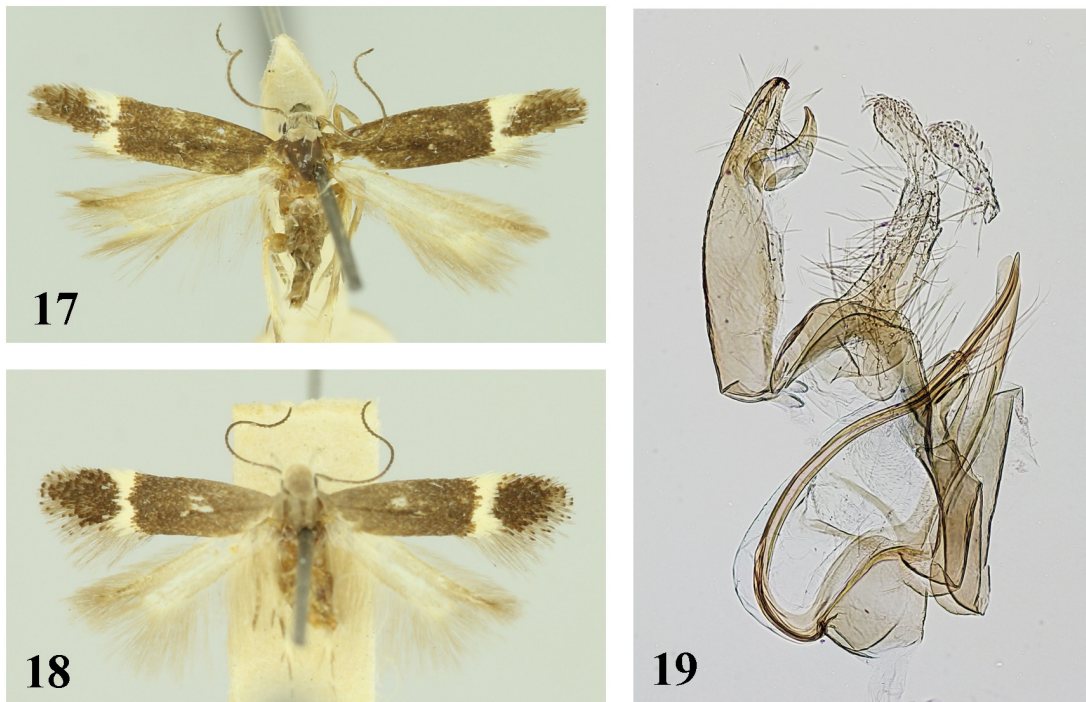
Remarks. *Anacampsis telaviviella* was described from an unstated number of specimens collected 19.v.1930 in Tel-Aviv ("Tel-Aviv, Düne"). A holotype was not selected in the original description, and two specimens in SMNK should be considered as syntypes. There are three additional males in SMNK collected sympatrically with syntypes of *A. languidella* 25.v.1932, of which one (Fig. 17) looks similar to the one of syntype (Fig. 18) and also bears a type label, but it does not belong to the type series.

The genitalia slide of the female syntype is uninformative, because the antrum is not visible and both ductus

bursae and corpus bursae are missing. The male genitalia of a specimen externally similar to *A. telaviviella* (Fig. 19), but collected 25.v.1932, look indistinguishable from those of *A. anthyllidella* and *A. languidella*. The status of this and the previous species remains unclear and can hardly be clarified without studying morphology and DNA of additional material.



FIGS 15, 16. *Stomopteryx lacteolella* Turati. 15. Adult. 16. Male genitalia (gen. slide 19007, J. Šumpich).



FIGS 17–19. *Aproaerema telaviviella* Ams. 17. Adult, ♂. 18. Adult, syntype, ♂. 19. Male genitalia (gen. slide 29/18, O. Bidzilya).

Tribe Chelariini Le Marchand, 1947

Nothris gregerseni Karsholt & Šumpich, 2015

Nothris gregerseni Karsholt & Šumpich, 2015—*Zootaxa* 4059 (3): 481, figs 11–13, 52–61, 110, 121–128.

Material examined. 5 ♂, 1 ♀, Golan Heights, Katzrin, 12.ix.2003 (gen. slide 119/18♀; 120/18♂, O. Bidzilya) (Kravchenko); 1 ♂, Shaues, vii.1965 (TAU).

Distribution. Denmark, Sweden, Turkey, Russia: Ural, Altai Mts, Southern Siberia (Karsholt & Šumpich 2015: 483), Israel (new record).

Nothris sabulosella Rebel, 1935

Nothris sabulosella Rebel, 1935—Mitt. Münch. Ent. Ges. 25: 41.

Material examined. 2 ♂, Golan Heights, Katzrin, 12.ix.2003 (Kravchenko) (gen. slide 121/18, O. Bidzilya) (TAU).

Distribution. Turkey, Israel (new record).

Nothris congressariella (Bruand, 1858)

Ypsolopha congressariella Bruand, 1858—Ann. Soc. ent. Fr. (3) 6: 471, pl. 11, fig. 7.

Records. Bodenheimer 1937: 102 (*deklaratella*).

Material examined. 1 ♀, Jerusalem, Palästina, 18.iv.1930 (Amsel) (SMNK).

Distribution. From the Canary Islands, Tunisia, SW Europe, Greece, Macedonia to Turkey; northwards to south-western part of the British Isles (Karsholt & Šumpich 2015: 474).

Nothris verbascella ([Denis & Schiffermüller], 1775)

Tinea verbascella Denis & Schiffermüller, 1775—Ank. syst. Schmett. Wienergegend: 136.

Nothris verbascella clarella Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 298.

Records. Bodenheimer 1930a: 174 (*sulcella*; misidentification); Amsel 1935a: 298; Bodenheimer 1937: 101 (*verbascella clarella* & *sulcella*); Karsholt & Šumpich 2015: 484.

Material examined. Syntypes of *Nothris verbascella clarella*: 1 ♂, Karmel, Haifa, 7.v.1930 (Amsel); 1 ♂, Jericho (Pal.), Lichtfang, 28.iv.1930 (Amsel) (all SMNK); 1 ♂, Jericho, 28.iv.1930 (Amsel); 1 ♂, Tel Aviv, 18.iv.1925 (Bodenheimer) (all NHMW).

Host plant. In Israel the larvae have been recorded feeding on *Verbascum* sp. (Bodenheimer 1937: 101).

Distribution. Europe, northwards to central Scandinavia, European Russia, North Africa, Middle East, Turkey, Iran, Iraq, Turkmenistan (Karsholt & Šumpich 2015: 486).

Anarsia arachniota Meyrick, 1925

Anarsia arachniota Meyrick, 1925—Bull. Soc. ent. Egypte 9 (1–3): 210.

Records. Amsel 1933: 126; Amsel 1935c: 265; Bodenheimer 1937: 102.

Material. 2 ♂, Jericho (Pal), Lichtfang, 25.ix.1930 (Amsel) (gen. slide GU3114, 1302 Hendriksen) (SMNK).

Distribution. Egypt, Israel.

Anarsia acaciae Walsingham, 1896

Anarsia acaciae Walsingham, 1896—Proc. zool. Soc. Lond. 1896: 278.

Records. Buxton 1924: 334; Bodenheimer 1930a: 174; Bodenheimer 1930b: 280; Amsel 1933: 126; Bodenheimer 1937: 102; Halperin 1986: 128, 131.

Material examined. 4 ♂, 5 ♀, Shezaf N.R., Nahal Shahaq, 27.viii, 5.x.1999 (Yarom) Malaise Trap; 1 ♀, Nahal Gidron, , 21.x.1999, Malaise Trap (Yarom & Zeevi); 1 ♀, Nahal Shahak, Shesaf Nat. Rez., 11.vi.1997 (Yarom); 3 ♀, Ein Gedi, 2.viii.1971, Dead Sea valley, *Acacia raddiana*, 17.viii, 1.ix.1971 (Halperin); 1 ♀, Ein Gedi, 17.ix.1968

(all TAU); 2 ♂, 1 ♀, Nahal Oren, Mt. Carmel, light trap, 29.ix.1998 (Pavliček & Kravchenko) (gen. prep. 19644, J. Šumpich); 1 ♀, ditto but 5.x.1997 (gen. prep. 19645, J. Šumpich); 1 ♀, ditto but 19.vi.1998 (all NMPC).

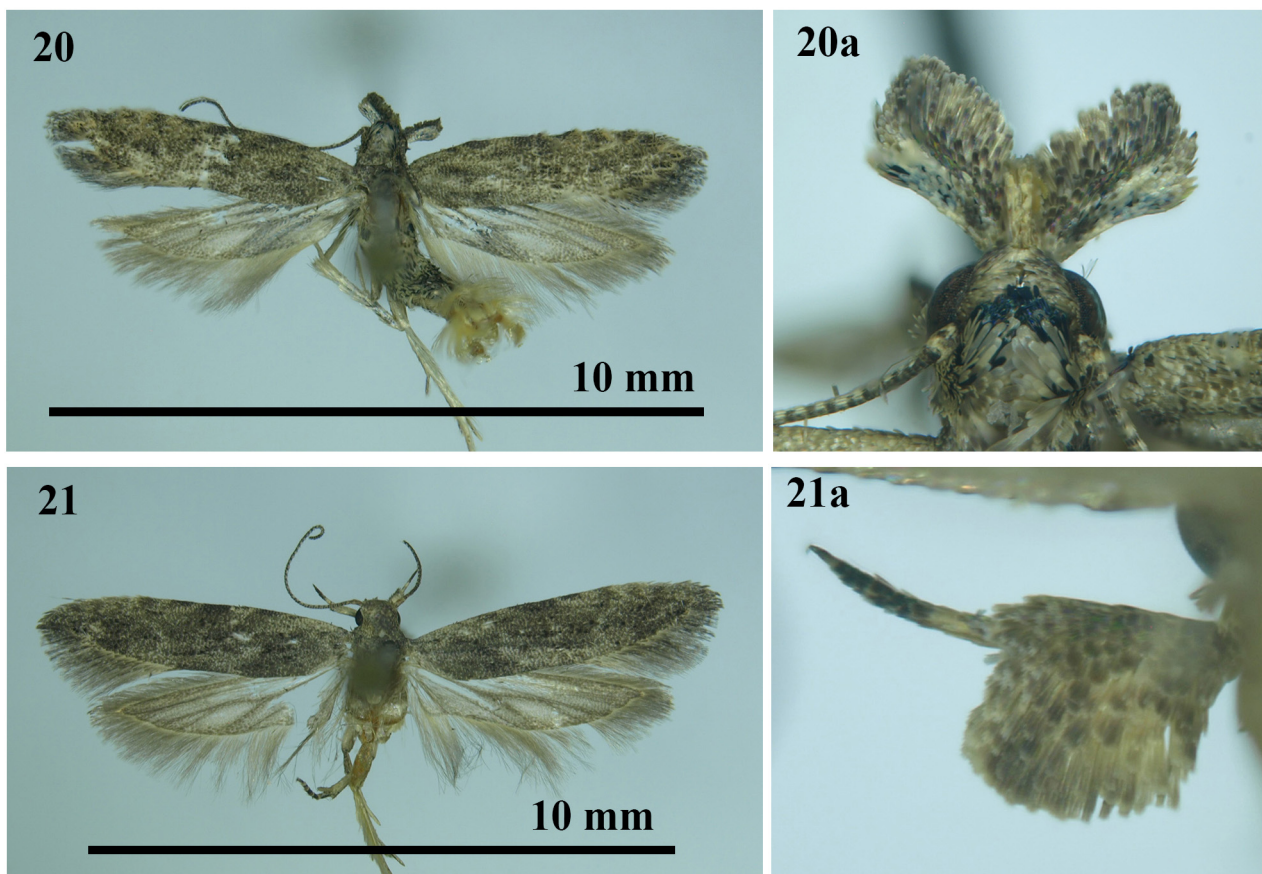
Host plant. In Israel larvae feed on *Acacia farnesiana* (L.) Wild. (Buxton 1924: 334; Bodenheimer 1930a: 174; 1930b: 280) and in pods of *A. tortilis* spp. *raddiana* (Savi) Brenan (Halperin 1986: 128, 131; Halperin & Sauter 1992: 134).

Distribution. Algeria, Yemen, Kenya (Bidzilya & Agassiz 2016: 118), Israel.

Anarsia balioneura Meyrick, 1921

Figs 20, 21.

Anarsia balioneura Meyrick, 1921—Ann. Transv. Mus. 8 (2): 79.



FIGS 20–21. *Anarsia balioneura* Meyr. 20. Adult, ♂. 20a. ♂, head and labial palpus, dorsal view. 21. Adult, ♀. 21a. ♀, labial palpus, lateral view.

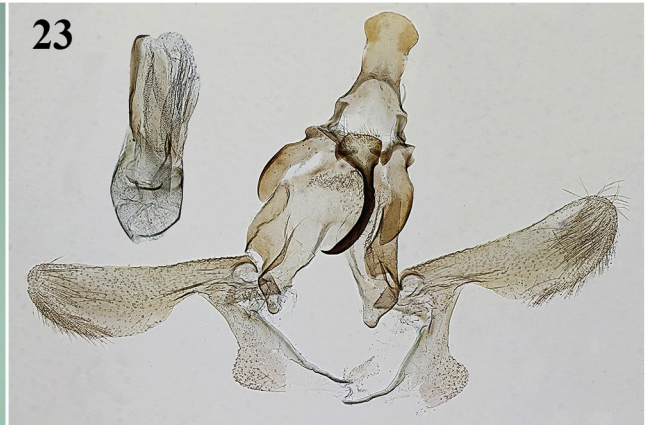
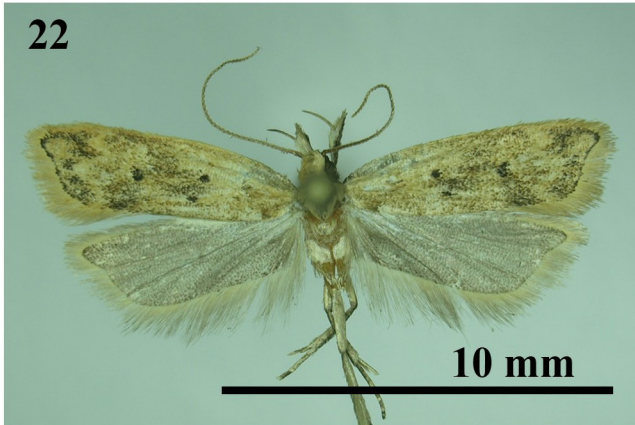
Material examined. 1 ♂, 4 ♀, Tel Aviv, Botanical garden, 14, 15.iv.2018, light (Kravchenko & Bidzilya) (TAU, ZMKU).

Additional material. 1 ♂, Cyprus (North), Akdeniz Plaji, 10 m, 1-3.iv.2008 (Skule & Hviid); 1 ♂, 1 ♀, Libya, Gharian, Wadi el Hira, 1.vi.1983 (Seneca) (all ZMUC).

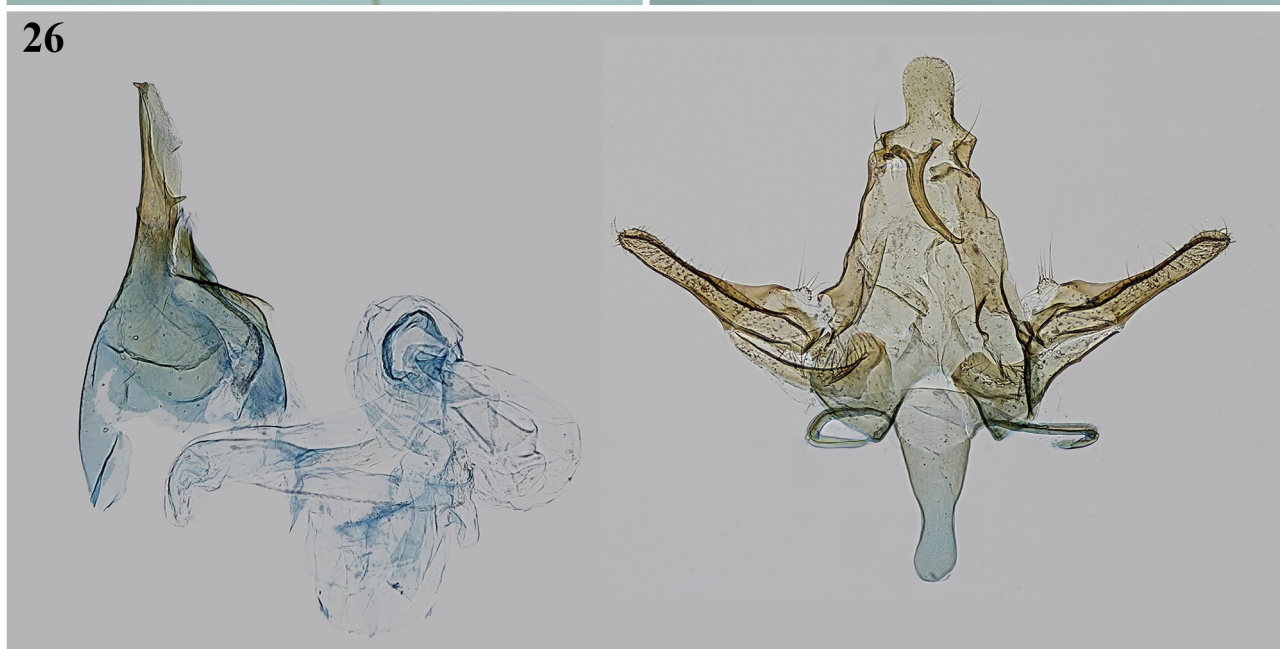
Host plant. The larva feeds on six *Acacia*-species in Kenya (Agassiz & Bidzilya 2016: 120). The specimens collected in the Botanical garden in Tel Aviv most likely live on one of the *Acacia*-species growing there.

Distribution. Zimbabwe, Malawi, Uganda, Kenya, Cape Verde (Agassiz & Bidzilya 2016: 120), Cyprus, Israel, Libya (new records).

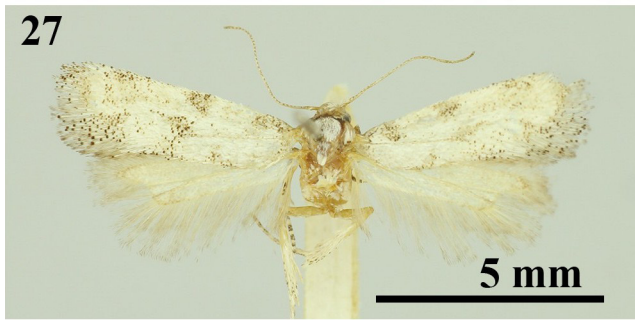
Remarks. The specimens from Israel match well both superficially and in genitalia specimens of *A. balioneura* from Africa.



FIGS 22–23. *Acompsia ballotellus* Ams. 22. Adult, ♂ (gen. slide 152/18, O. Bidzilya). 23. Male genitalia (gen. slide 152/18, O. Bidzilya).



FIGS 24–26. *Polyhymno* spp. 24. *P. chionarcha* Meyr., ♂. 25. *P. dumonti* Hartig, adult, ♂ (gen. slide 123/18, O. Bidzilya). 26. *P. dumonti* Hartig, male genitalia (gen. slide 123/18, O. Bidzilya).



FIGS 27–29. *Chrysoesthia amseli* Bidz. 27. Holotype, ♀. 28. Adult, ♂ (gen. slide 115/18, O. Bidzilya). 29. Male genitalia (gen. slide 115/18, O. Bidzilya).

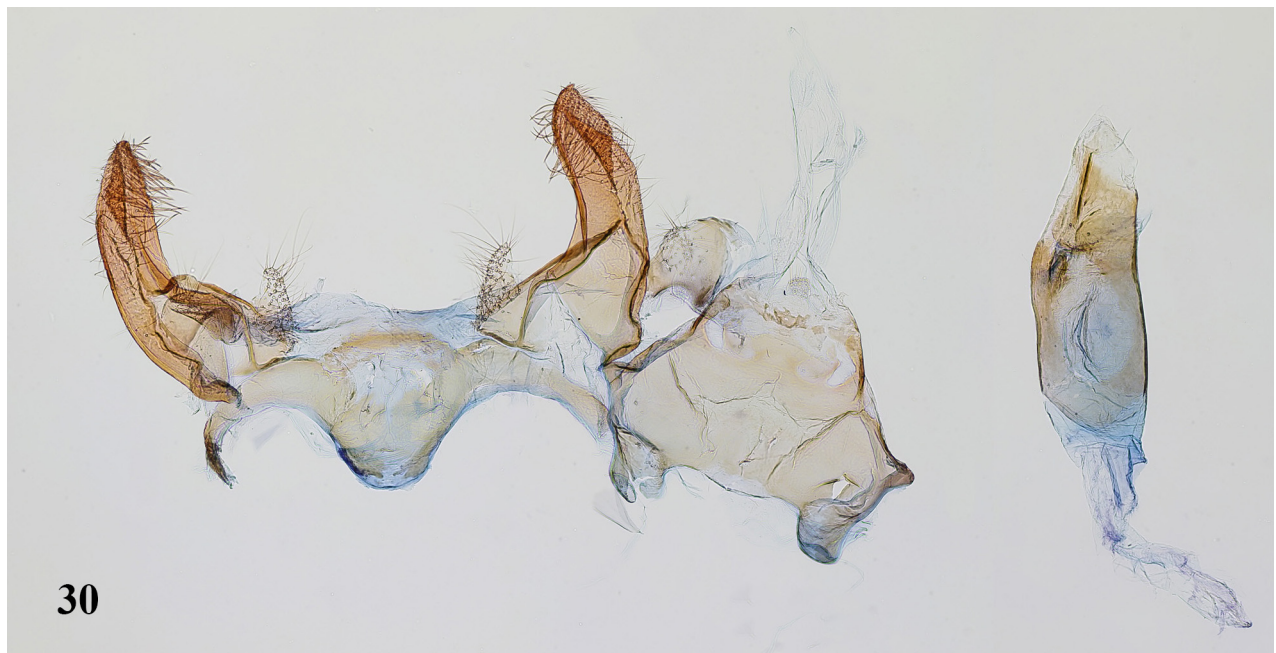


FIG. 30. *Oxypteryx jordanella* Rbl. Male genitalia (gen. slide 84/19, O. Bidzilya).

Anarsia lineatella Zeller, 1839

Anarsia lineatella Zeller, 1839—Isis [32] 1839 (3): 190.

Tinea pullatella Hübner, 1796—Samml. eur. Schmett. [8]: pl. 17, fig. 118.

Anarsia ? *pruniella* Clemens, 1860—Proc. Acad. Nat. Sci. Philad. 12: 169.

Ananarsia lineatella heratella Amsel, 1967—Beitr. naturk. Forsch. SüdwDtl. 26 (3): 27.

Ananarsia lineatella tauricella Amsel, 1967—Beitr. naturk. Forsch. SüdwDtl. 26 (3): 20.

Records. Bodenheimer 1930a: 174; Bodenheimer, 1930b: 235–236; Amsel 1933: 126; Bodenheimer 1937: 102.

Material examined. 1 ♀, Yizre'el, 40 km SE Haifa, 2.x.1972, coll. Besky | P.f. -2-1376 (1550), ex/on young fruit, plant coll. on: 10.ix.1972; 1 ♂ (abdomen missing), Wadi Auja, 10 km N Jericho, 6.ix.1971, coll. O. Herling | P.f. -1-657-769, ex/on ripe fruit, plant coll. on: 8.viii.1971 (SMNK); 4 ♂, 5 ♀, Jerusalem, 6.vi.2004 (Kravchenko) (genitalia slides 3408, 3409, 3410, 3411, 3412, 3413, 3414 Gregersen, 5499 Hendriksen, 5249 Karsholt) (TAU, ZMUC); 1 ♂, 5 ♀, East bank of Kenneret lake, Mevo Hama, 18.iv.2018, light (Kravchenko & Bidzilya) (ZMKU).

Host plant. In Israel the larvae have been recorded feeding on *Prunus* sp. and *Pyrus* sp. (Bodenheimer 1930a: 174, 1937: 102). Bodenheimer (1930b: 235) gave a detailed description of its biology. The record of larva from *Prosopis* (Halperin & Sauter 1992: 121) should most likely be referred to *Anarsia acaciae*.

Distribution. Central and southern Europe, North Africa, eastwards through the Middle East and Turkey to Central Asia, India (Kashmir), China (Xinjiang, Shaanxi), Primorskiy krai of Russia, North Korea and Japan, Australia (Lvovsky & Piskunov 1999: 49; Park & Ponomarenko 2007: 182; Li 2002: 238; Beljaev & Ponomarenko 2005: 2), North America, Canada (introduced) (Gregersen & Karsholt 2017:78).

Subfamily Dichomeridinae Hampson, 1918

Dichomeris acuminatus (Staudinger, 1876)

Mesophleps (?) *acuminatus* Staudinger, 1876—Stettin. ent. Ztg 37: 148.
Hypsolophus ianthes Meyrick, 1887—Trans. ent. Soc. Lond. 1887 (3): 273.
Ypsolophus rusticus Walsingham, [1892]—Proc. Zool. Soc. Lond. 1891: 525.
Ypsolophus lotellus Constant, 1893—Ann. Soc. ent. Fr. 62: 398.
Ypsolophus ammodxanthus Meyrick, 1904—Proc. Linn. Soc. N.S.W. 29 (2): 430.
Ypsolophus ochrophanes Meyrick, 1907—J. Bombay nat. Hist. Soc. 17 (4): 981.
Ypsolophus sublotellus Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 114.

Records. Amsel 1933: 126; Amsel 1935c: 265 (*lotellus*); Bodenheimer 1937: 102 (*lotellus*).

Distribution. South Europe, North Africa, China (Guandun, Taiwan), Japan, India, Sri Lanka, South Africa, Namibia, Kenya, Comoros, Mauritius, Reunion, Seychelles, Madagascar, Australia, Hawaii Isl., USA (Florida) (Ponomarenko 2009: 253; De Prins & De Prins 2018).

Dichomeris bodenheimeri (Rebel, 1926)

Rhinosia bodenheimeri Rebel, 1926—Verh. zool.-bot. Ges. Wien 74/75 (s.b.): (203).

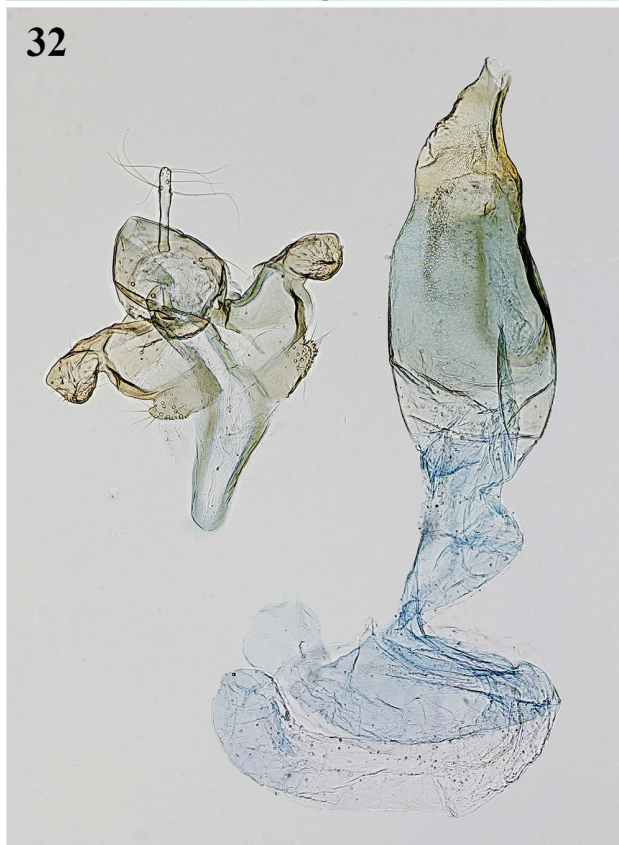
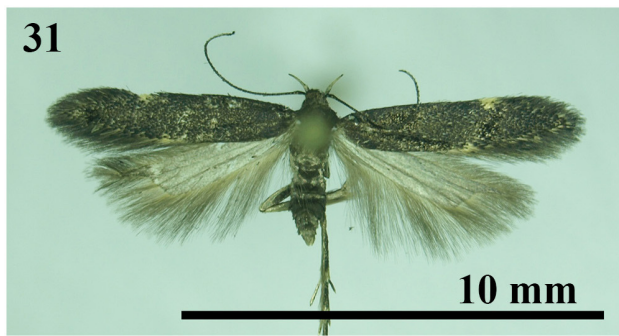
Records. Amsel 1933: 126; Amsel 1935b: 210; Amsel 1935c: 264; Bodenheimer 1937: 101.

Material examined. 1 ♂, 1 ♀, 1 ex, Südende d. Toten Meers, 15-27.iii.1933 (Amsel); 2 ♂, Jordanbrucke, Jericho, 10.iv.1930 (all SMNK); 1 ♂, Sinai, Wadi Hibran, 11.iv.1973 (Freidberg) (TAU); 1 ♂, 1 ♀, Negev, Nahal (Wadi) Zin, black light, 8.x.1999 (Pavliček) (NMPC).

Host plant. In Israel the larvae were recorded feeding on *Prosopis farcta* (Banks & Sol.) J. F. Macbr., (Rebel 1926: 204) and *Ononis* sp. (Bodenheimer 1937: 101), both Fabaceae.

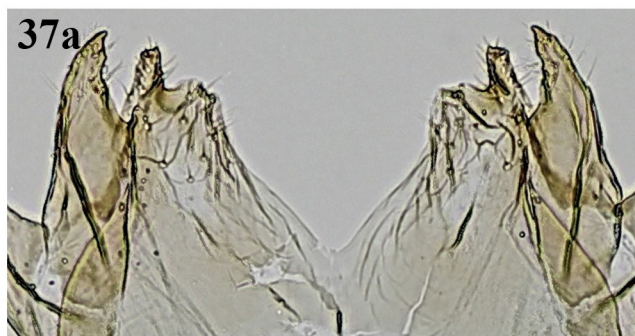
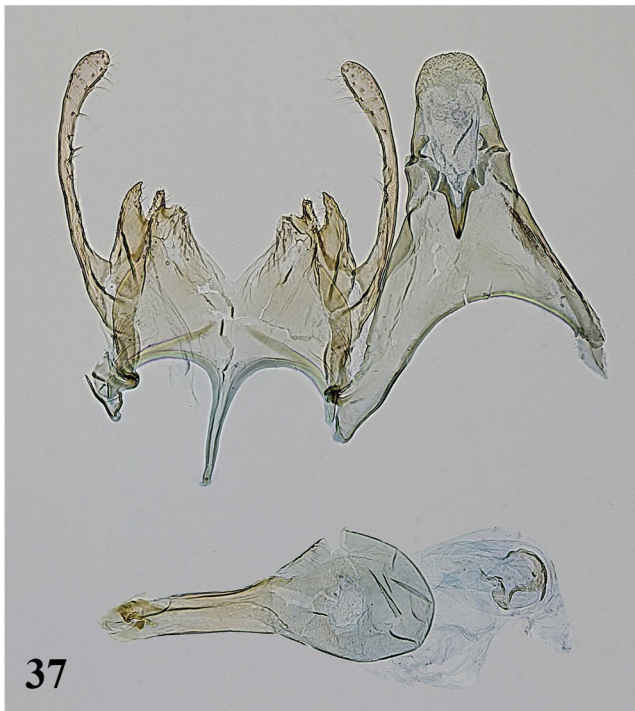
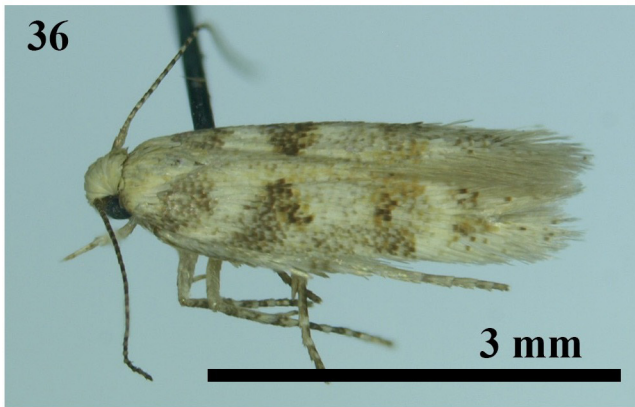
Distribution. Israel, Jordan, Saudia Arabia, Yemen, Kenya (Amsel 1956: 61; Amsel 1961: 59; Agassiz & Bidzilya 2016: 127).

Remarks. *Rhinosia bodenheimeri* was described from a male and a female bred (in May) from larvae found in December 1926, feeding from cases on *Prosopis stephaniana* (M. Bieb.) Kunth. ex Spreng. The type locality is given as “Palästina” without exact location.



FIGS 31–32. *Oxypteryx immaculatella* Dgl. 31. Adult, ♂. 32. Male genitalia (gen. slide 124/18, O. Bidzilya).

FIGS 33–35. *Metzneria freidbergi* sp. n. 33. Adult, holotype, ♂ (gen. slide 134/18, O. Bidzilya). 34. Male genitalia, paratype (gen. slide 137/18, O. Bidzilya), 34a. Phallus (enlarged). 35. Male genitalia, holotype (gen. slide 124/18, O. Bidzilya). 35a. Phallus (enlarged).



FIGS 36–38. *Scrobipalpa aravensis* sp. n. 36. Adult, Holotype, ♀ (gen. slide 136/18, O. Bidzilya). 37. Male genitalia, paratype (gen. slide 127/18, O. Bidzilya). 37a. Posterior margin of vinculum and sacculi (enlarged). 38. Female genitalia, holotype (gen. slide 136/18, O. Bidzilya).

***Dichomeris limbipunctellus* (Staudinger, 1859)**

Hypsolophus limbipunctellus Staudinger, 1859—Stettin. ent. Ztg 20 (7–9): 245.

Ypsolophus millierellus Stainton, 1873—Nat. Hist. Br. Tin. 13: 336.

Dichomeris neatodes Meyrick, 1923—Exot. Microlep. 3 (1–2): 35.

Material examined. 1 ♂, Bat-Shlomo, 25.viii.1968 (TAU); 2 ♂, 1 ex, Jerusalem, 6.vi.2004 (Kravchenko) (TAU); 1 ♂, Nahal Oren, Mt. Carmel, valley bottom, 29.ix.1998 (Pavliček & Kravchenko); 1 ♂, ditto but 21.viii.1998; 1 ♂, ditto but 5.x.1997; 1 ♂, ditto but 11.vii.1997 (all NMPC).

Distribution. Spain, South France, Italy, Greece, Cyprus, North Africa, and Israel (new record).

Dichomeris alacella (Zeller, 1839)

Gelechia alacella Zeller, 1839—*Isis* [32] 1839 (3): 199.

Records. Halperin & Sauter 1992: 134.

Material examined. 1 ♂, Nahal Oren, Mt. Carmel, light trap, 21.vi.1997 (Pavliček & Kravchenko); 1 ♂, ditto but 24.vi.1997; 1 ♀, ditto but 21.viii.1998 (gen. slide 19015, J. Šumpich) (all NMPC).

Host plant. In Israel the larvae have been recorded feeding on *Cercis siliquastrum* L. (Halperin & Sauter 1992: 134).

Distribution. Europe, Russia: European part and Western Siberia, Iran (Junnilainen *et al.* 2010: 63), Israel.

Brachmia blandella (Fabricius, 1798)

Tinea blandella Fabricius, 1798—*Suppl. ent. syst.* 499.

Gelechia gerronella Zeller, 1850—*Stettin. ent. Ztg.* 11 (5): 155.

Records. Stainton 1867: 40 (*gerronella*); Amsel 1933: 126 (*gerronella*); Bodenheimer 1937: 101 (*gerronella*).

Distribution. Europe to Ural Mountains, Turkey, Caucasus, Middle East (Karsholt & Riedl 1996: 121; Junnilainen *et al.* 2010: 64).

Helcystogramma lamprostoma (Zeller, 1847)

Gelechia lamprostoma Zeller, 1847—*Isis von Oken* 1847 (11): 851.

Anacampsis scutata Meyrick, 1894—*Trans. ent. Soc.* 1894 (1): 14.

Records. Bodenheimer 1930a: 174; Amsel 1935b: 210; Bodenheimer 1937: 101; Halperin & Sauter 1992: 121.

Material examined. 1 ♂, Huliot, 27.x.1968 (Shoham); 1 ♀, same data but 22.vi.1968; 1 ♀, Jerusalem, 6.vi.2004 (Kravchenko) (all TAU); 1 ♀, Nahal Oren, Mt. Carmel, light trap 29.x.1998 (Pavliček & Kravchenko); 1 ♀, ditto but 30.x.1997; 1 ♀, ditto but 19.vi.1998; 1 ♂, ditto but 21.v.1998 (all NMPC).

Host plant. In Israel the larvae have been recorded feeding on *Prosopis* sp. (Halperin & Sauter 1992: 121).

Distribution. Spain (with Canary Islands), Portugal, France, Malta, Italy (Sicilia and Sardinia), North Africa, Turkey, Middle East, India, Indonesia, Cameroon, Cape Verde, Ethiopia, Nigeria, Gambia, Kenya, Tanzania, Zimbabwe, Malawi, Namibia, South Africa (Karsholt & Riedl 1996: 120; Ponomarenko 2009: 283; De Prins & De Prins 2018).

Remarks. *Gelechia lamprostoma* was described from a single male from Sicily. Later it was moved to the genus *Anacampsis* by Wocke (1871), and the species was placed in the genus *Onebala* Walker, 1864 (Meyrick 1925: 138). Although Hodges (1986) had transferred the closely related *Gelechia zulu* Walsingham, 1881 to *Helcystogramma*, Old World lepidopterists (e.g. Karsholt & Riedl 1996: 120; Ponomarenko 2009: 283) listed *lamprostoma* in *Dichomeris* Hübner, 1818. The genitalia of *O. lamprostoma* match the genus *Helcystogramma* Zeller, 1877 and the species was placed in the latter genus in recent papers (Agassiz & Bidzilya 2016; Karsholt & Huemer 2017), but without any accompanying remarks.

Helcystogramma lutatella (Herrich-Schäffer, [1854])

Anacamptis lutatella Herrich-Schäffer, [1854]—Syst. Bearb. Schmett. Europ. 5 (65): 201.

Material examined. 1 ♂, Nahal Oren, Mt. Carmel, light trap 24.vi.1997 (Pavliček & Kravchenko) (gen. slide 19009, J. Šumpich) (NMPC).

Distribution. Europe, Caucasus, S Siberia (Junnilainen *et al.* 2010: 64), Israel (new record).

Acompsia ballotellus (Amsel, 1935) **comb. nov.**

Hypsolophus ballotellus Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 298, pl. 10, fig. 64.

Records. Amsel 1933: 126; Amsel 1935a: 298; Bodenheimer 1937: 101.

Material examined. Syntypes of *Hypsolophus ballotellus*: 1 ♀, Ain Karem b. Jerusalem, 24.iv.1930, ex. 1. *Ballota* (Amsel); 1 ♂, same data, but 14.v.1930 (SMNK); 1 ♀, same data but 23.iv.1930 (gen. slide GU651) (SMNK); 1 ♂, Katzrin vicinity, Zavitan stream, 21.iv.2018, evening collection, on *Ballota saxatilis* (Bidzilya) (gen. slide 152/18, O. Bidzilya) (ZMKU).

Host plant. Larvae live on *Ballota* spp. (Amsel 1935a: 298).

Distribution. Israel.

Remarks. *Hypsolophus ballotellus* was described from an unstated number of specimens (“24. 4, 14. 5, ex 1. *Ballota* sp.”) from Ain Karim, Jerusalem. A holotype was not selected in the original description, and three specimens in SMNK should be considered as syntypes.

Our study of the male genitalia of this species (Fig. 23) shows that it matches in all details to those of the genus *Acompsia* Hübner, [1825], subgenus *Telephila* Meyrick, 1923. Hence, the following new combination is proposed: *Acompsia* (*Telephila*) *ballotellus* (Amsel, 1935) **comb. nov.** Subgenus *Telephila* comprises two species—*A. (T.) schmidtiellus* (Heyden, 1848) and *A. (T.) syriaca* Huemer & Karsholt, 2002. *Acompsia ballotellus* differs from both these in the darker forewing with more extensive black pattern especially along the wing apex (Fig. 22). In *A. ballotellus* the apex of the forewing is moreover more acute, and the hindwings are darker than in *A. syriaca*. The male genitalia mostly resemble those of *A. schmidtiellus*, but the valva is narrower and the sclerite in the phallus is absent (Fig. 23). *Acompsia syriaca*, which is also without a sclerite in the more slender phallus, moreover differs by the longer arms of the vinculum and the different shape of the uncus (for comparison see Huemer & Karsholt (2002: figs 40, 57)).

Acompsia ballotellus is a little known species, and apart from Bodenheimer (1937: 101) it has to our knowledge not been mentioned in the literature since its description. There are a few references to it (as “*Brachmia bellotella*”) on the internet starting with Robinson *et al.* (2010–2018), but the species has apparently not been formally transferred to the genus *Brachmia* Hübner, 1825. *Hypsolophus* Illiger, 1801 is an emendation of *Ypsolopha* Latreille, 1796, the type genus of the family Ypsolophidae, and the placement of *ballotellus* in *Hypsolophus* may be the reason why the species was overlooked by authors of Gelechiidae literature.

Subfamily Apatetrinae Le Marchand, 1947

Tribe Pexicopiini Hodges, 1986

Harpagidia magnetella (Staudinger, 1871)

Gelechia magnetella Staudinger, 1871—Berl. ent. Z. 1870: 310.

Harpagidia pallidibasella Ragonot, 1895—Bull. Soc. ent. Fr. 1895: 107.

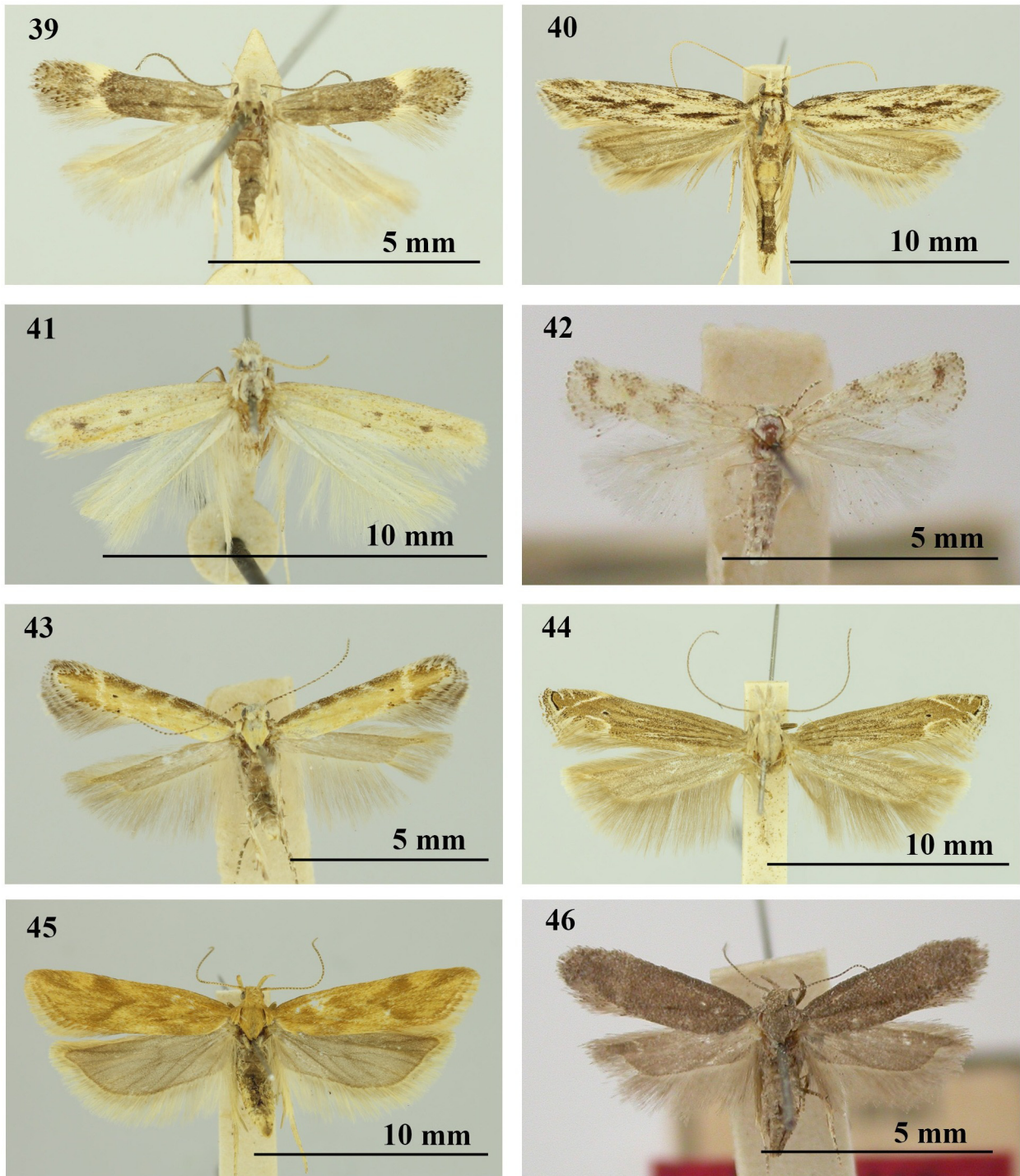
Gelechia melitophanes Meyrick, 1931—Exot. Microlep. 4: 85.

Records. Meyrick, 1931: 85 (*melitophanes*).

Material examined. 1 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (TAU).

Distribution. Greece, Turkey, Armenia, Iraq, Lebanon (Zerny 1934: 19), Israel.

Remarks. *Gelechia melitophanes* was described from a pair collected in "Palestine, Mt. Carmel, May" by S. Tapukhi (Meyrick 1931: 85). According to Clarke (1969:175) they are labelled "Place of Sacrifice, 2.5.30 S. Tapukhi. Carmel", both are males and both are in very poor condition.



FIGS 39–46. Type specimens of Gelechidae described from Israel and Palestine and some regional endemics. Adults. 39. *Schützeia languidella* Ams., syntype, ♂, Tel-Aviv, Düne, 25.v.1932. 40. *Oxypteryx jordanella* Rbl., ♂, Jericho, 22.ii.1931. 41. *Epidola semitica*, "Type", ♂, Ain Kerem, 20.vi.1930. 42. *Chrysophora halymella* Ams., "Type", Georgskloster, 21.iv.1930. 43. *Aristotelia brizelloides* Ams., syntype, ♂, Jericho, 31.v.1930. 44. *Paltodora striatella magna* Ams., lectotype, Kirjat Anawim, 28.iii.1930. 45. *Rhinosia flavella aflavella* Ams., paralectotype, ♂, Tabgha, Tiberias, März, 1930. 46. *Gelechia haifella* Ams., holotype, ♀, Karmel, 7.v.1930.

***Pexicopia malvella* (Hübner, 1805)**

Tinea malvella Hübner, 1805—Samml. eur. Schmett. [8]: pl. 41, fig. 281.

Recurvaria lutarea (Haworth, 1828)—Lepid. Britannica (4): 549.

Records. Amsel 1933: 125; Bodenheimer 1937: 100.

Distribution. Europe to Ural Mountains, North Africa, the Caucasus, Turkey, Central Asia to China (Xinjiang), SW Siberia (Li 2002: 432; Junnilainen *et al.* 2010: 65).

***Platyedra subcinerea* (Haworth, 1828)**

Recurvaria subcinerea Haworth, 1828—Lepid. Britannica (4): 548.

Gelechia vilella Zeller, 1847—Isis von Oken 1847 (11): 845.

Platyedra argillosella Herrich-Schäffer, 1855—Syst. Bearb. Schmett. Europ. 5 (58): fig. 514.

Gelechia bathrosticta Meyrick, 1936—Exotic Microlep. 5 (2): 43.

Records. Amsel 1933: 125 (*vilella*); Amsel 1935c: 263 (*vilella*); Bodenheimer 1937: 100 (*vilella*).

Material examined. 2 ♂, Jericho (Pal), Lichtfang, 11.iv.1930 (Amsel); 1 ♂, Jerusalem, Palastina, 27.vi.1931 (Einsler) (all SMNK); 1 ♂, Ramat, Hasharon, 5.ii.1972 (Herling) (all TAU); 1 ♂, Nahal Oren, Mt. Carmel, light trap, 24.vi.1997 (Pavliček & Kravchenko) (NMPC).

Distribution. Europe eastwards to Caucasus (Karsholt & Riedl 1996: 122; Ponomarenko 2008: 92), North Africa, Middle East, Iran, Transcaucasia, Afghanistan, Tadjikistan, Turkmenistan, Uzbekistan, Khabarovskiy krai of Russia (Lvovsky & Piskunov 1999: 75), USA (Lee *et al.* 2009: 32), Canada (Pohl *et al.* 2018: 83).

***Pectinophora gossypiella* (Saunders, 1844)**

Depressaria gossypiella Saunders, 1844—Trans. ent. Soc. Lond. 3: 285.

Gelechia umbripennis Swinhoe, 1885—Proc. Zool. Soc. Lond. 1885: 884.

Records. Buxton 1924: 334; Bodenheimer 1930a: 173; Bodenheimer 1930b: 347–349; Amsel 1933: 125; Bodenheimer 1937: 100; Bytinski-Salz 1966: 34; Kugler 1966: 85.

Material examined. 2 ♂, 6 ♀, Huliioth, 24, 27, 28.viii, 5.ix.1968 (Shoham); 1 ♀, Maoschaim, 7.ix.1968 (Shoham) (all TAU).

Host plant. A serious pest on cotton. In Israel larvae feed on *Gossypium* sp. Bodenheimer (1930b: 347–349) gives details on the biology.

Distribution. Tropical and subtropical regions of the world (CABI 2019).

***Sitotroga cerealella* (Olivier, 1789)**

Alucita cerealella Olivier, 1789—Encycl. Méth. Dict. Ins. 4: 121.

Tinea hordei Kirby, 1815—Introd. Ent. 1: 172.

Gelechia arctella Walker, 1864—List Spec. Lepid. Insects Colln Br. Mus. 29: 635.

Gelechia (*Sitotroga*?) *coarctatella* Zeller, 1877—Horae Soc. Ent. Ross. 13: 353.

Gelechia melanarthra Lower, 1900—Proc. Linn. Soc. N.S.W. 25 (3): 416.

Epitheatia palearis Meyrick, 1913—Exotic Microlep. 1 (3–5): 65.

Nesolechia horogramma Meyrick, 1921—Exotic Microlep. 2 (14): 425.

Syngenomictis aenictopa Meyrick, 1927—Insects Samoa 3: 78.

Aristotelia ochrescens Meyrick, 1938—Dt. ent. Z. Iris 52: 3.

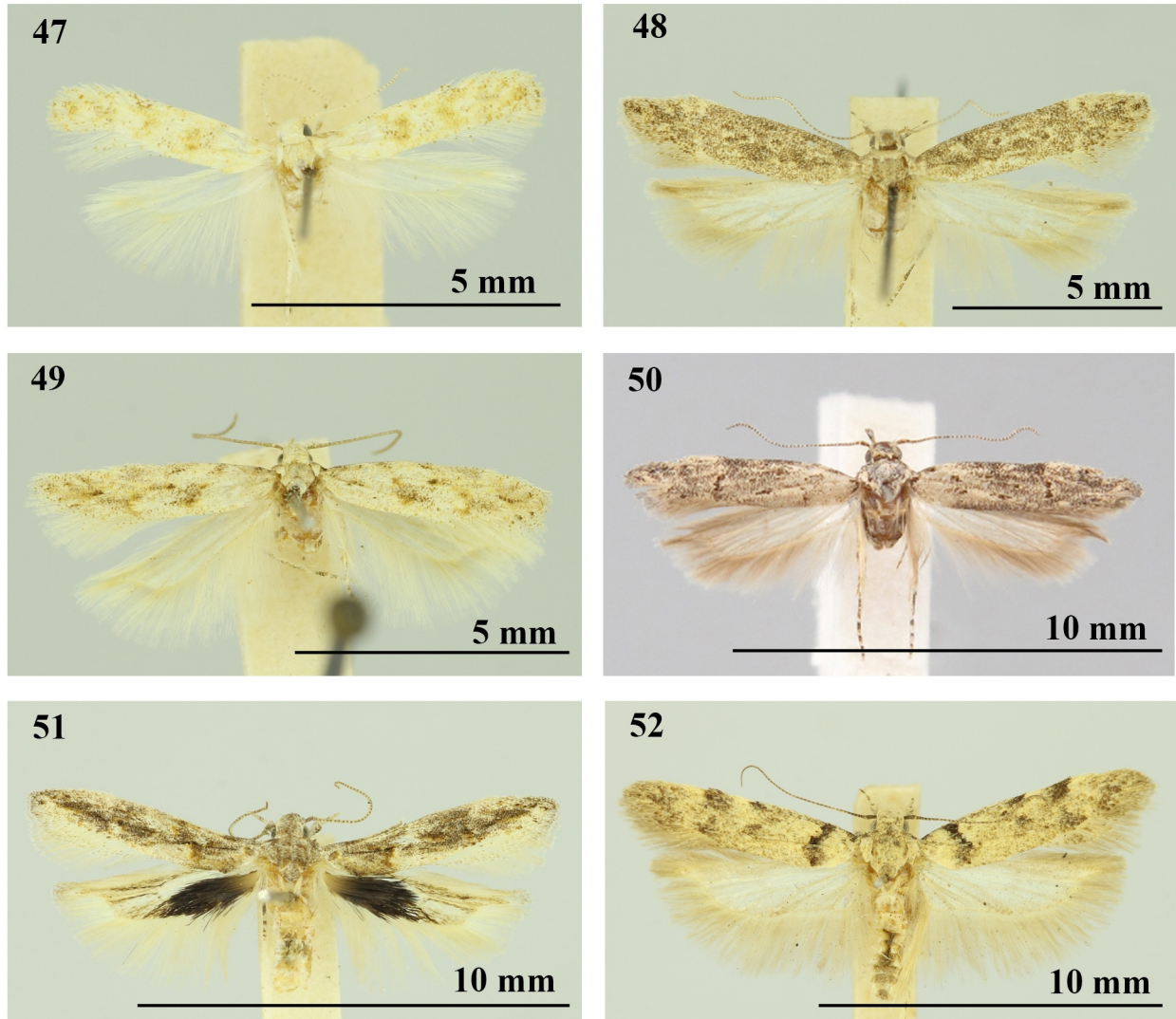
Chelaria asemodes Meyrick, 1938—Trans. R. ent. Soc. Lond. 87: 513.

Records. Bodenheimer 1930a: 174; Bodenheimer 1930b: 380–382; Amsel 1933: 126; Amsel 1935c: 264; Bodenheimer 1937: 101; Kugler 1966: 85; Donahay & Calderon 1969: 229.

Material examined. 1 ♂, Jerusalem, Palästina, 4.vi.1931 (Eisner); 1 ♂, 1 ♀, Abu-Goash, Jerusalem, 21.iv.1930 (Amsel) (all SMNK).

Host plant. In Israel the larvae were recorded as a very common pest, feeding on *Triticum* sp. (Bodenheimer 1930a: 174; 1930b: 380–382; 1937: 101).

Distribution. Tropical and subtropical regions of the world (CABI 2019).



FIGS 47–52. Type specimens of Gelechidae described from Israel and Palestine and some regional endemics. Adults. 47. *Scrobipalpa voltinelloides* Pov., holotype, ♂, Jericho, 31.v.1930. 48. *Gnorimoschema pseudobsolletum* Pov. & Gregor, holotype, ♀, Südende d. Toten Meers, 15-27.iii.1933. 49. *Lita rhamnifoliae* Ams. & Hering, lectotype, ♂, Jerusalem, 6.iv.1930. 50. *Is-trianis kravchenkoi* Bidz. & Karsh., holotype, ♂, Jericho, May, 2004. 51. *Pseudoteleia squamodorella* Ams., ♂, Kasr ek Jehud, 9.iv.1930. 52. *Gelechia hessi* Ams., lectotype, ♂, Georgskloster, 15.iv.1930.

Tribe Apatetrini Le Marchand, 1947

Chrysoesthia halymella (Amsel, 1935)

Fig. 42

Chrysophora halymella Amsel & Hering, 1931—Dt. ent. Z. Iris 1931: 124, pl. 2, fig. 11 (nomenclatorially unavailable).
Chrysophora halymella Amsel, 1935—Mus. Berl. 20 (2): 302.

Records. Amsel & Hering 1931: 124; Amsel 1933: 126; Amsel 1935a: 302; Amsel 1935c: 264; Bodenheimer 1937: 101.

Material examined. 1 ♀, “Georgskloster, *Atriplex haly.*, 21.4.1930., Z. 293” | “Palästina Expedition, 18.2-4.6.1930, H.G. Amsel” | “Typus, leg. H. Amsel “(SMNK).

Host-plant. *Atriplex halymus* L. (Amsel & Hering 1931: 124; Amsel 1935a: 302).

Distribution. Israel.

Remarks. *Chrysophora halymella* was first described from a few leaf mines found in early April. The description dealt only with the leaf mine, although it is apparent from the text that some moths emerged (“Die Puppenruhe dauerte etwa 8 bis 10 Tage” [=the pupation rest lasted about eight to ten days]). The description by Amsel & Hering (1931) is nomenclatorially invalid according to ICZN (1999: article 13.6.2), stating: “A name proposed after 1930 which is based on the work of an extant animal is excluded from zoological nomenclature [Art. 1.3.6]”. *Chrysophora halymella* was later made nomenclatorially available by Amsel (1935a: 302).

According to the original description, an unstated number of specimens were bred from *Atriplex halymus* L. in Georgskloster 21.iv. The single female in SMNK labelled as “type” should be considered as a syntype.

***Chrysoesthia amseli* (Bidzilya, 2008) comb. nov.**

Metanarsia amseli Bidzilya, 2008—SHILAP Revta lepid. 36 (144): 534.

Material examined. 1 ♂, 6 km S Sode Boger, Avdat, 15.x.1987 (Dorn) (gen. prep. 7778, G. Derra) (coll. G. Derra); 6 ♂, Negev, Nahal (Wadi) Zin, 8.x.1999, black light (Pavliček) (gen. slide 115/18, O. Bidzilya) (NMPC).

Remarks. *Metanarsia amseli* was described from two females collected in South Iran (Bidzilya 2008). The males from Israel are identical externally to types of *M. amseli*, but the black pattern is more distinct (Fig. 28). Bidzilya (2005) considered that *Metanarsia* Staudinger, 1871 can be separated from *Chrysoesthia* Hübner, 1825 by the absence of the signum in the corpus bursae of the female genitalia. However, Karsholt & Vives Moreno (2014) noted that the absence of pecten on antennal scape and short labial palpus in *Chrysoesthia* are more reliable characters for separating this genus from *Metanarsia*. Based on this suggestion we transfer *M. amseli* to *Chrysoesthia*: *Chrysoesthia amseli* (Bidzilya, 2008) **comb. nov.** Below we provide the first description of the male genitalia of this species.

Male genitalia (Fig. 29). Uncus rounded; gnathos narrow, elongated, membranous, with rounded apex; tegumen trapezoidal, anterior margin straight; valva broad at base, distal portion straight, apex weakly broadened, rounded, extending to about half length of uncus; basal part of sacculus broad, sub-rectangular, inner margin with two or three denticles, posteromedial corner rounded, distal half narrow, digitate, straight, extending to $\frac{3}{4}$ length of valva; saccus stout, broad at base, then narrowed towards truncate apex; phallus as long as valva, weakly constricted in middle, distal $\frac{3}{4}$ narrowed, apex weakly pointed.

Remarks. The male genitalia are most similar to those of *Ch. hispanica* Karsholt & Vives, 2014 but differ in sub-triangular rather than horseshoe shaped saccus and apically less narrowed phallus. The two species are externally different.

Biology. Host plant unknown. Adults have been collected in April in Iran and in October in Israel.

Distribution. South Iran, Israel (new record).

Subfamily Thiotrichinae Karsholt, Mutanen, Lee & Kaila, 2013

***Palumbina guerinii* (Stainton, 1858)**

Stathmopoda (?) *guerinii* Stainton, 1858—Ent. Annual: 152.

Palumbina terebintella Rondani, 1876—Bull. Soc. ent. Ital. 8: 23.

Tinea pistaciae Anagnostopoulos, 1935—The Pistachio Nut Tree in Greece: 50, figs 14–20.

Records. Halperin 1986: 126, 28, 130, 131.

Material examined. 4 ♂, 2 ♀, Em Karem, 10.ix.1975, *Pistacia palaestinensis*, 26.ix, 1.x.1975 (Halperin); 1 ♀, Kfar Hassidin, 17.x.1974, *Pistacia atlantica*, 18.xi.1974 (Halperin) (all TAU); 2 ♂, 2 ♀, Tel-Aviv, Botanical garden. 14, 16.iv.2018, light (Bidzilya & Kravchenko) (ZMKU).

Host plant. In Israel larvae feed on *Pistacia terebinthus* ssp. *palaestina* (Boiss.) Engl. and *P. atlantica* Desf. (Halperin 1986: 126, 128, 130, 131). They were also observed in galls of aphids (Hemiptera: Eriosomatidae, Fordinae) on *Pistacia* spp. and in galls of *Tetraneura caerulescens* (Passerini, 1856) (Hemiptera: Aphididae) on *Ulmus* sp. (Halperin & Sauter 1992: 134).

Distribution. South Europe (Karsholt & Riedl 1996: 118), Morocco, Lebanon, Cyprus, Turkey (Sattler 1982: 27), Israel, Kenya (Miller *et al.* 2014: 140).

***Polyhymno chionarcha* Meyrick, 1913**

Fig. 24

Polyhymno chionarcha Meyrick, 1913—Ann. Transv. Mus. 3 (4): 282.

Polyhymno abaiella Amsel, 1974—Beitr. naturk. Forsch. SüdWdtl. 33: 201. **Syn. nov.**

Material examined. 1 ♂, Wingeit, 26.viii.1968 (TAU).

Additional material. 1 ♂, 1 ♀, SW Arabien, Wadi Tihama, 850 m, 23.iv.1979, Asirgrbirge (Amsel) (gen. slide 85/19♂; 86/19♀, O. Bidzilya) (SMNK). *Polyhymno abaiella* 2♂, 1 ♀, S Iran, Strasse Bandar-Abbas-Sirjan, km 24, 250 m, 2.iv.1972 (Amsel) (gen. slide 87/19♂; 88/19♀, O. Bidzilya) (SMNK).

Host plant. Host plant in Israel unknown. In Kenya larvae feed on *Acacia tortilis* (Forssk.) Heyne and *Vachellia xanthophloea* (Benth.) Hurter (Agassiz & Bidzilya 2016).

Distribution. South Africa, Namibia, Zimbabwe, Mozambique, Tanzania, Kenya (Agassiz & Bidzilya 2016: 128), Israel (new record), S Iran, Saudia Arabia (new record).

Remarks. *Polyhymno abaiella* was described based on a series of specimens collected in South Iran. The species is similar externally to *P. chionarcha* except for brown pattern along costal margin which are reduced in most specimens from the type series. The specimens with fully or partially reduced brown irroration along the costa occur both among type specimens of *P. abaiella* and *P. chionarcha* from Africa, thus this character is rather variable. The genitalia of both sexes of *P. abaiella* fully agree with those of *P. chionarcha* studied by us from South Africa, Kenya and Saudia Arabia. Hence, we consider *Polyhymno abaiella* Amsel, 1974 **syn. nov.** of *P. chionarcha* Meyrick, 1913.

***Polyhymno dumonti* (Hartig, 1936) comb. nov.**

Stigmatoptera dumonti Hartig, 1936—Z. öst. EntVer. 21: 45, pl. 2, fig. 11.

Material examined. 1 ♂, Ein-Gedi, 17.ix.1968 (gen. slide 123/18, O. Bidzilya) (TAU).

Additional material. Paratype of *Stigmatoptera dumonti* ♂, Tunisie merid., Bou-Hedma, 8.vi.1929 (Dumont) (SMNK); 1 ♂, Libya, Gharian, Wadi el Hira, 15.iv.1983 (Seneca); 1 ♀, same data but 27.v.1983 (ZMUC); 1 ♀, Sudan, El Damer, Hudelba, 15.iii.1962 (Remane) | K. Sattler, B. M. (N.H.), 1974-570 (NHMUK).

Remarks. The genus *Stigmatoptera* Hartig, 1936 was established as monotypic for the new species *S. dumonti* collected in South Tunisia. *Stigmatoptera dumonti* agrees both externally (Fig. 25) and in the male genitalia (Fig. 26) with species of the genus *Polyhymno* Chambers, 1874. Hence, we synonymize *Stigmatoptera* with the latter here: *Stigmatoptera* Hartig, 1936 **syn. nov.** of *Polyhymno* Chambers, 1874.

Here we describe the male genitalia of *S. dumonti* for the first time.

Male genitalia. Uncus parallel-sided, twice as long as broad, posterior corners rounded; gnathos hook weakly curved, extending anteriorly to about 1/3 of tegumen; tegumen trapezoidal, weakly narrowed posteriorly, anterior margin with short rounded medial emargination; valva slightly shorter than tegumen, broad at base, distal portion straight, narrow, with strongly sclerotized ventral margin, apex rounded; sacculus short, hump-shaped; saccus about ¾ length of valva, gradually narrowed apically, constricted after half length, apex rounded; phallus strongly swollen at base, distal portion as long as basal portion, narrow, gradually tapering, with short narrow thorn before middle and on the apex, ductus ejaculatorius containing well developed lamina.

Remarks. *Polyhymno dumonti* externally resembles *P. hostilis* (Meyrick, 1918), but the forewing is light brown rather than dark brown, white pattern along costal margin is broader, medial white area is narrower and white medial

subtriangular streak before apex is absent. Two thorns in the phallus are characteristic for *P. albifulvella* Agassiz & Bidzilya, 2016 too, but the latter differs externally and the thorns are located in middle of the phallus.

Distribution. Tunisia, Israel (new record), Libya (new record), Sudan (new record).

Subfamily Anomologinae Meyrick, 1926

Parapodia sinaica (Frauenfeld, 1859)

Gelechia sinaica Frauenfeld, 1859—Verh. zool.-bot. Ges. Wien 9: 323.

Parapodia tamaricicola Walsingham, 1911—Bull. Soc. Ent. Fr. 1912: 305.

Cecidonostola tamariciella Amsel, 1958—Beitr. naturk. Forsch. SüdwDtschl. 17 (1): 81.

Records. Amsel 1933: 126; Amsel 1935b: 210; Amsel 1935c: 264; Bodenheimer 1937: 101; Halperin & Sauter 1992: 127.

Material examined. 1 ♀, Nahal David Spill, nr. Ein Gedi, 27.iii.1973, coll. S. Umon | Na-3-1570-4322, ex/en small galls, plant col. on: 13.iii.1972 (SMNK); 2 ♂, Field School, Hazeva, 28.iii, 6.iv.1998 (Ashkenazi); 1 ♂, Shezaf N.R., Nahal Shahaq, 3.iv.1999, Malaise Trap (Yarom); 1 ♀, Israel, N. David, 20.iii.1972 (Furth) (all TAU).

Host plant. In Israel larvae were recorded feeding on *Tamarix* sp. (Bodenheimer 1937: 101; Halperin & Sauter 1992: 127).

Distribution. South France, Spain, North Africa, Cyprus, Middle East, NE Arabia, Iraq, Turkmenistan, Uzbekistan (Sattler 1962: 53; Ivinskis & Piskunovas 1994: 42; Vives Moreno 2014: 173).

Epidola semitica Amsel, 1942

Fig. 41

Epidola semitica Amsel, 1942—Veröff. dt. Kol. Übersee-Mus. 3: 221, pl. 16, fig. 10.

Material. “Type” of *semitica* ♂, Ain Kerem, Jerusalem, 20.vi.1930 (Einsler) (gen. slide GU 58); “type” of *semitica*, 1 ♀, same data; 1 ♀ (abdomen missing), same data (all SMNK); 1 ♀, Wingeit, 11.v (gen. slide 199/18, O. Bidzilya) (TAU).

Distribution. Israel.

Remarks. *Epidola semitica* was described from a single (worn) male collected by W. Einsler 20th June 1930 at Ain Karem near Jerusalem. The holotype was stated to be deposited in the collection of the Deutsches Kolonial- und Übersee-Museum in Bremen, but could not be traced there during a visit by OK in 2008. No additional specimens were mentioned in the original description, although we were able to examine three additional specimens in SMNK, collected on the same occasion. Two of them, male and female, bear a “type” label. It is unclear to us if any of these represents the holotype.

Amblypalpis olivierella Ragonot, 1886

Amblypalpis olivierella Ragonot, 1886—Ann. Soc. ent. Fr. (6) 5 (Bull.): ccix.

Records. Bodenheimer 1930a: 174; Bodenheimer 1930b: 369; Amsel 1933: 127; Bodenheimer 1937: 102; Halperin 1986: 122, 124, 129, 131.

Material examined. 1 ♂, Mikmoret, 9 km N Netanya, 20.xii.1970, coll. D. Gerling | Ta.-0-8, ex/spinde gall, plant. col. on: 17.xii.1970 (SMNK); 1 ♂, Micmoret, 20.ii.1977 (Lupo); 1 ♂, Holon, 26.ii.1977 (Lupo); 2 ♂, Netanya, 25.ix.1984, *Tamarix*, 20, 23.xi.1984 (Halperin) (all TAU); 2 ♂, Nahal Kziv, light trap, 5.xii.1998 (Pavliček & Kravchenko); 1 ♂, Nahal Oren, Mt. Carmel, light trap, 9.xii.1998 (Pavliček & Kravchenko) (all NMPC).

Host plant. Stem galls of *Tamarix* sp. (Halperin 1986: 122, 124, 129, 131). According to Bodenheimer (1930a: 174; 1930b: 369) the larvae are so heavily parasitized that moths very rarely emerge from the galls.

Distribution. Algeria, Tunisia, Israel, Saudi Arabia (Amsel 1958: 81), Spain (Vives Moreno 2019: 53).

Aristotelia brizelloides Amsel, 1935

Fig. 43

Aristotelia brizelloides Amsel, 1935—Mitt. Zool. Mus. Berl. 20 (2): 302.

Records. Bodenheimer 1930a: 174 (*brizella*); Amsel 1933: 125; Amsel 1935a: 302; Amsel 1935c: 264; Bodenheimer 1937: 101.

Material examined. Syntypes of *Aristotelia brizelloides*: 1 ♂, Jericho (Pal.), Lichtfang, 31.v.1930 (Amsel) (gen. prep. 252/08, O. Bidzilya); 2 ♂, 1 ♀, same data (gen. prep. GU 715, GU 716); 1 ♀, same data but 16.iv.1930; 1 ♂, Georgskloster, Wadi el Kelt, Lichtfang, 21.vi.1930 (all SMNK); 3 ♂, 1 ♀, Jericho, Kalia, v.2004 (Kravchenko) (gen. slide 5496 Hendriksen) (TAU, ZMUC).

Distribution. Israel.

Remarks. *Aristotelia brizelloides* was described from an unstated number of specimens collected at Georgskloster 16.iv, 21 and 31.v. A holotype was not selected in the original description. Six specimens from these dates in SMNK are labelled “Jericho”, which is situated 7 km east of Georgskloster (Amsel 1935c: 228), and these should be considered as syntypes.

The species is closely related to *A. brizella*, but seems to differ in small details of the male genitalia. The genus *Aristotelia* is in need of revision, and we prefer to retain *A. brizelloides* as distinct pending future revisionary work, preferably including molecular data.

Dirhinosia nitidula (Stainton, 1867)

Gelechia nitidula Stainton, 1867—Tineina of Syria and Asia Minor: 44.

Records. Stainton 1867: 45; Amsel, 1933: 126; Bodenheimer 1937: 101; Tokár & Gozmány 2004: 71.

Material examined. Lectotype of *Gelechia nitidula* ♂, “*Gelechia nitidula* / St., Palestine, Plains of Jordan, Cambridge 1865, Tristram J. J. 1847, B. M. genitalia slide ♂ no. 12726, Walsingham Collection, 1910–427” (coll. NHMUK); 4 ♂, 1 ♀, Karmel, Haifa, 7.v.1930 (Amsel) (SMNK).

Distribution. Israel, Syria, Lebanon, Turkey (Tokár & Gozmány 2004: 72–73).

Remarks. *Gelechia nitidula* was described from two males and three females from the “plains of Jordan”. The lectotype cited above was designated by Tokár & Gozmány (2004: 72).

Dirhinosia arnoldiella (Rebel, 1905)

Rhinosia arnoldiella Rebel, 1905—Ann. nat. Mus. Wien 20: 212.

Records. Amsel 1933: 126; Amsel 1935c: 264; Bodenheimer 1937: 101; Tokár & Gozmány 2004: 72.

Material examined. 2 ♂, Kefar, Shamuël, 29.iv.1968; 1 ♂, Latrain, 25.iv.1968 (TAU).

Distribution. Greece, Israel, Turkey (Tokár & Gozmány 2004: 73).

Megacraspedus albella (Amsel, 1935)

Chilopselaphus albella Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 302, pl. 10, fig. 57.

Records. Amsel 1933: 127 Amsel 1935c: 266; Bodenheimer 1937: 102; Huemer & Karsholt 2018: 114–115.

Distribution. Iran, Israel (Huemer & Karsholt 2018).

Remarks. *Chilopsephalus albella* was described from few specimens (“selten”) collected by H. G. Amsel at “Georgskloster”, 1st of April 1930. A lectotype was published by Huemer & Karsholt (2018: 144).

***Ptocheuusa inopella* (Zeller, 1839)**

Gelechia (Brachmia) inopella Zeller, 1839—Isis [32] 1839 (3): 201.

Ptocheuusa amesella Chrétien, 1907—Bull. Soc. ent. Fr. 1907: 339.

Records. Amsel 1933: 126; Amsel 1935c: 264; Bodenheimer 1937: 101.

Material examined. 1 ♂, Ain Karim, Jerusalem (Pal.), 18.v.1930 (Amsel) (gen. slide 83/19, O. Bidzilya) (SMNK).

Distribution. Europe eastward to Ural Mountains (Junnilainen *et al.* 2010: 22), Israel.

***Ptocheuusa paupella* (Zeller, 1847)**

Gelechia paupella Zeller, 1847—Isis von Oken 1847 (11): 858.

Apheloseitia inulella Curtis, 1850—Ann. Mag. nat. Hist. (2) 5 (26): 117.

Apatetris leucoglypta Meyrick, 1918—Exotic Microlep. 2 (4): 117.

Material examined. 1 ♂, 1 ♀, Kefar Shamuël, 22.viii.1968 (gen. slide 17/19, O. Bidzilya) (TAU).

Distribution. Europe, India (?) (Elsner *et al.* 1999), Israel (new record).

***Oxypteryx immaculatella* (Douglas, 1850) comb. nov.**

Figs 31, 32

Gelechia immaculatella Douglas, 1850—Trans. ent. Soc. Lond. (n.s.) 1: 67.

Eulamprotes phaeella Heckford & Langmaid, 1988—Ent. Gaz. 39: 2.

Material examined. 4 ♂, Keshet, 21.iv.2018, at light (Kravchenko & Bidzilya) (gen. slide 124/18, O. Bidzilya) (TAU, ZMKU).

Distribution. Great Britain, Denmark, Germany, Portugal, Spain (Heckford *et al.* 1999; Gaedike *et al.* 2017), Israel (new record).

Remarks. The species is closely related to *O. atrella*. The record of latter species from Israel needs verification (see remarks under incorrect and doubtful records for Israel).

***Oxypteryx jordanella* Rebel, 1911**

Fig. 40

Oxypteryx jordanella Rebel, 1911—Verh. zool-bot. Ges. Wien 61: 151.

Records. Amsel 1933: 126; Amsel 1935c: 265; Bodenheimer 1937: 101.

Material examined. 1 ♂, Jericho, Palästina, 22.ii.1931 (Amsel) (gen. slide 84/19 O. Bidzilya) (SMNK).

Remarks. The genus *Oxypteryx* Rebel, 1911 was established as monotypic for the new species *O. jordanella* described on the basis of two females collected in Jordan. The genus was considered as a synonym of *Acompsia* by Meyrick (1925: 142) or as a separate genus by Amsel (1935c: 265). The male genitalia of *Oxypteryx* (Fig. 30) agree in all details with *Eulamprotes* Bradley, 1971 so that the following synonymy is proposed: *Eulamprotes* Bradley, 1971 **syn. nov.** of *Oxypteryx* Rebel, 1911.

Here we describe the male genitalia of *O. jordanella* for the first time.

Male genitalia. Uncus very small, hump-shaped, with a few apical setae; gnathos reduced; tegumen trapezoidal, as long as broad at base, anteromedial emargination four times as broad as long, subrectangular; valva elongated, parallel-sided to 2/3 length, then narrowed towards pointed apex, with subapical fold; sacculus short, subtriangular, 1/3-1/4 length of valva; transtilla lobes rounded; vinculum band-shaped; saccus short, weakly narrowed towards

rounded apex; phallus three times as long as broad, slightly longer than valva, distal 1/3 weakly narrowed, with sub-ovate membranous area in basal portion, vesica without cornuti.

Remarks. *Oxypteryx jordanella* is well recognized externally by its large size and for *Eulamprotes* unique wing pattern. The short saccus is the most characteristic feature in the male genitalia.

Distribution. Israel, Jordan, Saudi Arabia (Amsel 1961: 59).

Isophrictis magna (Amsel, 1935)

Fig. 44

Paltodora striatella magna Amsel, 1935—Veröff. dt. Kol. Übersee-Mus. 1 (2): 265.

Records. Stainton 1867: 46 (*striatella*); Amsel 1935c: 265; Amsel 1936: pl. 15, fig. 28–31; Bodenheimer 1937: 101 (*Isophrictis striatella magna*); Povolný 1983: 142, figs 21, 22.

Material examined. Lectotype of *Paltodora striatella magna* ♂, Kirjat Anawim, Jerusalem, 28.iii.1930 (Amsel) (gen. slide Ams. 5444) (SMNK). Paralectotypes of *magna*, 4 ♂, 2 ♀, same data (gen. slide 58/08♂; 145/17♀, O. Bidzilya); 1 ♂, Ramalah, Jerusalem, 21.iv.1930 (gen. slide 147/17, O. Bidzilya) (Amsel) (all SMNK).

Distribution. Israel; possibly also Greece (Crete) (Huemer & Karsholt 2017: 174).

Remarks. *Paltodora striatella magna* was described from a series of both sexes collected in the vicinity of Jerusalem. Based on study of type material, Povolný (1983) showed that *magna* is a separate species that differs from *I. striatella*, *I. anthemidella* (Wocke, 1871) and some other species of *Isophrictis*. However, the status of *magna* is rather questionable as the genus is in need of revision.

Isophrictis kefersteiniellus (Zeller, 1850)

Ypsolophus kefersteiniellus Zeller, 1850—Stettin. Ent. Ztg. 11 (5): 142.

Records. Bodenheimer 1930a: 174; Amsel 1933: 126; Bodenheimer 1937: 101.

Material examined. 1 ♂, Kirjat Anawim, Jerusalem, 2.v.1930 (Amsel) (SMNK); 2 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (TAU).

Distribution. Spain, Portugal, France, Italia (with Sardinia and Sicily), Malta, Balkan Peninsula, Greece (Crete) (Karsholt & Riedl 1996: 105), Israel.

Monochroa lucidella (Stephens, 1834)

Cleodora lucidella Stephens, 1834—Ill. Br. Ent. (Haustellata) 4 (2): 221.

Xystophora scordiscella Rebel, 1904—Ann. Mus. Wien 19: 352.

Catabranchmia unipunctella Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 297, pl. 10, fig. 63.

Monochroa lucidella immaculatella Huemer, 1996—Z. Arb.Gem. öst. Ent. 48: 23.

Records. Amsel 1935a: 297 (*unipunctella*); Bodenheimer 1937: 101 (*unipunctella*); Sattler (1992: 108).

Material examined. Lectotype of *Catabranchmia unipunctella* ♂, Jordanbrücke bei Jericho, 29.iv.30, Amsel, Lichtfng. | Palästina Expedition, 18.ii-4.vi.1930 (Amsel) | Lectotype *Catabranchmia unipunctella* Ams., teste K. Sattler, 1980, ♂ | GENITALIA: K. Sattler, 734 | (gen. slide Nr. 5790♂) (SMNK).

Distribution. Europe, Middle East, Russia (Zabaikalskiy krai, Amur region) (Ponomarenko 2008: 89; Junnilainen *et al.* 2010: 24).

Remarks. The lectotype was designated by Sattler (1992: 108).

***Monochroa hornigi* (Staudinger, 1883)**

Doryphora hornigi Staudinger, 1883—Stett. ent. Z. 44: 184.

Monochroa nordmanella Bruun, 1958—Notul. ent. 37: 118.

Material examined. 2 ♂, 1 ♀, Jerusalem, 6.vi.2004 (Kravchenko) (TAU, ZMUC).

Distribution. Central and North Europe, Italy, Portugal (Karsholt 2011), Israel (new record).

***Metzneria aestivella* (Zeller, 1839)**

Gelechia aestivella Zeller, 1839—Isis [32] 1839 (3): 202.

Gelechia carlinella Stainton, 1851—Suppl. Cat. Br. Tineidae and Pteroph.: 5.

Gelechia selaginella Mann, 1855—Verh. zool.-bot. Ges. Wien 5 (Abh.): 565.

Parasia torridella Mann, 1859—Wien. Ent. Mon. 3 (6): 174.

Records. Amsel 1933: 124; Amsel 1935c: 262 (*selaginella*); Bodenheimer 1937: 100 (*selaginella*); Englert 1974: 395.

Material examined. 2 ♂, Georgskloster, Wadi el Kelt, 21.v.1930, Lichtfg. (Amsel) (GU 321, Englert); 1 ♂, 2 ♀, Ain Karem, Jerusalem, 14.v., 19.viii.1930 (Amsel); 3 ♂, 3 ♀, Tel-Aviv (Düne), 21.viii.1930 (Einsler); 1 ♂, Jericho, Palästine, 31.v.1930 (Amsel); 1 ex., Kirjat Anawim, Jerusalem, 2.v.1930 (Amsel) (all SMNK); 12 exx, slopes by Yflach, 31.v.2004 (Kravchenko); 1 ♂, 30 exx, Jerusalem, 6.vi.2004 (Kravchenko) (TAU, ZMUC).

Distribution. Europe, North Africa, Middle East (Syria, Israel), Turkey (Englert 1974: 395). A record from Zabaikalskiy krai of Russia (Bidzilya 2000: 287) refers to *M. transbaikalica* Bidzilya, 2018 (Bidzilya & Nupponen 2018: 383).

***Metzneria agraphella* (Ragonot, 1895)**

Parasia agraphella Ragonot, 1895—Bull. Soc. ent. Fr. 1895: cvi.

Metzneria incognita Walsingham, 1904—Ent. Month. Mag. 40: 220.

Records. Amsel 1933: 125; Amsel 1935c: 263; Bodenheimer 1937: 100; Englert 1974: 407.

Material examined. 1 ♂, Mizpe Harel, 31°48'N 34°58'E, 28.vii.2013 (Kuslitzky) | flower head *Echinops* (TAU); 2 ♂, Jerusalem, 6.vi.2004 (Kravchenko) gen. slide 5500 Hendriksen (TAU, ZMUC); 1 ♂, 1 ♀, Nahal Oren, Mt. Carmel, wally bottom, 29.ix.1998 (Pavliček & Kravchenko); 1 ♂, ditto but 27.v.1999; 2 ♂, ditto but 21.v.1998; 1 ♀, ditto but 17.v.1997; 3 ♂, 1 ♀, Golan Heights, Masd al Shams, oak forest, 26.viii.2000 (Pavliček) (all NMPC).

Host plant. In Israel the larva was bred from flower head of *Echinops* sp. (first host plant record).

Distribution. Portugal (Corley: 215: 88), Morocco, Algeria, Greece (Crete), Turkey, Syria, Israel (Englert 1974: 407).

***Metzneria aprilella* (Herrich-Schäffer, 1854)**

Parasia aprilella Herrich-Schäffer, 1854—Syst. Bearb. Schmett. Europ. 5 (65): 207, (64) fig. 963.

Parasia neuropteralla var. *igneella* Tengström, 1859—Förhandl. Sällsk. Fauna Fenn.: 183.

Metzneria sanguinolentella Joannis, 1910—Bull. Soc. ent. Fr. 1910: 295.

Records. Bodenheimer 1930a: 173; Amsel 1933: 125; Bodenheimer 1937: 100.

Distribution. Europe, Turkey, Middle East (Lebanon, Syria, Israel), Armenia, Kazakhstan, Uzbekistan, Iran, Asiatic part of Russia (Kemerevo region, Altai) (Englert 1974: 409; Elsner *et al.* 1999: 25; Bidzilya 2002: 65; Ponomarenko 2008: 88; Falkovitsh & Bidzilya 2009: 70).

Remarks. The variable *M. aprilella* probably represents an unrevised species complex.

***Metzneria artificella* (Herrich-Schäffer, 1861)**

Anacampsis artificella Herrich-Schäffer, 1861—Neue Schmett. (3): 31, fig. 158.

Parasia litigiosella Millièrè, 1879—Naturaliste 1 (18): 139.

Metzneria pannonicella Rebel, 1915—Rovart. Lapok 22: 186.

Records. Amsel 1933: 124 (*pannonicella*); Amsel 1935c: 263 (*litigiosella*); Bodenheimer 1937: 100 (*pannonicella*).

Material examined. 1 ♀, Kirjat Anawim, 22.iv.1930 (Amsel) (GU 204 W.E.); 1 ♀, Jericho, Palastina, 11.iv.1930 (Amsel) (all SMNK); 4 ♂, 2 ♀, Nizzanim, 17.vi.2012, seedhead *Echinops philistaeus* (Freidberg) (TAU); 1 ♂, 2 ♀, Jerusalem, 6.vi.2004 (Kravchenko) (TAU, ZMUC).

Host plant. In Israel larvae were bred from *Echinops philistaeus* Feinbrun & Zohary (new host plant record).

Distribution. South and East Europe to South Ural and the Volga region (Junnilainen *et al.* 2010: 21), Zabai-kalskiy krai of Russia (Ponomarenko 2008: 88), China (Hebei) (Li 2002: 110), Lebanon (Zerny 1934: 18), Israel.

***Metzneria aspretella* Lederer, 1869**

Metzneria aspretella Lederer, 1869—Horae Soc. ent. ross. 6 (2): 92, pl. 5, fig. 13.

Records. Bodenheimer 1930a: 173; 1937: 100.

Material examined. 1 ♂, 1 ♀, Har Hermon, 2000 m, 28.vi.2012 (Freidberg) | *Cousinia hermonis* (2011) collected in 2012 (gen. slide 143/18, O. Bidzilya) (TAU).

Host plant. In Israel the larva was bred from *Cousinia libanotica* DC. (as *C. hermonis*) (new host plant record).

Distribution. Iran, Afghanistan (Englert 1974), Israel.

***Metzneria clitella* Rebel, 1903**

Metzneria clitella Rebel, 1903—Verh. zool.-bot. Ges. Wien 53: 411.

Metzneria tricolor Turati, 1930—Atti Soc. Ital. Sci. nat. 69: 79.

Records. Amsel, 1933: 124; Bodenheimer 1937: 100; Englert 1974: 401.

Material examined. 1 ♂, Jericho, Palastina, 11.iv.1930 (Amsel) (gen. slide GU149, Englert) (SMNK).

Distribution. Algeria, Tunisia, Libya, Israel, South Iran (Turati 1930: 79; Englert 1974: 401).

***Metzneria diamondi* Amsel, 1949**

Metzneria diamondi Amsel, 1949—Bull. Soc. Fouad Ier Ent. 33: 321.

Records. Englert, 1974: 408.

Distribution. Saudi Arabia, Israel, Afghanistan, Iran, Uzbekistan (Englert 1974: 408; Falkovitsh & Bidzilya 2009: 70).

***Metzneria ehikeella* Gozmány, 1954**

Metzneria ehikeella Gozmány, 1954—Ann. hist.-nat. Mus. Natn. Hung. 46: 276.

Material examined. 2 ♂, Bet Dagan, 17.viii.2009, 5.ix.2010 (Kuslitzky) | seedhead *Centaurea procurrens* (gen.

slide 153/18, O. Bidzilya); 1 ♂, Mizpe Netofa, 15.vii.2010 (Kuslitzky) | seedhead *Carlina curetum*; 1 ♂, 2 ♀, Rehovod., 10.vi., 7.vii.2009 (Kuslitzky) | reared from *Centaurea procurrens*; 1 ♀, Rehovot, , 20.viii.2010 (Kuslitzky) / flower of *Centaurea procurrens*; 1 ♂, 1 ♀, same data but 2.viii.2010 (Argov) / seedhead *Centaurea procurrens* (TAU); 1 ♀, Giv'at Brenner, 12.vii.2009 (Kuslitzky) | from flower of *Centaurea procurrens*; 3 ♀, Nes Ziyayona, 5.viii.2010 (Kuslitzky) | flower of *Centaurea procurrens*; 1 ♀, Mashmar Dawid, 21.vii.2010 (Kuslitzky) | seedhead *Centaurea hyalolepis* (TAU, ZMKU).

Host plant. In Israel larvae feed on *Centaurea procurrens* Spreng, *Carlina curetum* Halácsy and *Centaurea hyalolepis* Boiss. (new host plant records).

Distribution. Europe to Ural Mountains, Algeria, Turkey, Syria, Lebanon, Caucasus, Central Asia to China (Xinjiang) (Englert 1974; Elsner *et al.* 1999; Li 2002: 107; Junnilainen *et al.* 2010: 21), Israel (new record).

Metzneria freidbergi sp. nov.

Type material. Holotype ♂, Israel, Mt. Hermon, 2000 m, 22.v.1973 (Freidberg) (gen. slide 134/18, O. Bidzilya) (TAU). Paratypes: 1 ♂, same data as holotype; 2 ♂, 2 ex (abdomen missing), same data as holotype but ex. *Cousinia hermonis*, 1.vi. [1973] (gen. slide 137/18, O. Bidzilya) (TAU, ZMKU).

Diagnosis. Light brown forewing with extensive grey pattern along veins and with three distinct black markings are characteristic for the new species. Other species of *Metzneria* lack distinct black markings (*M. lappella* Zeller, 1839, *M. neuropterella* Zeller, 1839) or are smaller in size (*M. santolinella* (Amsel, 1936), *M. intestinella* (Mann, 1864) and others). The male genitalia are recognized by two long and two short cornuti in the phallus in combination with the rounded sacculus. *M. ehikeella* and *M. diamondi* Amsel, 1949 are similar in the shape of valva and sacculus but have only two long cornuti in the phallus. *M. lappella* (Linnaeus, 1758) has usually 4-5 cornuti of subequal length.

Description. Adult (Fig. 33). Wingspan 21.8-23.0 mm. Head and labial palpus yellow-brown, segment 2 mottled with grey on outer side, segment 3 with dark brown medial ring, scape light brown, antennal segments ringed light brown and dark; thorax light brown with dark brown medial line; ground colour of forewing light brown, veins, fold, termen and costal and dorsal margins mottled with grey, black dot in the end of fold near dorsal margin, another black touch in middle and black spot in the corner of cell, cilia grey brown-tipped; hindwing grey.

Male genitalia (Figs 34, 35). Uncus short, broadly rounded; gnathos absent; tegumen trapezoid, anterior margin twice as long as posterior margin; valva narrow, broadened at 3/4 length, dorso-caudal corner rounded, inner margin nearly straight, outer margin curved, apex with pointed tip; sacculus rounded, narrowed apically; saccus triangular; phallus about three times as long as broad, apex elongated, pointed or truncate distally, laterally with sclerotized plate covered with spines, vesica with two long thorn-shaped and two short needle-shaped cornuti.

Female genitalia. Unknown.

Biology. Larvae feed on *Cousinia libanotis* DC. (as *C. hermonis*). Adults were recorded from late May to early June at an altitude of 2000 m.

Distribution. Israel: Hermon Mountain.

Etymology. The new species is named after the collector of the type series, Dr. Amnon Freidberg, a well-known Israeli dipterist and enthusiastic collector of insects all over the world, curator of insect collection during many years in the Steinhardt Museum of Natural History, Israel National Center for Biodiversity Studies and Department of Zoology, Tel Aviv University.

Metzneria lappella (Linnaeus, 1758)

Phalaena (Tinea) lappella Linnaeus, 1758—Syst. Nat. (Edn 10) 1: 537.

Records. Bodenheimer 1930a: 173; 1937: 100.

Distribution. Europe, Asian part of Russia (Kemerevo region, Altai, Tuva, Zabaikalskiy krai), Turkey, Middle East, Armenia, Middle Asia, Kazakhstan, North America (introduced) (Elsner *et al.* 1999: 25; Bidzilya 2002: 67).

***Metzneria littorella* (Douglas, 1850)**

Gelechia littorella Douglas, 1850—Trans. ent. Soc. Lond. (n.s.) 1: 67.

Metzneria quinquepunctella Herrich-Schäffer, 1855—Syst. Bearb. Schmett. Europ. 5 (65): 172, (62) fig. 573.

Records. Englert 1974: 398.

Material examined. 1 ex., Südende d. Toten Meers, 15-27.iii.1933 (Amsel) (SMNK).

Distribution. England, South Europe to Lower Volga and West Caucasus, North Africa, Israel (Englert 1974: 398; Ponomarenko 2008: 88).

***Metzneria tenuiella* (Mann, 1864)**

Gelechia tenuiella Mann, 1864—Wien. Ent. Mon. 8 (6): 186, pl. 4, fig. 16.

Apodia seminivora Walsingham, 1903—Ent. Mon. Mag. 39: 263.

Metzneria insignificans Walsingham, [1908]—Proc. Zool. Soc. Lond. 1907: 926.

Metzneria infelix Walsingham, [1908]—Proc. Zool. Soc. Lond. 1907: 926.

Records. Amsel 1933: 124 (*infelix*); Amsel 1935c: 262 (*infelix*); Bodenheimer 1937: 100 (*infelix*); Englert 1974: 392.

Material examined. 1 ♂, Jerusalem, Palästina, 3.vi.1930 (Amsel) (gen. slide GU 148, Englert) (SMNK).

Distribution. Canary Islands, South Europe, Turkey, Israel (Englert 1974: 392).

***Spiniphallellus fuscescens* Bidzilya & Karsholt, 2008**

Spiniphallellus fuscescens Bidzilya & Karsholt, 2008—Nota lepid. 31(2): 208.

Material examined. 1 ♂, Golan Heights, vi.2001 (Kravchenko) (gen. slide 113/18, O. Bidzilya) (TAU).

Distribution. Turkey (Bidzilya & Karsholt 2008), Israel (new record).

***Bryotropha domestica* (Haworth, 1828)**

Recurvaria domestica Haworth, 1828—Lepid. Britannica (4): 551.

Gelechia domesticella Doubleday, 1859—Zool. Syn. List Br. Butts Moths (edn 2): 30.

Lita (?) punctata Staudinger, 1876—Stettin. ent. Ztg 37: 146.

Gelechia domestica var. *salmonis* Walsingham, [1908]—Proc. Zool. Soc. Lond. 1907: 937.

Bryotropha algericella Chrétien, 1917—Ann. Soc. Ent. Fr. 85: 469.

Records. Bodenheimer 1930a: 173; Amsel 1933: 125; Amsel 1935c: 263; Bodenheimer 1937: 100; Karsholt & Rutten 2005: 100.

Material examined. 1 ♀, Haifa, 25.v.1921 (Buxton); 1 ♀, Jerusalem, 5.v.1922 (Buxton) (all NHMUK); 1 ex., Jerusalem, Abu-Gosch, 28.vi.1931 (Einsler) (SMNK); 1 ex., Ain Karim, Jerusalem, 01.vi.1930 (Amsel); 1 ex., Georgskloster, Wadi el Kelt, 1.iv.1930 (all NHMW); 1 ♀, Michmoret, 26.viii.1968 (gen. slide 165/18, O. Bidzilya) (TAU); 1 ♀, slopes by Yflach, 31.v.2004 (Kravchenko); 3 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (all TAU); 5 ♂, 3 ♀, E bank of Kenneret lake, Mevo Hama, 18.iv.2018, light (Kravchenko & Bidzilya) (TAU, ZMKU).

Distribution. Europe eastwards to Hungary and Slovakia, Canary Islands, North Africa, Middle East, Saudi Arabia, Yemen, Turkmenistan (Karsholt & Rutten 2005: 99–100).

***Bryotropha arabica* Amsel, 1952**

Bryotropha arabica Amsel, 1952—Fragm. ent. Roma 1: 120.

Records. Amsel 1933: 125 (*terrella*); Amsel 1935c: 263 (*terrella*); Amsel 1952: 120; Bodenheimer 1937: 100 (*terrella*); Karsholt & Rütten 2005: 105.

Material examined. Lectotype of *Bryotropha arabica* ♂, Georgskloster, Wadi el Kelt, Lichtfang, 1.iv.1930 | Palästina. Expedition, 19.ii-4.vi.1930 (Amsel) | GU 182 | Lectotype of *Bryotropha arabica* Amsel, 1952, O. Karsholt design., 1999 (SMNK); 5 ♂, Kirjat Anawim, Jerusalem, 28.iii, 22,28,29.iv.1930 (Amsel); 1 ♂, 1 ♀, Jerusalem, 9.iv.1930 (Amsel); 1 ♂, 1 ♀, Georgskloster, Wadi el Kelt, 1.iv.1930 (Amsel) (all SMNK); 1 ♂, Nahal Zin, v.2000; 1 m, Nahal HaKikkar, v.1999; 1 ♀, Huloth, 20.iv.1968 (Shoham) (all TAU).

Distribution. South Europe, North Africa, Turkey, Middle East (Lebanon, Syria, Israel), Saudi Arabia, Yemen, Iraq, Iran, Turkmenistan (Karsholt & Rütten 2005: 105).

***Bryotropha hulli* Karsholt & Rütten, 2005**

Bryotropha hulli Karsholt & Rütten, 2005—Tijdschr. Ent. 148 (1): 131, figs 87–91, 201–202, 254, 298–299, 345–346.

Records. Karsholt & Rütten 2005: 131.

Material examined. Paratypes of *hulli* ♂, Karmel, Hiafa, 7.v.1930 (gen. slide A. 834); 1 ♂, Ain Karim, Jerusalem (Pal.), 1.vi.1930 (Amsel) (gen. slide A. 833), 1 ♀, same place, 14.v.1930 (Amsel) (gen. slide AR0460) (SMNK); 1 ♂, Ein Gedi, 17.viii.1968 (gen. slide 203/18, O. Bidzilya) (TAU); 1 ♀, East bank of Kenneret lake, Mevo Hama, 18.iv.2018, light (Bidzilya & Kravchenko) (gen. slide 132/18, O. Bidzilya) (ZMKU).

Distribution. Croatia, Macedonia, Bulgaria, Greece, Cyprus, Turkey, Syria, Israel (Karsholt & Rütten 2005: 133).

***Bryotropha plebejella* (Zeller, 1847)**

Gelechia plebejella Zeller, 1847—Isis von Oken 1847 (11): 850.

Gelechia imperitella Staudinger, 1859—Stettin ent. Ztg 20 (7–9): 242.

Aristotelia ancillula Walsingham, [1908]—Proc. Zool. Soc. Lond. 1907: 930.

Bryotropha inexpectella Nel, 1999—Bull. Soc. ent. Fr. 104: 348.

Records. Bodenheimer 1930a: 173; Amsel 1933: 125; Amsel 1935c: 263; Bodenheimer 1937: 100; Halperin & Sauter 1992: 134 (cf. *plebejella*); Karsholt & Rütten 2005: 105.

Material examined. 3 ♀, Jerusalem, Palästina (Amsel), 8.iv.1930 (gen. prep. 2334, H. Hendriksen; slide No. AR0330, L.M. Rütten); 1 ♂, Kirjat, Anawim, Jerusalem, 2.v.1930 (gen. slide No. AR0275, A.L.M. Rütten); 2 ♀, Jerusalem, 9.iv.1930 (Amsel) (gen. slide HH2334; AR0330) (all SMNK); 2 ♀, Jerusalem, 6.vi.2004 (Kravchenko) (TAU).

Host plant. Halperin & Sauter (*op cit.*—as “*Bryotropha cf. plebejella* Z.”) recorded larvae feeding on twigs of *Quercus ithaburensis* Decne. and green twigs of *Tamarix aphylla* (L.) H. Karst. infested with *Waxiella tamaricis*. We consider these records as doubtful because larvae of *Bryotropha* species feed on mosses (Bryophyta) (Karsholt & Rütten 2005: 85).

Distribution. Madeira and Canary Islands, South Europe, North Africa, Turkey, Syria, Israel (Karsholt & Rütten 2005: 134).

***Bryotropha affinis* (Haworth, 1828)**

Recurvaria affinis Haworth, 1828—Lepid. Britannica (4): 551.

Gelechia tegulella Herrich-Schäffer, 1854—Syst. Bearb. Schmett. Europ. 5 (65): 182.

Gelechia tectella Herrich-Schäffer, 1854—Syst. Bearb. Schmett. Europ. 5 (65): 182.

Gelechia affinella [= *affinis*] Doubleday, 1859—Zool. Syn. List Br. Butts Moths (edn 2): 30 (emend.).

Anacampsoides affinitella [= *affinis*] Bruand, 1859—Ann. Soc. ent. Fr. (3) 6: 643 (emend.).

Records. Amsel 1933: 125; Bodenheimer 1937: 100.

Material examined. 1 ♀, slopes by Yflach, 31.v.2004 (Kravchenko) (TAU).

Distribution. Europe (Karsholt & Rutten 2005: 139), Israel.

***Ornativulva heluanensis* (Debski, 1913)**

Teleia heluanensis Debski, 1913—Bull. Soc. ent. Egypte 3: 111.

Teleia frankeniivorella Chrétien, 1917—Ann. Soc. Ent. Fr. 85: 474.

Teleja [sic] *oasicolella* Turati, 1924—Atti Soc. ital. Sci. nat. 63: 161, pl. 6, fig. 4.

Lita siculella Mariani, 1937—G. Sci. nat. econ. Palermo 39 (3): 9, pl. 1, fig. 1.

Records. Sattler, 1967: 39; Sattler 1976: 103.

Material examined. 2 ♂, Südende d. Toten Meers, 15-27.iii.1933 (Amsel); 2 ♂, Jordanbrücke, Jericho, 10,29. iv.1930 (Amsel) (gen. slide GU 3800♂, Genitalia K. Sattler, 303a; GU 3799♂, Genitalia K. Sattler, 299b) (all SMNK); 3 ♂, 4 ♀, Ein Gedi, 29, 30.iii.1968 (TAU); 2 ♂, 1 ♀, 18 exx, Jericho, Kalia, v.2004 (Kravchenko) (gen. slide 5489 Hendriksen) (TAU, ZMUC).

Distribution. South Europe to Lower Volga, Cape Verde, Canary Islands, North Africa, Middle East, Central Asia to Mongolia and West China (Quinghai) (Sattler 1976: 103; Emelyanov & Piskunov 1982: 391).

***Ornativulva zonella* (Chrétien, 1917)**

Teleia zonella Chrétien, 1917—Ann. Soc. Ent. Fr. 85: 474.

Teleia cimelion Amsel, 1935—Veröff. dt. Kol. Übersee-Mus. 1: 210.

Ornativulva iranella Sattler, 1967—Beitr. naturk. Forsch. Südwdtl. 26 (3): 44, pl. 3, fig. 19.

Records. Amsel 1935b: 210 (*cimelion*); Bodenheimer 1937: 101 (*cimelion*); Sattler 1967:45; Sattler 1976: 106.

Material examined. *Teleia cimelion* “Typus”, ♀, Südende d. Toten Meers, 15-27.iii.1933 (Aigner); 1♂, “Paraty-
pus”, same data (gen. slide GU 3997♂, Genitalia K. Sattler, 297a) (all SMNK); 3 ♂, 1 ♀, Hazeva, Field School, 28.iii, 10.iv, 4,18.v.1998, Malaise Trap (Ashkenazi) (gen. slide 162/18♂, 158/18♀, O. Bidzilya) (TAU).

Distribution. Algeria, Tunisia, Saudi Arabia, Israel, South Iran, China (Xinjiang) (Sattler 1976: 106; Li 2002: 91).

***Ornativulva erubescens* (Walsingham, 1904)**

Gelechia erubescens Walsingham, 1904—Ent. mon. Mag. 40: 265.

Records. Sattler 1967:46; Sattler 1976: 108.

Material examined. 1 ♀, Field School, 24.viii.1997, Malaise Trap (Ashkenazi) (gen. slide 151/18, O. Bidzilya); 1 ♂, Hazeva, Field School, 21.iv.1998, Malaise Trap (Ashkenazi) (gen. slide 154/18, O. Bidzilya) (all TAU); 3 ♂, 3 ♀, 77 exx, Jericho, Kalia, v.2004 (Kravchenko) (gen. slide 5490 Hendriksen) (TAU, ZMUC); 4 ♂, 8 ♀, Nahal (Wadi) Zin, black light, 8.x.1999 (Pavliček) (NMPC).

Distribution. North Africa, Saudi Arabia, Israel, Iran, Uzbekistan, Pakistan (Sattler 1976: 47; Falkovitsh & Bidzilya 2009: 70).

***Ornativulva arabica* Sattler, 1967**

Ornativulva arabica Sattler, 1967—Beitr. naturk. Forsch. Südwdtl. 26 (3): 48, pl. 4, fig. 25 [not 24], pl. 10, fig. 51.

Material examined. 2 ♂, 1 ♀, Hazeva, Field School, 22, 28.iii, 6.iv.1998, Malaise Trap (Ashkenazi) (gen. slide 140/18♀, 145/18♂, O. Bidzilya) (all TAU); 1 ♀, Nahal (Wadi) Zin, black light, 8.x.1999 (Pavliček) (gen. slide 19002, J. Šumpich) (NMPC).

Distribution. Sudan, Saudi Arabia (Sattler 1976: 115), Israel (new record).

Ornativalka sesostrella (Rebel, 1912)

Gelechia sesostrella Rebel, 1912—Dt. ent. Z. Iris 26 (1): 88.

Material examined. 1 ♂, 1 ♀, Hazeva, Field School, , 23.v.1998, Malaise Trap (Ashkenazi) (gen. slide 146/18♀, 155/18♂, O. Bidzilya); 1 ♂, Hazeva, Field School, 8.x.1997, Malaise Trap (Ashkenazi) (gen. slide 138/18 O. Bidzilya); 1 ♀, Nahal Shezaf, 30.iii.1998 (Ashkenazi) (gen. slide 157/18, O. Bidzilya); 1 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (all TAU).

Distribution. Algeria, Tunisia, Egypt, Sudan, Saudi Arabia, South Iran, Pakistan (Sattler 1976), Israel (new record).

Ornativalka misma Sattler, 1976

Ornativalka misma Sattler, 1976—Bull. Br. Mus. nat. Hist. (Ent.) 34 (2): 121, 100 (list), pl. 1, fig. 6, pl. 3, fig. 23, pl. 17, fig. 94, pl. 25, fig. 117.

Records. Halperin & Sauter 1992: 135.

Host plant. In Israel the larvae have been recorded feeding on green twigs of *Tamarix* sp. (Halperin & Sauter 1992: 135).

Distribution. Sudan (Sattler 1976: 122), Israel.

Ornativalka macrosignella Sattler, 1967

Ornativalka macrosignella Sattler, 1967—Beitr. naturk. Forsch. Südwdtl. 26 (3): 60, pl. 5, fig. 29, pl. 11, fig. 55.

Records. Amsel 1933: 125 (*tamaricalis* Z. Misspelling and misidentification); Amsel 1935c: 263 (partim) (*tamariciella*; misidentification); Bodenheimer 1937: 101 (*tamariciella*); Sattler 1967:61; Sattler 1976: 124.

Material examined. 1 ♂, Jordanbrucke bei Jericho, 29.iv.1930, Lichtfn. (Amsel); 1 ex (abdomen missing), Sü-dende des Toten Meeres, 15-27.iii.1933 (Amsel) (all SMNK); 1 ♂, 5 ♀, Jericho, Kalia, v.2004 (Kravchenko) (gen. slide 5487 Hendriksen) (TAU, ZMUC).

Distribution. Algeria, Tunisia, Egypt, Saudi Arabia, Sudan, Israel, South Iran, Uzbekistan, Turkmenistan (Amsel 1958: 81; Sattler 1976: 124; Falkovitsh & Bidzilya 2009: 69).

Ornativalka antipyramis (Meyrick, 1925)

Gelechia antipyramis Meyrick, 1925—Bull. Soc. ent. Egypte 9 (1–3): 209.

Material examined. 1 ♀, Hazeva, Field School, 16.v.1998, Malaise Trap (Ashkenazi) (gen. slide 202/18, O. Bidzilya); 1 ♂, Jericho, Kalia, v.2004 (Kravchenko) (all TAU); 3 ♂, 1 ♀, Nahal (Wadi) Zin, black light, 8.x.1999 (Pavliček) (gen. slide 19001, J. Šumpich) (NMPC).

Distribution. Cape Verde, Canary Islands, Morocco, Algeria, Tunisia, Libya, Egypt, Sudan, Socotra, Jordan, Saudi Arabia, United Arab Emirates, South Iran, Pakistan (Sattler 1976: 129), Israel (new record).

***Ornativulva plutelliformis* (Staudinger, 1859)**

Gelechia plutelliformis Staudinger, 1859—Stettin ent. Ztg 20 (7–9): 239.

Alucita olbiaella Millière, [1861]—Icon. Desc. Chenilles Lepid. 1: 193, pl. 21, fig. 1–6.

Hypsolophus siewersiellus Christoph, 1867—Stettin Ent. Ztg 28 (4–6): 239.

Gelechia sinuatella Walsingham, 1904—Ent. mon. Mag. 40: 223.

Records. Amsel 1933: 125; Amsel 1935b: 210; Bodenheimer 1937: 100; Sattler 1967:72.

Material examined. 2 ♀, Südde d. Toten Meers, 15-27.iii.1933 (Amsel); 1 ♀, Jordan valley, Zerqua R. Colny, c. 100 m below S.L., at light (Trevor Trought); 1 ♀, Jordanbrücke, Jericho, 31.v.1930 (Amsel) (all SMNK); 6 ♂, 15 ♀, Hazeva, Field School, 24.ii, 8,21,26.iii, 20, 21, 23, 26.iv, 4,9,17,18.v.1998 (Ashkenazi); 3 ♀, Hazeva, Field School, 7.i, 16.vii, 24.viii.1997 (Maklakov); 1 ♀, En-Zin, 17.i.1999 (Yarom & Kravchenko); 1 ♀, Wingeit, 26.viii.1968; 1 Ein Gedi, 16.viii.1968 (all TAU); 3 ♂, 2 ♀, 103 exx, Jericho, Kalia, v.2004 (Kravchenko) (gen. slide 5490 Hendriksen) (TAU, ZMUC).

Host plant. In Israel the larvae were recorded feeding on *Tamarix* sp. (Bodenheimer, 1937: 100).

Distribution. South Europe to Lower Volga, Canary Islands, North Africa, Middle East (Israel, Jordan, Lebanon, Syria), Saudi Arabia, Iran, Iraq, Afghanistan, Pakistan, Turkmenistan, Uzbekistan, China (Xinjiang) (Sattler 1976: 132; Li 2002: 84; Falkovitsh & Bidzilya 2009: 69).

***Ornativulva mixolitha* (Meyrick, 1918)**

Phthorimaea mixolitha Meyrick, 1918—Exotic Microlep. 2 (5): 135.

Ornativulva mixolitha (Meyrick, 1918)—Sattler, 1976: 135.

Material examined. 1 ex, Südde d. Toten Meers, 15-27.iii.1933 (Amsel) (gen. slide Am. 1645) (SMNK).

Distribution. North Africa, Russia (Volgograd region), Turkey, Israel (new record), Iran, Iraq, Afghanistan, Uzbekistan, Pakistan, India (Bihar), China (Xinjiang), Mongolia (Sattler 1976: 136; Li 2002: 78; Falkovitsh & Bidzilya 2009: 70).

***Ornativulva triangulella* Sattler, 1967**

Ornativulva triangulella Sattler, 1967—Beitr. naturk. Forsch. SüdwDtl. 26 (3): 68, pl. 6, fig. 33, pl. 13, fig. 60, 60a.

Material examined. 2 ♂, 1 ♀, Jericho, Kalia, v.2004 (Kravchenko) (gen. slide 5488 Hendriksen) (TAU, ZMUC).

Distribution. Algeria, Tunisia, Kuwait, Iraq, Iran, Afghanistan (Sattler 1976: 128), Israel (new record).

***Deltophora fasciella* Sattler, 1979**

Deltophora fasciella Sattler, 1979—Bull. Br. Mus. nat. Hist. (Ent.) 38 (6): 282, 276 (list).

Records. Amsel 1935c: 263 (*maculata*; misidentification, partim); Sattler 1979: 283.

Material examined. Paratype of *Deltophora fasciella* ♀, Georgskloster, Wadi el Kelt, 15.iv.1930 | Palästina Expedition, 19.ii-4.vi.1930 (Amsel) | Prep. Nr. 5554♀ | Genitalia slide K. Sattler 632c | Paratype *Deltophora fasciella* Sattler, ♀, teste K. Sattler, 1977 (SMNK).

Distribution. Israel, Saudi Arabia, Sudan (?), Iran, Afghanistan (Sattler 1979: 283).

***Deltophora maculata* (Staudinger, 1879)**

Teleia maculata Staudinger, 1879—Horae Soc. Ent. Ross. 15 (2–3): 314.

Records. Amsel 1935c: 263 (partim); Bodenheimer 1937: 101; Sattler 1979: 287.

Material examined. 1 ♀, Kirjat Anawim, Jerusalem, 22.iv.1930 (gen. slide GU 3726♀); 1 ♂, Ramallah, Jerusalem, 21.iv.1930 (Amsel) (SMNK); 1 ♀, Ein Gedi, 17.viii.1965 (TAU).

Distribution. Spain, France, Italy (Karsholt & Riedl 1996: 108), Ukraine (Crimea) (Bidzilya *et al.* 2003: 67), Greece (including Crete), Turkey, Syria, Lebanon, Israel, Georgia, Armenia, Azerbaijan, Turkmenistan, Afghanistan, Iran (Sattler 1979: 286, 287).

Subfamily Gelechiinae Stainton, 1854

Tribe Gelechiini Stainton, 1854

Mirificarma aflavella (Amsel, 1935)

Fig. 45

Rhinosia flavella aflavella Amsel, 1935—Veröff. dt. Kol. Übersee-Mus. 1 (2): 275.

Records. Amsel 1935c: 275; Amsel 1936: Pl. 15, fig. 33, 35; Bodenheimer 1937: 101; Pitkin 1984: 25.

Material examined. Lectotype of *Rhinosia flavella aflavella* ♂, Tabgha, Tiberias, März 1930 (Amsel) | Lectotype *Rhinosia flavella aflavella* Amsel, det L.M. Pitkin, 1982 | L.M.P. Genitalia Slide No. 17, ♂; paratype 1 ♂, 2 ♀, same data as for holotype (all SMNK).

Distribution. Greece, Turkey, Israel (Pitkin 1984: 26).

Mirificarma flavella (Duponchel, 1844)

Acompsia flavella Duponchel, 1844—in Godart, Hist. Nat. Lépid. Fr. (Suppl.) 43: 512, pl. 89, fig. 7.

Gelechia segetella Zeller, 1847—Isis von Oken 1847 (11): 847.

Records. Stainton 1867: 40; Caradja 1920: 112; Amsel 1933: 126; Amsel 1935c: 264.

Material examined. 1 ♂, Hulioth, 19.iv.1968 (TAU).

Distribution. South Europe, North Africa, Turkey, Israel (Pitkin 1984: 27; Huemer & Karsholt 1999: 129).

Mirificarma eburnella (Denis & Schiffermüller, 1775)

Tinea eburnella Denis & Schiffermüller, 1775—Ank. syst. Schmett. Wienergegend: 140.

Tinea formosella Hübner, 1796—Samml. eur. Schmett. [8]: 62, pl. 23, fig. 160 (preocc. *Tinea formosella* Denis & Schiffermüller, 1775).

Carcina flammella Hübner, [1825]—Verz. Bek. Schmett. (23-27): 410 (repl. *Tinea formosella* Hübner, 1796).

Gelechia rufeoformosella Bruand, 1859—Ann. Soc. ent. Fr. (3) 6: 652 (repl. *Tinea formosella* Hübner, 1796).

Records. Stainton 1867: 40 (*formosella*); Amsel 1933: 126 (*formosella*); Amsel 1935c: 264 (*formosella*); Bodenheimer 1937: 101 (*formosella*).

Material examined. 4 ♂, Hulioth, 9, 19, 20, 26.iv.1968 (TAU); 1 ♀, East bank of Kenneret lake, Mevo Hama, 18.iv.2018, light (Kravchenko & Bidzilya) (ZMKU).

Distribution. Europe, North Africa, Turkey, Lebanon, Syria, Israel, Armenia, USA (presumably introduced) (Pitkin 1984: 28).

***Mirificarma mulinella* (Zeller, 1839)**

Gelechia mulinella Zeller, 1839—Isis [32] 1839 (3): 199.
Gelechia caminariella Fuchs, 1902—Stettin ent. Ztg 63 (1): 323.
Gelechia nigraesilvae Amsel, 1950—Beitr. nat. Forsch. SüdWdtl. 9: 27.

Material examined. 2 ♂, Nahal Kziv, light trap, 5.xii.1998 (Pavliček & Kravchenko) (gen. slide 19017, J. Šumpich); 2 ♂, ditto but 13.xi.1998; 1 ♂, Nahal Oren, Mt. Carmel, light trap 20.xi.1997 (Pavliček & Kravchenko); 4 ♀, ditto but 15.xi.1998 (all NMPC).

Distribution. Europe, North Africa, Greece (Crete), Cyprus, Zabaikalskiy krai of Russia (Huemer & Karsholt 1999: 137; Ponomarenko 2008), Israel (new record).

***Mirificarma ocellinella* (Chrétien, 1915)**

Gelechia ocellinella Chrétien, 1915—Ann. Soc. ent. Fr. 84: 317.

Material examined. 1 ♀, En-Zin, 17.i.1999, light trap (Yarom & Kravchenko) (gen. slide 116/18, O. Bidzilya) (TAU); 1 ♀, Negev centr., Nahal Haro'a, black light, 15.i.2000 (Pavliček) (gen. slide 19008, J. Šumpich) (NMPC).

Distribution. Morocco, Algeria, Tunisia, Libya, Jordan (Pitkin 1984: 30), Israel (new record).

***Mirificarma rhodoptera* (Mann, 1866)**

Gelechia rhodoptera Mann, 1866—Verh. zool. bot. Ges. Wien 16: 353, pl. 1, fig. 10.

Records. Halperin & Sauter 1992: 126, 134.

Host plant. In Israel the larvae have been recorded living in green twigs of *Spartium junceum* L. (Halperin & Sauter 1992: 134).

Distribution. Romania, Greece, Turkey, Lebanon (Karsholt & Huemer 1999: 126), Israel.

Remarks. Without studying the material of *M. rhodoptera* from Israel we cannot exclude that this record belongs to closely related *M. minimella* Karsholt & Huemer, 2001 known from Greece.

***Athrips fagoniae* (Walsingham, 1904)**

Leobatus fagoniae Walsingham, 1904—Ent. mon. Mag. 40: 221.
Lita cervinella Turati, 1934—Atti Soc. Ital. Sci. Nat. 73: 197.
Gelechia rosinansella Lucas, 1942—Bull. Soc. ent. Fr. 47: 125.

Material examined. 1 ♂, 'En Gedi, 5.ii.2003 (Weinstein); 1 ♀, En-Zin, 17.i.1999 (Yarom & Kravchenko) (TAU).

Distribution. Canary Islands, Tunisia, Algeria, Libya, Egypt, Jordan (Amsel 1955b: 279; Sattler 1968: 113; Bidzilya 2005: 36), Israel (new record).

***Athrips rancidella* (Herrich-Schäffer, 1854)**

Gelechia rancidella Herrich-Schäffer, 1854—Syst. Bearb. Schmett. Europ. 5 (65): 176, (58) fig. 534.
Gelechia triatoma Mühlig, 1864—Stettin Ent. Ztg 25 (1–3): 101.
Gelechia vepretella Zeller, 1871—Stettin Ent. Ztg 32 (1–3): 64.
Gelechia superfetella Peyerimhoff, 1877—Pet. Nouv. Ent. 9 (164): 102.
Cremona cotoneastri Busck, 1934—Proc. Ent. Soc. Wash. 36 (4): 83, pl. 14, fig. 1–5.
Epithectis cerasivorella Kuznetsov, 1960—Trudy zool. Inst. Leningr. 27: 34.
Gelechia haifella Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 300. **Syn. nov.**

Records. Bodenheimer 1930a: 173 (*vepretella*); Bodenheimer 1930b: 239, 249 (*vepretella*); Amsel 1933: 125 (*haifella*); Amsel 1935a: 300 (*haifella*); Amsel 1935c: 263 (*vepretella*); Bodenheimer 1937: 100 (*haifella* & *vepretella*); Bytinski-Salz 1966: 34.

Material examined. Holotype of *Gelechia haifella* ♀ “Karmel, Haifa, 7.5.1930, H. Amsel” | *Gelechia haifella*, Typus, leg. H. Amsel” (gen. slide 251/08♀, O. Bidzilya) (SMNK); 1 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (TAU).

Host plant. In Israel larvae were reported from *Prunus* sp. and *Pyrus* sp. (Bodenheimer 1930a: 173; 1930b: 239, 249).

Distribution. Europe, Transcaucasia, Middle East, Iran, Syria, Turkmenistan, USA (Bidzilya 2005: 50), Israel. *A. rancidella tadzhika* Bidzilya, 2005 from Tadjikistan should probably be considered as a separate species according to data from DNA analysis (Karsholt & Huemer 2017).

Remarks. *Gelechia haifella* was described from a single female collected on Mount Carmel near Haifa. The holotype (Fig. 46) agrees both externally and in the genitalia with *Athrips rancidella*. Hence, the following synonymy is established: *Gelechia haifella* Amsel, 1935 **syn. nov.** of *Athrips rancidella* (Herrich-Schäffer, 1854).

Aroga aristotelis (Millière, [1876])

Gelechia aristotelis Millière, 1876—Cat. Léop. Alpes-Maritimes (3): 324, 414, pl. 1, f. 1-2

Gelechia astragali Staudinger, 1879—Horae Soc. Ent. Ross. 15 (2-3): 304.

Gelechia fulminella Millière, 1883—Ann. Soc. Linn. Lyon (n.s.) 29: 161, pl. 2, fig. 4.

Gelechia lacertella Walsingham, 1904—Ent. mon. Mag. 40: 222.

Gelechia aplasticella Rebel, 1913—Ann. Mus. Wien. 27: 330 (nom. nud.).

Gelechia hyrcanella Toll, 1947—Z. wien. ent. Ges. 32: 113, pl. 4, fig. 20.

Material examined. 2 ♂, Golan Heights, Katzrin, Sept. 2003 (Kravchenko) (TAU).

Distribution. Mediterranean area, from Spain to Turkey, Ural Mnts, Iran (Huemer & Karsholt 1999: 159), Israel (new record).

Tribe Gnorimoschemini Povolný, 1964

Scrobipalpa aptatella (Walker, 1864)

Gelechia aptatella Walker, 1864—List Spec. Lepid. Insects Colln Br. Mus. 29: 636.

Gelechia heliopa Lower, 1900—Proc. Linn. Soc. N.S.W. 25 (3): 417.

Records. Bodenheimer 1930a: 174 (*heliopa*); Bodenheimer 1930b: 314 (*heliopa*); Amsel 1933: 125 (*heliopa*); Amsel 1935c: 263 (*heliopa*); Bodenheimer 1937: 101 (*heliopa*); Bytinski-Salz 1966: 34 (*heliopa*).

Host plant. In Israel larvae feed in stem galls on *Nicotiana tabacum* L. (Bodenheimer 1930a: 174; 1930b: 314–315).

Distribution. East Australia. Introduced into Oriental and Afrotropical regions. Records from Europe (France, Italy, former Yugoslavia) are unconfirmed (Huemer & Karsholt 2010: 59).

Remarks. We did not examine material of this species, but accept it on the basis of the illustrations and text by Bodenheimer (1930b: 314), based on galls with larvae found at Tel Josef, Dagania.

Scrobipalpa ocellatella (Boyd, 1858)

Gelechia ocellatella Boyd, 1858—Ent. Weekly Int. 4 (96): 143.

Gelechia ocellatella Stainton, 1859—Ann. Mag. nat. Hist (3) 3: 212 (preocc.).

Gelechia submissella Stainton, 1859—Ann. Mag. nat. Hist (3) 3: 212.

Gelechia horticolella Rössler, 1866—Verz. Schmett. Nassau: 240.

Lita atriplicella var. *clarella* Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 100.

Lita ocellatella obscurior Rebel, 1926—Bull. Soc. ent. Egypte 10: 187.

Gnorimoschema ocellatellum orientale Gregor & Povolný, 1954—Zool. ent. Listy 3: 91.

Records. Bodenheimer 1930a: 174; Bodenheimer 1930b: 338–339; Amsel 1933: 125; Amsel 1935b: 211; Amsel 1935c: 263; Bodenheimer 1937: 100; Kugler 1966: 85.

Material examined. 1 ♂, Bat Shlomo, 20.viii.1968; 2 ♂, Hulioth, 17.x.1968 (Shohom); 1 ♂, 1 ♀, Ein Gedi, 16.viii.1968; 1 ♀, Abu Gosh, 21.viii.1968; 1 ♀, Vingeit, 26.viii.1968; 2 ♀, Bat Zion, 28.viii.1968 (gen. slide 200/18, O. Bidzilya) (all TAU).

Host plant. In Israel the larvae were recorded feeding on *Beta* sp. (Bodenheimer 1930a: 174; 1930: 338–339; 1937: 100).

Distribution. Europe, North Africa, Turkey, Middle East, Armenia, Turkmenistan (Lvovsky & Piskunov 1999: 84; Huemer & Karsholt 2010: 165).

***Scrobipalpa gecko* (Walsingham, 1911)**

Lita gecko Walsingham, 1911—Ent. mon. Mag. 47: 12.

Records. Povolný 2002: 44.

Material examined. 1 ♂, 1 ♀, Hazeva, Field School, 20.iv, 4,11,16,17,18.v.1998, Malaise Trap (Ashkenazi) (gen. slide 126/18♂, O. Bidzilya); 1 ♀, Nahal Gidron, 22.xi.1999, Malaise Trap (Yarom & Zeevi); 1 ♀, Nahal Shahaq, Shesaf Nat. Rez., 5.xi.1999, Malaise Trap (Yarom & Zeevi) (all TAU).

Distribution. Greece, Algeria, Tunisia, Israel (Povolný 2002: 44; Huemer & Karsholt 2010: 168).

***Scrobipalpa erichi* Povolný, 1964**

Scrobipalpa erichi Povolný, 1964—Acta Soc. ent. Cechoslov. 61: 356.

Records. Povolný 1964: 356.

Material examined. Paratype of *Scrobipalpa erichi* 1 ex., Jordanbrücke bei Jericho, 10.iv.1930, Lichtfang (Amsel) (gen. slide A. 809); 1 ♀, Jordanbrücke bei Jericho, 29.iv.1930, Lichtfang (Amsel) (gen. slide A. 800) (all SMNK); 1 ♀, Tel Aviv, Botanical garden, 15.iv.2018, light (Bidzilya & Kravchenko) (TAU, ZMKU).

Distribution. South of Central and Eastern Europe, Turkey, Middle East, Central Asia to Mongolia and China (Xinjiang, Inner Mongolia) (Bidzilya & Li 2010: 4; Karsholt & Huemer 2010: 170).

***Scrobipalpa portosanctana* (Stainton, 1859)**

Gelechia portosanctana Stainton, 1859—Ann. Mag. nat. Hist (3) 3: 212.

Gelechia eremaula Meyrick, 1891—Ent. mon. Mag. 27: 57.

Gelechia lyciella Walsingham, 1900—Ent. mon. Mag. 36: 217.

Lita desertella Rebel, 1901—Dt. ent. Z. Iris 13 (2): 165.

Phthorimaea bertramella Lucas, 1940—Bull. Soc. ent. Fr. 44: 228.

Teleia leroyella Lucas, 1949—Bull. Soc. ent. Fr. 54: 143.

Gnorimoschema reisseri Povolný & Gregor, 1955—Z. Wiener ent. Ges. 40: 85.

Gnorimoschema philolycii Hering, 1957—Bestimm. Blattminen Eur. 1-3: 26.

Scrobipalpa gallincolella auct.

Records. Povolný 2002: 43 (*gallincolella*).

Material examined. 1 ♂, Hazeva, Field School, 26.iv.1998, Malaise Trap (Ashkenazi); 1 ♂, 1 ♀, Nahal Shahaq, Shesaf Nat. Rez., 16.iv.1999, Malaise Trap (Yaromi) (all TAU).

Distribution. South Europe, Madeira, Canary Islands, North Africa, Middle East (Huemer & Karsholt 2010: 160).

***Scrobipalpa heratella* Povolný, 1967**

Scrobipalpa heratella Povolný, 1967—Acta sc. nat. Brno 1: 218.

Records. Povolný 2002: 46.

Distribution. Algeria, Tunisia, Egypt, Israel, Jordan, Saudi Arabia, Iran, Afghanistan (Povolný 2002: 46).

***Scrobipalpa voltinelloides* Povolný, 1972**

Fig. 47

Scrobipalpa voltinelloides Povolný, 1972: 231—Acta sc. nat. Brno 1: 231.

Records. Povolný 1972: 231.

Material examined. Holotype of *Scrobipalpa voltinelloides* ♂, Jericho (Pal), Lichtfang, 31.v.1930 (Amsel) (gen. slide A.795). Paratype of *Scrobipalpa voltinelloides* ♀, Jericho (Pal), Lichtfang, 1.iv.1930 (Amsel) (gen. slide A.816); 1 ♂, Jordanbrucke bei Jericho, 29.iv.1930, Lichtfng. (Amsel) (gen. slide A.828) (all SMNK).

Distribution. Israel.

***Scrobipalpa usingeri* Povolný, 1969**

Scrobipalpa usingeri Povolný, 1969—Acta sc. nat. Brno 3 (12): 19.

Records. Povolný 2002: 75.

Material examined. 1 ex, Jordanbrucke bei Jericho, 29.iv.1930, Lichtfng. (Amsel) (gen. slide A. 824) (SMNK).

Distribution. Saudi Arabia, Israel, Mongolia (Povolný 2002: 75). A record from Volgograd region of Russia (Povolný 1980: 250) needs confirmation (Huemer & Karsholt 2010: 83).

***Scrobipalpa traganella* (Chrétien, 1916)**

Lita traganella Chrétien, 1915—Ann. Soc. ent. Fr. 84: 321.

Records. Povolný 1967: 230.

Material examined. 8 ex., Jordanbrucke bei Jericho, 29.iv.1930, Lichtfng. (Amsel) (gen. slides A.832, A.804; A.830); 2 ex., Südende d. Toten Meers, 15-27.iii.1933 (Amsel) (gen. slides A.839, A.840) (all SMNK); 2 ♂, 1 ♀, Jericho, Kalia, v.2004 (Kravchenko) (genitalia slide 5707 Hendriksen) (TAU, ZMUC).

Distribution. Spain (with Canary Islands), Portugal (with Madeira), South France, Cape Verde, North Africa, Israel, Lebanon, Syria, Saudi Arabia, Pakistan (Povolný 2002: 74; Huemer & Karsholt 2010: 137).

***Scrobipalpa superstes* Povolný, 1977**

Scrobipalpa superstes Povolný, 1977—Acta ent. bohemoslov. 74: 189, figs. 10-14, 31.

Records. Povolný 1972: 189 as *Scrobipalpa otregata* (misidentification).

Material examined. Paratype of *Scrobipalpa otregata* ♀, Jordanbrucke, 10.iv.1930 (gen. slide A.808); Paratype: 1 ♀, Jordanbrucke bei Jericho, 10.iv.1930; PT, 29.iv.1930, Lichtfang (gen. slide A. 827) (all SMNK).

Additional material. 2 ♂, 3 ♀, 76 exx, Greece, Lesvos, 7 km E Skala Kallonis, salt pans, 1 m, 15.x.2008 (Kaila & Kullberg) (genitalia slides Karsholt 5208, 5210, 5211, 5213) (MZH, ZMUC).

Distribution. Portugal, Spain, Italy (Sardinia, Sicily) (Huemer & Karsholt (2010: 115), Greece (Lesvos) (new record), Israel (new record).

Remarks We were able to examine a large series from the Greece Island of Lesvos, which fits *S. superstes* both in genitalia and in DNA barcode (M. Mutanen *in litt.*).

Scrobipalpa indignella (Staudinger, 1879)

Fig. 48

Bryotropha indignella Staudinger, 1879—Horae Soc. Ent. Ross. 15(2–3): 308.

Gnorimoschema hyoscyamivora Gerasimov, 1940—Opredelitel roda *Phthorimea* povrezhdajushchih kartofel, tomaty i drugie kultury semeistva paslenovykh: 77.

Gnorimoschema pseudoboletellum Povolný & Gregor, 1955—Z. Wiener ent. Ges. 40: 83.

Scrobipalpa grossa Povolný, 1966—Acta ent. bohemoslov. 63: 400.

Records. Povolný & Gregor 1955: 83 (*pseudoboletella*).

Material examined. Holotype of *pseudoboletellum* ♀, Südenende d. Toten Meers, 15–27.iii.1933 (Amsel) (gen. slide GU. 3202) (SMNK).

Distribution. Ukraine, Russia (Stavropolskiy krai, Volgograd region, Orenburg region), Turkey, Syria, Israel, Saudi Arabia, Iraq, Afghanistan, Turkmenistan, Western China (Xinjiang) (Povolný 2002: 64; Bidzilya & Budashkin 2009: 17; Bidzilya & Li 2010: 5; Huemer & Karsholt 2010: 124; Junnilainen *et al.* 2010: 48).

Scrobipalpa obsoletella (Fischer von Röslerstamm, 1841)

Lita obsoletella Fischer von Röslerstamm, 1841—Abbildungen Schmettkde: 225, pl. 79.

Phthorimaea bipunctella Hartig, 1941—Mitt. Münch. Ent. Ges. 31: 158.

Phthorimaea calaritanella Amsel, 1952—Fragm. ent. Roma 1: 128.

Scrobipalpa obsoletella hospes Povolný, 1964—Acta Soc. ent. Cechoslov. 61: 354.

Records. Amsel 1935b: 211; Bodenheimer 1937: 100.

Material examined. 2 ♀ (abdomen missing), Südenende d. Toten Meers, 15–27.iii.1933 (Amsel) (SMNK).

Distribution. Palaearctic region eastwards to Mongolia, North China and Zabaikalskiy krai of Russia, North America (?), South Africa (Bidzilya & Li 2010: 6; Huemer & Karsholt 2010: 87).

Scrobipalpa pulchra Povolný, 1967

Scrobipalpa pulchra Povolný, 1967—Acta sc. nat. Brno 1: 223.

Records. Povolný 1967: 224; Huemer & Karsholt 2010: 167.

Material examined. 1 ♂, Jordanbrucke, 29.iv.1930 (A.796); 1 ♀, Toten Meer (Amsel) (all SMNK); 2 ♂, 2 ♀, Hazeva, Field School, 18, 20.iv, 4.v.1998, Malaise Trap (Ashkenazi) (all TAU); 1 ♂, 1 ♀, 10 exx, Jericho, Kalia, v.2004 (Kravchenko) (gen. slide 5495 Hendriksen) (TAU, ZMUC); 1 ♀, Nahal (Wadi) Zin, black light, 8.x.1999 (Pavliček) (gen. slides 19005, 19006, J. Šumpich) (NMPC).

Distribution. Latvia, Eastern Ukraine, South Ural Mountains of Russia, Turkey, Israel, Central Asia to Mongolia (Huemer & Karsholt 2010: 267; Bidzilya *et al.* 2011: 66).

Scrobipalpa suaedivorella (Chrétien, 1915)

Lita suaedivorella Chrétien, 1915—Ann. Soc. ent. Fr. 84: 320.

Lita detersipunctella Toll, 1947—Z. wien. ent. Ges. 32: 114, pl. 4, fig. 23.

Material. 1 ♂, 1 ♀, Nabi Musa, 27.i, 18.ii; 2 ♀, Birkat Zefira, 9.ii.2016, ex *Suaeda asphaltica* (Keidar); 1 ♂, Mirpe, Jericho, 18.i.20-16 (Dorchin); 1 ♂, Arad, 9.ii.2016, ex *Suaeda asphaltica* (Kedar) (all TAU).

Host plant. The larvae were recorded feeding on various species of *Suaeda* (Falkovitsh & Bidzilya 2006: 75; Huemer & Karsholt 2010: 142), *Salicornia* sp. and *Salsola tetragona* Delile (Povolný 2002: 72). In Israel the species was bred from *Suaeda asphaltica* (Boiss.) Boiss. (new host plant record).

Distribution. Spain, Russia (Volgograd region) (Huemer & Karsholt 2010: 142), Tunisia, Algeria, Turkey, Iran (Povolný 2002: 72), Kazakhstan, Turkmenistan, Uzbekistan, (Falkovitsh & Bidzilya 2009: 77), Israel (new record).

***Scrobipalpa halymella* (Millière, 1864)**

Gelechia halymella Millière, 1864—Icon. Desc. Chenilles Lepid. 1: 352, pl. 42, fig. 4–8.

Records. Huemer & Karsholt 2010: 185.

Material examined. 6 ♂, 7 ♀, 168 exx, Jericho, Kalia, v.2004 (Kravchenko) (gen. slides 5491, 5492, 5494, Hendriksen) (TAU, ZMUC).

Distribution. Portugal, South France (with Corsica), Malta, Italy, Canary Islands, Morocco, Tunisia, Israel (Huemer & Karsholt 2010: 185).

***Scrobipalpa camphoromella* Nel, 1994**

Scrobipalpa camphoromella Nel, 1999—Alexanor 20 (1998): 462.

Records. Huemer & Karsholt, 2010: 187.

Material examined. 1 ♂, Jericho, Kalia, v.2004 (Kravchenko) (gen. slides 5705, Hendriksen) (TAU).

Distribution. Spain, South France, Greece, Israel (Huemer & Karsholt 2010: 186–187).

***Scrobipalpa instabilella* (Douglas, 1846)**

Anacamptis instabilella Douglas, 1846—Zoologist 4: 1270.

Lita lagunella Chrétien, 1910—Naturaliste 32: 272.

Lita strobilacella Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 101.

Lita salsolella Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 301.

Phthorimaea halymiphaga Amsel, 1952—Fragm. ent. Roma 1: 127.

Records. Amsel 1933: 125 (*salsolella*); Amsel 1935a: 301 (*salsolella*); Amsel 1935c: 263 (*salsolella*); Bodenheimer 1937: 100 (*salsolella*).

Material examined. 1 ♂, 2 ♀, Jordanbrucke bei Jericho, 10, 29.iv.1930, Lichtfng. (Amsel) (gen. slides A.807, A.815, A.802); 1 ♀, Jericho (Pal), Lichtfang, 31.v.1930 (gen. slide A.822); 1 ♂, Georgskloster, Wadi el Kelt, 21.v.1930 (gen. slide A.835); 1 ♀, Südende d. Toten Meers, 15-27.iii.1933 (Amsel) (gen. slide A.841) (all SMNK).

Host plant. In Israel the larvae were recorded feeding on *Salsola orientalis* Gmel. (as *S. rigida* Pall.) (Amsel 1935a: 301). It is described by Amsel (1935c: 263).

Distribution. Western and southern Europe, North Africa, Israel (Huemer & Karsholt 2010: 189).

Scrobipalpa ergasima (Meyrick, 1916)

Phthorimaea ergasima Meyrick, 1916—Exot. Microlep. 1 (18): 568.

Teleia hyoscyamella Rebel, 1912—Dt. ent. Z. Iris 26 (1): 89 (preocc. *Lita hyoscyamella* Stainton).

Lita mignatella Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 100 (Chrétien).

Phthorimaea intestina Meyrick, 1921—Ann. Transv. Mus. 8 (2): 74.

Gnorimoschema mirabile Povolný & Gregor, 1955—Z. Wiener ent. Ges. 40: 85.

Gnorimoschema pervada Clarke, 1962—Ent. News 73: 101.

Scrobipalpa epithymella auct.

Records. Bodenheimer 1930a: 174; Bodenheimer 1930b : 323; Amsel & Hering 1931: 147; Bodenheimer 1937: 100; Bytinski-Salz 1966: 34 (all as *Lita epithymella* Stgr., misidentification).

Material examined. 1 ♀, Jericho, 31.v.1930 (Amsel) (gen. slide GU 320y) (SMNK); 1 ♀, Jericho, Kalia, v.2004 (Kravchenko); 1 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (all TAU).

Host plant. Apest on *Solanum melongena* L. (Bodenheimer 1930a: 174; Amsel & Hering 1931: 147; Bytinski-Salz 1966: 34).

Distribution. South of Palaearctic region to Japan, Africa, South-East Asia (Huemer & Karsholt 2010: 192).

Scrobipalpa aravensis sp. nov.

Material. Holotype ♀, ISRAEL, Shezaf N.R., Nahal Shahaq, 25.iii.1999, Malaise Trap (Yarom) (gen. slide 136/18, O. Bidzilya) (TAU). Paratype: 1 ♂, same data as for holotype (gen. slide 127/15, O. Bidzilya) (TAU).

Diagnosis. The small species is defined externally by the white forewing with three dark brown fasciae and black spot in middle and in the corner of cell. *S. brandti* Povolný, 1972 and *S. sinevi* Piskunov, 1900 are lighter, without black markings, the fasciae are light brown rather than dark brown. The transverse posterior margin of vinculum in combination with sub-triangular sacculus and narrow saccus are characteristic for the male genitalia. *S. acuminatella* (Sircom, 1850) is similar in having the transverse posterior margin of vinculum but differs in sub-rectangular rather than sub-triangular sacculus, the broader saccus and tapered rather than sub-rectangular vincular process (see for comparison Huemer & Karsholt 2010: 375, fig. 17). *S. heratella* Povolný, 1967 has somewhat similar posterior margin of vinculum and vincular process but differs in the longer uncus, larger sacculus and weakly inflated caecum of the phallus (Povolný 2002: Pl. 8, fig. 73). The female genitalia can be recognized by the strongly edged anteromedial emargination of sternum VIII and narrow medially curved signum. Some other species of *Scrobipalpa* (e.g. *S. albofusca* Povolný, 1971, *S. sindibad* Povolný, 1981 and *S. superstes* Povolný, 1977) have somewhat similar female genitalia but differ in the above mentioned and some other characters.

Description. Adult (Fig. 36). Wingspan. 7.0 mm (♀)—7.7 (♂) mm. Head, thorax and tegulae white with single yellow or light brown-tipped scales. Labial palpus white with light brown basal and apical rings, inner surface of segment 2 entirely white, scape brown, antennal segment brown and white ringed; forewing white, dorsal margin weakly mottled with light brown, three transverse dark brown fasciae at base, before middle and at 2/3 length, diffuse black spot with few ochreous scales in mid-width within the second fascia, several black scales in the corner of cell, apex mixed with brown, cilia white, brown-tipped. Hindwing light grey.

Male genitalia (Fig. 37). Uncus longer than broad, weakly narrowed, posterior margin weakly rounded; gnathos short; tegumen sub-triangular, anteromedial emargination deep, broadly rounded; valva gradually curved, apex weakly broadened, extending to the top of uncus; sacculus about 1/3 length of valva, broad at base, gradually curved inwards, distal portion narrowly tapered towards pointed tip, gap to valva big; posterior margin of vinculum transverse laterally, with deep and broad v-shaped medial emargination, vincular process short, sub-rectangular, extending below tip of sacculus; saccus slender, extending far beyond apex of pedunculus; phallus short, straight, caecum strongly inflated, rounded, half length of phallus.

Female genitalia (Fig. 38). Papillae anales sub-ovate, covered with short setae; apophyses posteriores 3.5 times as long as apophyses anteriores; segment VIII as broad as long, subgenital plates broadly separated, covered with microtrichia, with honey-comb sclerotized patches extending posteriorly from the base of apophyses anteriores to 2/3 length of medial margin, ventromedial depression weakly sclerotized, not extending beyond the anterior margin of sternum VIII, anteromedial emargination of segment VIII deep, triangular, strongly edged; apophyses anteriores as long as segment VIII; colliculum narrow, ductus bursae gradually broadened towards elongated corpus bursae; signum narrow, strongly curved in middle.

Biology. Host plant unknown. Adults have been collected in late March. Both of the localities are situated in the Arava Valley which is harsh desert in the southern part of Israel characterized by riverbed vegetation only. The annual precipitation is low, with number of rain days per year ranging from 0 to 10 (Jaffe 1988).

Distribution. Israel: Shezaf Nature reserve.

Etymology. The species is named after its type locality—Arava desert in southern Israel.

Phthorimaea operculella (Zeller, 1873)

Gelechia (? *Bryotropha*) *operculella* Zeller, 1873—Verh. Zool.-bot. Ges. Wien 23 (Abh.): 262, pl. 3, fig. 17.

Gelechia terrella Walker, 1864—List Spec. Lepid. Insects Colln Br. Mus. 30: 1024 (preocc. Denis & Schiffermüller, 1775).

Bryotropha solanella Boisduval, 1874—J. Soc. centr. Hort. Fr. 2 (8): 713.

Gelechia tabacella Ragonot, 1879—Bull. Soc. ent. Fr. 5 (9): cxlvi.

Gelechia sedata Butler, 1880—Cist. Ent. 2: 560.

Gelechia piscipellis Howard, 1897—U.S. Dept. Agric. Div. Ent. Bull. 7: 76.

Phthorimaea argentinae Povolný, 1989—Steenstrupia 15: 95.

Records. Bodenheimer 1930a: 174 (*solanella*); Amsel 1933: 125 (*solanella*); Bodenheimer 1937: 100; Bytinski-Salz 1966: 34; Yathom & Meisner 1966: 167.

Material examined. 2 ex., Daganya, (Palmoni) (SMNK); 1 ♀, Beth Dagon, 30.ix.1964 (Shoham); 5 ♀, Rehovoth, Volkomi, ix.1978 (Kaplan); 1 ♀, Tel Aviv, 15.vii.1975 (Kaplan); 1 ♀, Hazeva, Field School, 26.iv.1998, Malaise Trap (Ashkenazi); 1 ♂, slopes by Yflach, 31.v.2004 (Kravchenko) (all TAU).

Host plant. In Israel larvae feed on *Solanum tuberosum* L. and *S. melongena* L.

Distribution. Neotropical species that has been introduced into all continents except Antarctica (CABI 2019).

Tuta absoluta (Meyrick, 1917)

Phthorimaea absoluta Meyrick, 1917—Trans. Ent. Soc. Lond. 1917 (1): 45.

Records. Seplyarsky *et al.* 2010: 445.

Material examined. 1 ♂, 2 ♀, Keshet, 21.iv.2018, at light (Kravchenko & Bidzilya) (TAU, ZMKU).

Host plant. *Lycopersicum* sp. (Seplyarsky *et al.* 2010: 445).

Distribution. Neotropical species that was introduced to Europe in 2007 (Huemer & Karsholt 2010: 206). Now known from most countries of Africa, as well as from Middle East, Central and South Asia (CABI 2019).

Ephysteris promptella (Staudinger, 1859)

Gelechia (*Lita*) *promptella* Staudinger, 1859—Stettin ent. Ztg 20 (7–9): 241.

Tinea despectella Walker, 1863—List Spec. Lepid. Insects Colln Br. Mus. 28: 477.

Gelechia petiginella Mann, 1867—Verh. zool.-bot. Ges. Wien 17: 843.

Doryphora parvula Staudinger, 1879—Horae Soc. Ent. Ross. 15 (2–3): 323.

Aristotelia cacomicra Walsingham, [1908]—Proc. Zool. Soc. Lond. 1907: 931.

Ephysteris chersaeta Meyrick, 1908—Proc. Zool. Soc. Lond. 1908: 725.

Ephysteris oschophora Meyrick, 1910—Rec. Ind. Mus. 5: 219.

Phthorimaea dispensata Meyrick, 1921—Ann. Transv. Mus. 8 (2): 73.

Phthorimaea fanatica Meyrick, 1921—Ann. Transv. Mus. 8 (2): 73.

Metzneria xanthorhabda Gozmány, 1951—Folia ent. hung. 4 (3): 20.

Ephysteris (*Ephysteris*) *promptella australiae* Povolný, 1977—Acta Ent. Mus. Nat. Pragae 39: 423.

Material examined. 1 ♀, Kabri, 16 km NE Haifa, 12.x.1971 (Gerling); 1 ♀, Abu-Gorke, 23.vi.1968; 1 ♀, Kefar-Shemud, 28.x.1968 (all TAU).

Distribution. South Europe, Canary Islands, Madeira, Middle East, Pakistan (Huemer & Karsholt 2010: 210), South China (Li & Bidzilya 2008: 47), Afrotropical and Indo-Australian regions (Povolný 2002: 87), Israel (new record).

Ephysteris arabiae Povolný, 1968

Ephysteris arabiae Povolný, 1968—Acta sci. nat. Brno 2 (3): 4.

Material examined. 3 ♀, Michmoret, 21.x.1968; 6 ♀, Bat Shlomo, 28.viii.1968 (all TAU).

Distribution. Saudi Arabia, Bahrein (Povolný 2002: 88), Israel (new record).

Ephysteris iberica Povolný, 1977

Ephysteris (Opacopsis) iberica Povolný, 1977—Acta ent. bohemoslov. 74 (3): 195.

Records. Huemer & Karsholt 2010: 220.

Material examined. 2 ♂, 2 exx, Jerusalem, 6.vi.2004, leg. Kravchenko, gen. slide 5498 Hendriksen (TAU, ZMUC); 1 ♂, Nahal Oren, Mt. Carmel, light trap 30.x.1997 (Pavliček & Kravchenko); 1 ♂, ditto but 5.vii.1999; 1 ♂, ditto but 11.vii.1997; 2 ♂, ditto but 27.v.1999; 1 ♀, Nahal Oren, Mt. Carmel, light trap 17.v.1997 (Pavliček & Kravchenko) (all NMPC).

Distribution. Widely distributed in the Mediterranean region from France and Spain through Croatia to Crete and Israel (Huemer & Karsholt 2010: 220).

Ochrodia subdiminutella (Stainton, 1867)

Gelechia subdiminutella Stainton, 1867—Tineina of Syria and Asia Minor: 45.

Gelechia jamaicensis Walsingham, 1897—Proc. Zool. Soc. Lond. 1897: 76.

Gnorimoschema bucolica Meyrick, 1904—Proc. Linn. Soc. N.S.W. 29 (2): 317.

Lita zygophyllella Rebel, 1912—Dt. ent. Z. Iris 26 (1): 89.

Phthorimaea ericnista Meyrick, 1914—Ann. S. Afr. Mus. 10 (8): 245.

Aristotelia ferritincta Turner, 1919—Proc. R. Soc. Qd 31 (10): 115.

Phthorimaea ochrodeta Meyrick, 1923—Exot. Microlep. 3 (1–2): 23.

Phthorimaea extorris Meyrick, 1923—Exot. Microlep. 3 (1–2): 50.

Phthorimaea crocoleuca Meyrick, 1923—Exot. Microlep. 3 (1–2): 51.

Lita unitella Turati, 1930—Atti Soc. ital. Sci. nat. 69: 78.

Aristotelia tribulivora Dumont, 1931—Bull. Soc. ent. Fr. 1931: 149.

Aristotelia pulverea Janse, 1950—Moths S.Afr. 5 (2): 115.

Scrobipalpa turgida Janse, 1951—Moths S.Afr. 5 (3): 218.

Gnorimoschema infallax Gozmány, 1960—Ann. hist.-nat. Mus. nat. hung. 52: 417.

Gnorimoschema tractatum Gozmány, 1960—Ann. hist.-nat. Mus. nat. hung. 52: 418.

Records. Stainton 1867: 46; Meyrick 1923: 23 (*ochrodeta*); Bodenheimer 1930b: 377 (*ochrodeta*); Amsel 1933: 125 (*suasoria*); Amsel 1935c: 263 (*ochrodeta & suasoria*); Bodenheimer 1937: 101 (*ochrodeta & suasoria*).

Material examined. 1 ♂, Jericho (Pal), Lichtfang, 30.iv.1930 (gen. slide Am. 4853); 1 ♂, Jericho (Pal), Lichtfang, 11.iv.1930 (gen. slide A. 790); Jericho Z. 307, Balanitis aegyptica, 7-25.v.1930 (gen. slides A. 285, A. 284); 1 ♂, Tel-Aviv (Dune), 7.iii.1932 (Einsler) (gen. slide A. 844); 1 ♂, Georgskloster, Wadi el Kelt, Lichtfang, 1.iv.1930 (gen. slide A. 818); 1 ♀, same but 21.v.1930 (gen. slide A. 814); 1 ♂, same data but 11.iv.1930 (gen. slide A. 799) (all SMNK); 1 ♂, Hazeva, Field School, 6.iv.1998, Malaise Trap (Ashkenazi) (gen. slide 161/18, O. Bidzilya); 1 ♀, Ein Gedi, 17.viii.1968 (gen. slide 125/18, O. Bidzilya) (all TAU); 2 ♂, 2 ♀, 8 exx, Jericho, Kalia, v.2004 (Kravchenko) (gen. slide 5706 Hendriksen); 2 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (all TAU, ZMUC).

Host plant. *Balanites aegyptiaca* L. (Delile) (Bodenheimer 1930b: 377; Amsel & Hering, 1931: 124).

Distribution. South Europe, subtropical and tropical Africa and Asia (Huemer & Karsholt 2010: 222).

Remarks. *Gelechia subdiminutella* was described from two males and one female from Jaffa (“Jeffa”) and “plains of Jordan” (Stainton 1867: 45). The type specimens in NHMUK match the specimens listed above from Jericho, Kalia.

Phthorimaea crocoleuca was described from one male, which Buxton bred from a larva feeding on *Balanites*

aegyptiaca by Jericho (Meyrick 1923: 51). The holotype has lost its abdomen (Clarke 1969: 160–161), but externally matches well *O. subdiminutella*.

The genus *Ochrodia* is in need of revision. Povolný (e.g., 1972: 93–94) was unable to separate the species and synonymized all *Ochrodia* species under *O. subdiminutella*. A revision of the genus is outside the scope of this study, but we notice that our material includes at least two *Ochrodia* species.

***Microlechia rhamnifoliae* (Amsel & Hering, 1931)**

Fig. 49

Lita rhamnifoliae Amsel & Hering, 1931—Dt. ent. Z. Iris 1931: 142, pl. 1, fig. 1.

Records. Amsel & Hering, 1931: 142; Amsel, 1933: 125; Amsel, 1935a: 301; Bodenheimer 1937: 100.

Material examined. “Lectotype” ♂, here designated, “Jerusalem, ex l. *Rhamnus palästina*, 6.iv.1930, Z. 248, H. Amsel” | Typus, leg. H. Amsel, *Lita rhamnifoliae*” | (gen. slide Am. 5171) | Lectotypus, *Megalocypha rhamnifoliae* Amsel, teste Povolný, 1976”. Paralectotypes: 2 ♂, Tel-Aviv, Z. 225, *Rhamnus palästina*, 29.iii.1930, H. Amsel (gen. slide Am.291, Am.254); 1 ex, Jerusalem, ex.l., *Rhamnus palästina*, 6.iv.1930, z.248 (Amsel) (gen. slide Am.255) (all SMNK).

Host plant. *Rhamnus lycioides* subsp. *graeca* (Boiss. & Reut.) Tutin (as *R. palastina* Boiss.) (Amsel & Hering, 1931: 142).

Distribution. Canary Island, Morocco, Greece, Israel, Saudi Arabia, South Africa (Povolný 2002: 100; Huemer & Karsholt 2010: 227).

Remarks. *Lita rhamnifoliae* was first described based on larvae living on “*Rhamnus palästina*” found at Yarkon by Tel-Aviv, Georgskloster by Jericho and near Jerusalem in February and March 1930. Later this species was described again (Amsel 1935a: 301) based on an unstated number of adults reared most likely from the same larvae which were mentioned in the first description.

Povolný (1978: 47, 48) stated that type material of larvae is not preserved and selected a neotype from series of specimens mentioned by Amsel in the “second” description (Amsel 1935a: 301).

We traced four specimens in SMNK labelled as cited above (Fig. 48). The collecting days given on their labels differ from those in the original description and probably represent the days when the adults emerged. One of them, dated 6.iv.1930, is labelled as lectotype and bears additional label “Neotypus”. We consider this specimen as part of the original type series, and hence we set aside the neotype designated by Povolný (*op cit.*) (ICZN 1999: article 75.8). At the same time we designate this specimen as lectotype of *Lita rhamnifoliae* Amsel & Hering, 1931 in order to stabilize nomenclature.

***Microlechia chretieni* Turati, 1924**

Microlechia chretieni Turati, 1924—Atti Soc. ital. Sci. nat. 63: 163, pl. 6, fig. 5.

Gelechia micradelpha Walsingham, 1900—Ent. mon. Mag. 36: 217 (preocc. *Gelechia micradelpha* Lower, 1897);

Phthorimaea microcasis Meyrick, 1929—Exot. Microlep. 3 (17): 532 (repl. *Gelechia micradelpha* Walsingham, 1900)

Hedma abzacella Dumont, 1932—Soc. ent. Fr (Livre centen.): 714, fig. 18–22.

Teleia hyoscyamella Amsel & Hering, 1931—Dt. ent. Zs 1931: 132, pl. 1, fig. 5; homonym of *Teleia hyoscyamella* Rebel, 1912.

Teleia hyoscyami Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 299 pl. 12, fig. 147.

Megalocypha polioptera Janse, 1960—Moths S.Afr. 6 (2): 197.

Records. Amsel & Hering 1931: 132 (*hyoscyamella*); Amsel 1933: 125 (*hyoscyami*); Amsel 1935a: 299 (*hyoscyami*); Amsel 1935c: 263 (*hyoscyami*); Bodenheimer 1937: 101 (*hyoscyami*).

Material examined. Syntypes of *Teleia hyoscyami*: 1 ♂, 1 ♀, Jerusalem, ex la. 26.iii.1930, *Hyoscyamus* (Amsel) (UMB); syntypes of *Teleia hyoscyami*: 1 ♀, 2 ex., Jerusalem, Z. 232, 26.iii.1930, *Hyoscyamus* (Amsel) (gen. slide Am.5172, Am.554); 2 ex., Jerusalem, 6.iii.1930 (Amsel) (gen. slide Am.5174, Am.555) (all SMNK); 1 ♀, Jericho (Pal), Lichtfang, 16.iv.1930 (A. 831); 1 ♀, Ojerrasch., Transjord., 7.iv.1930 (gen. slide A. 825) (all SMNK); 1 ♀, Shezaf N.R., Nahal Shahaq, 25.iii.1999, Malaise Trap (Yarom) (TAU).

Host plant. *Hyoscyamus aureus* L. (Amsel & Hering 1931: 132).

Distribution. Portugal, Spain, South France, Italy (Sardinia), Croatia, Crete, Canary Islands, Tunisia, Algeria, Syria, Israel, Saudi Arabia, Bahrein, Iran, Afghanistan, South Africa (Povolný 2002: 99; Huemer & Karsholt 2010: 228).

***Caryocolum cauligenella* (Schmid, 1863)**

Gelechia cauligenella Schmid, 1863—Berl. ent. Z. 7: 63.

Records. Caradja 1920: 103; Bodenheimer 1937: 100; Huemer 1988: 479.

Distribution. Europe, Middle East (Huemer & Karsholt 2010: 256), Israel.

***Caryocolum proxima* (Haworth, 1828)**

Recurvaria proxima Haworth, 1828—Lepid. Britannica (4): 552.

Gelechia maculiferella Douglas, 1851—Trans. ent. Soc. Lond. (n.s.) 1: 102 (repl. *Recurvaria proxima* Haworth, 1928).

Gelechia maculivicinella Bruand, 1859—Ann. Soc. ent. Fr. (3) 6: 652.

Gelechia horticolla Peyerimhoff, 1871—Mitt. schweiz. Ent. Ges. 3: 411.

Records. Huemer & Karsholt 2010: 280.

Material examined. 1 ♂, slopes by Yflach, 31.v.2004 (Kravchenko) (gen. slide 5703, Hendriksen) (TAU).

Distribution. Europe, Turkey, Israel, U.S.A (presumably introduced) (Huemer & Karsholt 2010: 280).

***Caryocolum schleichi* (Christoph, 1872)**

Lita schleichi Christoph, 1872—Horae Soc. ent. Ross. 9: 22.

Caryocolum syriacum Povolný, 1977—Acta Univ. Agr. Brno (A) 25 (4): 171.

Lita dianthella Chrétien, 1925—Amat. Papillons 2: 246.

Caryocolum hackeri Derra, 1985—Entomofauna 6 (22): 373.

Material examined. 1 ♂, Nahal Oren, Mt. Carmel, light trap, 30.x.1997 (Pavlíček & Kravchenko) (gen. prep. 19649, J. Šumpich) (NMPC).

Distribution. Europe to Mongolia and southern Siberia, Turkey, Syria, Jordan, Afghanistan (Huemer & Karsholt 2010: 265), Israel (new record).

Tribe Litini Bruand, 1859

***Recurvaria nanella* (Denis & Schiffermüller, 1775)**

Tinea nanella Denis & Schiffermüller, 1775—Ank. syst. Schmett. Wienergegend: 141.

Records. Bodenheimer 1937: 101.

Host plant. In Israel the larvae were recorded feeding on *Prunus* sp. (Bodenheimer, 1937: 101).

Distribution. Europe, North Africa, Central Asia, North America (presumably introduced) (Huemer & Karsholt 1999: 37), Lebanon (Zerny 1934: 20), Israel.

Stenolechia gemmella (Linnaeus, 1758)

Phalaena (*Tinea*) *gemmella* Linnaeus, 1758—Syst. Nat. (Edn 10) 1: 539.
Recurvaria nivea Haworth, 1828—Lepid. Britannica (4): 554 (emend.).
Lita nigrovittella Duponchel, [1839]—in Godart, Hist. Nat. Lépid. Fr. (Suppl.) 3: 315.
Gelechia (*Brachmia*) *lepidella* Zeller, 1839—Isis [32] 1839 (3): 202.

Records. Amsel 1933: 126; Amsel 1935c: 264; Bytinski-Salz & Sternlicht 1967: 130; Bodenheimer 1937: 101.

Material examined. Jordan: 1 ♀, Jordan Valley, Zerqa R. Colony, c. 100 m below S.L., 12.iv.1953, at light (Trough); 1 ♀, ditto but 18.ii.1954; 1 ♀, ditto but 18.i.1953 (gen. slide GU2281); 1 ♂, ditto but 19.xii.1952 (gen. slide GU2280) (all SMNK).

Host plant. In Israel larvae feed on *Quercus coccifera* L. (as *Q. calliprinos* Webb) (Bytinski-Salz & Sternlicht 1967: 130).

Distribution. Europe to Caucasus (Huemer & Karsholt 1999: 44), Israel, Jordan (new record).

Parastenolechia nigrinotella (Zeller, 1847)

Gelechia nigrinotella Zeller, 1847—Isis von Oken 1847 (11): 856.

Material examined. 1 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (TAU); 6 ♂, 1 ♀, Nahal Oren, Mt. Carmel, light trap 24.vi.1997 (Pavliček & Kravchenko); 6 ♂, 2 ♀, ditto but 19.vi.1998 (gen. prep. 19646, J. Šumpich) (all NMPC).

Distribution. Central and Southern Europe, Turkey (Huemer & Karsholt 1999: 46), Lebanon (Zerny 1934: 21), Israel (new record).

Carpatolechia decorella (Haworth, 1812)

Tinea decorella Haworth, 1812—Trans. Ent. Soc. Lond. 1: 338.
Gelechia humeralis Zeller, 1839—Isis [32] 1839 (3): 200.
Lita incretella Duponchel, [1845]—Cat. méth. lépid. Eur.: 338.
Anacamptis lyellella Humphreys & Westwood, 1845—Br. Moths 2: 190, pl. 106, fig. 10.
Lita humeralella Bruand, [1851]—Mém. Soc. Doubs 3 (3, livr. 5–6): 46.
Lita marmoripennella Bruand, [1851]—Mém. Soc. Doubs 3 (3, livr. 5–6): 47 [?81].
Gelechia pisticella Nowicki, 1860—Enum. Lepid. Haliciae Orient.: 188.
Gelechia scabra Staudinger, 1870—Horae Soc. ent. ross. 7 (1870): 254.
Gelechia erschoffii Frey, 1880—Lep. Schweiz.: 361; TL: Wallis.
Teleia subericolella Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 104.
Gelechia buckwelli Lucas, 1956—Bull. Soc. Sci. nat. Maroc 35: 256.
Carpatolechia dimitrescui Capuse, 1964—Ent. Tidskr. 85: 13, fig. 1–4.

Records. Halperin & Sauter 1992: 119, 135.

Material examined. 1 ♂, 1 ♀, Shoresh, 8.iv.1976, *Pistacia palaestina*, 13.v.1976 (Halperin); 2 ex, Jerusalem, 8.iv.1988, *Pistacia palaestina*, p. 12.iv, em. 4.v (Halperin); 1 ex, Ein Hemed, 7.iv.1976, *Pistacia palaestina*, in blossom, pupa 24.iv, em. 15.v.1976 (Halperin); 1 ♂, Nahal Amud, 19.iv.1976, *Pistacia palaestina*, pup. 6.v, em. 22.v.1976 (Halperin); 1 ex, Israel, Shaar Hagai, 8.iv.1976, *Pistacia palaestina*, pup. 15.ix, em. 13.v.1976 (Halperin); 2 ♂, Jerusalem, 6.vi.2004 (Kravchenko) (all TAU); 1 ♂, Haifa—Carmelia, light trap 3.x.1999 (Pavliček); 4 ♂, 7 ♀, Nahal Oren, Mt. Carmel, light trap 26.ii.1998 (Pavliček & Kravchenko); 1 ♂, ditto but 27.v.1999; 1 ♀, ditto but 24.vi.1997; 2 ♀, ditto but 25.ii.1999; 1 ♀, ditto but 1.iv.1997; 1 ♂, 1 ♀, ditto but 5.ii.2000 (Pavliček); 1 ♂, 3 ♀, ditto but 11.iii.2000 (all NMPC).

Host plant. In Israel larvae feed on *P. terebinthus* ssp. *palaestina* Boiss. (as *Pistacia palaestina* Boiss.) (Halperin & Sauter 1992: 135).

Distribution. Europe, Canary Islands, North Africa, Turkey, Israel, Kazakhstan (Huemer & Karsholt 1999: 68).

***Schneidereria pistaciella* Weber, 1957**

Schneidereria pistaciella Weber, 1957—Mitt. schweiz. ent. Ges. 30 (1): 68.

Records. Halperin & Sauter 1992: 135 (? *pistaciella*).

Host plant. In Israel the larvae have been recorded feeding in fruits of *Pistacia vera* L. (Halperin & Sauter 1992: 135).

Distrubution. Ukraine, Greece, Cyprus, Syria (Huemer & Karsholt 2001: 45). Probably also Iran and Iraq (Sattler 1982: 16), Israel.

Remarks. *Schneidereria pistaciella* can easily be separated from other species of Litini which larvae feed on *Pistacia* with exception for *Schneidereria pistaciicola* (Danilevsky, 1955). However, the latter is restricted to Central Asia. Therefore, we suggest that the questionable record of *S. pistaciella* from Israel (Halperin & Sauter 1992: 135) should most likely be referred to this species rather than to *S. pistaciicola*.

***Istrianis arenicolella* (Caradja, 1920)**

Teleia myricariella var. *arenicolella* Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 104.

Teleia amilcarella Lucas, 1933—Bull. Soc. ent. Fr. 38: 199.

Material examined. 1 ♂, Negev, Nahal (Wadi) Zin, black light, 8.x.1999 (Pavliček) (gen. slide 19010, J. Šumpich) (NMPC).

Distribution. Algeria, France, Spain (Bidzilya & Karsholt 2015: 414), Israel (new record).

***Istrianis kravchenkoi* Bidzilya & Karsholt, 2015**

Fig. 50

Istrianis kravchenkoi Bidzilya & Karsholt, 2015—Zootaxa 4059 (3): 421

Records. Bidzilya & Karsholt, 2015: 421.

Material examined. Holotype (♂) and 3 paratypes (♀) of *Istrianis kravchenkoi* Jericho, Kalia, v.2004 (Kravchenko) (gen. slides 5497, 5699, 5700, 5735 Hendriksen) (TAU, ZMKU, ZMUC).

Distribution. Israel.

***Istrianis wachtlii* (Rogenhoffer, 1881)**

Teleia wachtlii Rogenhofer, 1881—Verh. zool.-bot. Ges. Wien 30 (s.b.): 48.

Records. Amsel 1933: 125; Amsel 1935c: 263; Bodenheimer 1937: 100.

Material examined. 1 ♂, Jordanbrucke bei Jericho, 29.iv.1930, Lichtfang (Amsel) (gen. slide 37/16, O. Bidzilya) (SMNK).

Distribution. Egypt, Israel, Iran (Amsel 1959: 32; Bidzilya & Karsholt 2015: 425).

***Istrianis femoralis* (Staudinger, 1876)**

Teleia femoralis Staudinger, 1876—Stettin. ent. Ztg 37: 146.

Teleia comedonella Staudinger, 1880—Horae Soc. Ent. Ross. 15 (2–3): 312.

Teleia gravosensis Rebel, 1937—Zs. Öst. EntVer. 22 (2): 13.

Teleia angustipennis Rebel, 1941—Isv. tsarsk. prirodonauch. Inst. Sofia 14: 4.

Teleia funebrella Rebel, 1941—Isv. tsarsk. prirodonauch. Inst. Sofia 14: 5.

Records. Bidzilya & Karsholt 2015: 430.

Material examined. 1 ♀, Zomet, Elialain, 1.xi.1980 (Wool) / ex. *Smynthurodes betae* (TAU).

Distribution. Spain, Italy (incl. Sicilia), Croatia, Macedonia, Slovenia, Ukraine (Crimea), Turkey, Turkmenistan, Kyrgyzstan, Iran (Bidzilya & Karsholt 2015: 430).

Remarks. According to label, the larva was bred from galls produced by *Smynthurodes betae* Westwood, 1849 (Hemiptera, Aphididae).

***Istrianis squamodorella* (Amsel, 1935)**

Fig. 51

Pseudoteleia squamodorella Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 299, pl. 11, fig. 104–106, pl. 18, fig. 73–76.

Records. Amsel 1933: 125; Amsel 1935c: 264; Bodenheimer 1937: 101; Halperin & Sauter 1992: 125, 135.

Material examined. Lectotype of *Pseudoteleia squamodorella* ♂, Jordanbrücke, Jericho, Z. 342, *Populus euphr.*, 17.vii.1930, H.G. Amsel (gen. slide 26/11, O. Bidzilya) (SMNK). Paralectotypes of *squamodorella*: 2 ♂, 2 ♀, Kasr el Jehud, Jordan Z., 250, 9.iv.30, *Populus euphratica*, H.G. Amsel (gen. slide 26/11♀; 44/11♂, O. Bidzilya) (SMNK; NHMUK); 1 ♂, Jordanbrücke, Jericho, Z. 342, *Populus euphr.*, 17.vii.1930, H.G. Amsel (gen. slide 26/11, O. Bidzilya) (SMNK); 1 ♀, Jordanbrücke, Jericho, 29.iv.1930, Lichtfg. (Amsel) (NHMUK); 1 ♂ [abdomen missing], 1 ♀, Allen by Bridge Jericho, ex l. *Populus euphratica*, 22(?)iv.30 (Amsel) (gen. slide 400/14, O. Bidzilya) (MFN).

Host plant. In Israel the larvae have been recorded feeding on *Populus euphratica* Oliv. (Amsel 1935a: 300) and *Salix* sp. (Halperin & Sauter 1992: 125, 135).

Distribution. Israel, Jordan, Pakistan (Bidzilya & Karsholt 2015: 437).

Remarks. The type material was discussed and the lectotype was designated by Bidzilya & Karsholt (2015: 437–438).

***Teleiopsis sarcochroma* (Walsingham, 1900)**

Fig. 52

Gelechia sarcochroma Walsingham, 1900—Bull. Liverpool Mus. 3 (1): 3.

Gelechia hessi Amsel, 1935—Mitt. zool. Mus. Berl. 20 (2): 301.

Gelechia nigrorosea Walsingham, 1904—Ent. mon. Mag. 40: 266.

Records. Amsel 1933: 125 (*hessi*); Amsel 1925a: 301 (*hessi*); Amsel 1935c: 263 (*hessi*); Bodenheimer 1937: 100 (*hessi*); Sattler 1982: 23.

Material examined. Lectotype of *Gelechia hessi* ♂, Georgskloster, Wadi el Kelt, 15.iv.1930 (Amsel) | Palästina Expedition, 18.ii-4.iv.1930 (Amsel) | Lectotype. *Gelechia hessi* Amsel, ♂, teste K. Sattler, 1981; paralectotype, same data (all SMNK); 1 ♂, Hulioth, 20.vi.1969 (Shoham); 1 ♀, Negev, M. Haqatan, 25.ii.1981 (Furth) / on *Rhus tripartita* / Pupa 28.iii, em. 21.iv; 1 ♀, same data but Ma'ale Tanar, 13.iii.1981 / P. 30.iii, em. 22.iv; 1 ♂, 1 ♀, same data but Mamshit, 13.iii.1981 / P. 20, 23.iii., em. 19, 20.iv. (all TAU).

Host plant. In Israel the larvae were recorded feeding on *Searsia tripartita* (Ucria) Moffett (Sattler 1982: 23–24). One of the specimens listed above was bred from a pupa found on this plant.

Distribution. Algeria, Tunisia, Lybia, Israel, Yemen (Amsel 1961: 59; Sattler 1968: 116).

***Teleiopsis terebinthinella* (Herrich-Schäffer, 1856)**

Gelechia terebinthinella Herrich-Schäffer, 1856—Syst. Bearb. Schmett. Europ. 6: 40.

Records. Halperin & Sauter 1992: 119, 135.

Material examined. 2 ♂, 1 ♀, Jerusalem, 6.vi.2004 (Kravchenko) (genitalia slide 5698 Hendriksen) (TAU, ZMUC).

Host plant. In Israel the larvae have been recorded feeding on *Pistacia vera* L. (Halperin & Sauter 1992: 135).

Distribution. Southern and South-Eastern Europe, Turkey and the Middle East (Huemer & Karsholt 1999: 90), Israel.

***Teleiopsis diffinis* (Haworth, 1828)**

Recurvaria diffinis Haworth, 1828—Lepid. Britannica (4): 551

Lita dissimilella Treitschke, 1833—in Ochseneheimer, Schmett. Eur. 9 (2): 80

Lita friesella Zetterstedt, 1839—Insecta Lapponica: 1005

Gelechia scabidella Zeller, 1839—Isis [32] 1839 (3): 199

Gelechia diffinella [= *diffinis*] Doubleday, 1859—Zool. Syn. List Br. Butts Moths (edn 2): 30 (emend.)

Gelechia diffinis ab. *groenliensis* Strand, 1920—Archiv Naturg. 85 A (4): 65

Material examined. 1 ♂, Nahal Teqoa, 31°38'N 35°14'E, 650 m, 31.iii.2009 (Freidberg) (gen. slide 147/18, O. Bidzilya) (TAU).

Distribution. Europe, North Africa, South-Western and Central Asia (Huemer & Karsholt 1999: 91), Russia: North and Central of the European part to South Ural (Ponomarenko 2008; Junnilainen *et al.* 2010), Altaiskiy krai (Bidzilya & Nupponen 2018: 385), Israel (new record).

***Altenia mersinella* (Staudinger, 1879)**

Teleia mersinella Staudinger, 1879—Horae Soc. Ent. Ross. 14: 313

Stenolechia sagittella Caradja, 1920—Dt. ent. Z. Iris 34 (1/2): 110

Teleia melanostictella Ragonot, 1895—Bull. Soc. ent. Fr. 1895: 106

Gelechia praedicata Meyrick, 1923—Exot. Microlep. 3 (1–2): 19 (repl. *Gelechia melanostictella* Ragonot, 1895)

Telphusa tribolopsis Meyrick, 1927—Exot. Microlep. 3 (11): 344

Records. Amsel 1933: 125 (*tribolopsis*); Bodenheimer 1937: 101 (*tribolopsis*).

Material examined. 2 ♂, Karmel, Haifa, 7.v.1930 (Amsel) (SMNK); 1 ♀, Karmel, 7.ii.1981 (Wool) ex. *Aploneura* (TAU); 2 ♂, Nahal Oren, Mt. Carmel, light trap 17.v.1997 (Pavliček & Kravchenko); 1 ♂, ditto but 23.iv.1998; 1 ♂, ditto but 5.vii.1999 (all NMPC).

Host plant. According to the label one specimen was bred from galls (?) of *Aploneura* sp. (Hemiptera: Aphididae).

Distribution. Cyprus, Lebanon, Syria, Turkey (Huemer & Karsholt 2001: 47), Israel.

***Altenia wagneriella* (Rebel, 1926)**

Gelechia wagneriella Rebel, 1926—Verh. zool.-bot. Ges. Wien 74/75 (s.b.): 108

Klaussattleria danilevskyi Piskunov, 1973—Trudy Ent. Obsch. 56: 192

Material examined. 1 ♀, Nahal Oren, Mt. Carmel, light trap, 1.iv.1997 (Pavliček & Kravchenko) (gen. slide 19004, J. Šumpich) (NMPC).

Distribution. South-Eastern Europe, Turkey, Iran, Tadjikistan, Turkmenistan (Huemer & Karsholt 1999: 102), Israel (new record).

Altenia modesta (Danilevsky, 1955)

Teleia modesta Danilevsky, 1955—Ent. Obozr. 34: 114, figs. 5, 6.

Records. Halperin & Sauter 1992: 119, 135.

Host plant. In Israel the larvae have been recorded feeding on *Pistacia atlantica* Desf. (Halperin & Sauter 1992: 135).

Distribution. Algeria, Croatia, Greece, Cyprus, Central Asia (Karsholt & Huemer 1999: 103), Israel.

Stenovalva disclusa (Meyrick, 1923)

Telphusa disclusa Meyrick, 1923—Exot. Microlep. 3 (1–2): 16.

Stenovalva ghorella Amsel, 1955—Bull. Inst. Sci. nat. Belg. 31 (83): 10, pl. 1, fig. 7–9.

Records. Meyrick 1923: 16; Amsel 1933: 125; Bodenheimer 1937: 101; Sattler 1982: 24.

Material examined. 1 ♀, Samaria, western Yarihiv, 23.xi.1979 (Furth) / on *Rhus tripartita*; 1 ♀, Samaria, Sanniriya, 4.iv.1981 (Furth) / on *Rhus tripartita* / Pupated 14.iv, em. mid May; 1 ♀, W. Samaria, 1 km east Yarihiv, 11.i.1984, on *Rhus tripartita* (Furth) / pup. 25.i, em. 12.ii. (all TAU).

Host plant. In Israel larvae feed on *Searsia tripartita* (Ucria) Moffett (Sattler 1982: 24).

Distribution. Jordan (Amsel 1955a: 12), Israel.

Agnippe lunaki (Rebel, 1941)

Stenolechia lunaki Rebel, 1941—Isv. tsarsk. prirodonauch. Inst. Sofia 14: 6

Evippe penicillata Amsel, 1961—Beitr. Naturk. Forsch. SW-Dt. 20: 102

Material examined. 1 ♂, Bat-Shlomo, 20.viii.1953 (TAU); 1 ♂, 1 ♀, Jerusalem, 6.vi.2004 (Kravchenko) (TAU, ZMUC); 1 ♂, Nahal Oren, Mt. Carmel, light trap, 19.vi.1998 (Pavliček & Kravchenko) (NMPC).

Distribution. Macedonia, Greece (Crete), Turkey (Marash) (Bidzilya & Li 2010: 251), Israel (new record).

Incorrect and doubtful records for Israel

Nothris sulcella Staudinger, 1879

Records. Bodenheimer 1930a: 174.

Remarks. Bodenheimer (1930a: 174) recorded *N. sulcella* Staudinger, 1879 from Ben Schamen in April 1925, as the only *Nothris* species known from Palestine at that time. Ben Schamen is a suburb of present day Tel Aviv, and we presume that the specimen listed above from Tel Aviv under *N. verbascella* may represent Bodenheimer's *N. sulcella*. Thereby we disagree with Karsholt & Šumpich (2015: 488, 493) who suggested that *N. sulcella* sensu Bodenheimer (1930a) may belong to *N. skyvai* Karsholt & Šumpich, 2015. Both *N. sulcella* and *N. skyvai* are here removed from the list of Lepidoptera of Israel/Palestine until their presence can be confirmed.

Nothris skyvai Karsholt & Šumpich, 2015

Records. Karsholt & Šumpich (2015: 488, 493)

Remarks. See remarks under *N. sulcella*.

Anarsia spartiella (Schrank, 1802)

Records. Halperin & Sauter 1992: 134.

Remarks. The isolated record of *A. spartiella* from Israel needs verification. It should most likely be referred to another species from *A. spartiella*-group. Hence, we removed *A. spartiella* from the list of Israeli Gelechiidae until the presence of this species in Israel can be confirmed.

Megacraspedus cerussatellus Rebel, 1930

Records. Amsel, 1935b: 210; Bodenheimer 1937: 102.

Remarks. *Megacraspedus cerussatellus* is only known from the type series from the Alibotusch Mountains in South Bulgaria (Huemer & Karsholt 2018: 137–138). We were not able to examine specimens identified as *cerussatellus* from Israel, and we find it most likely that these were misidentified. It is highly unlikely that this species occurs in Israel, and it is herewith removed from the list of Israeli Gelechiidae.

Oxypteryx atrella (Denis & Schiffermüller, 1775) comb. nov.

Records. Bodenheimer 1930a: 174; Amsel 1933: 125; Bodenheimer 1937: 101.

Remarks. We did not find material supporting the occurrence of *O. atrella* in Israel. We suppose that Amsel (1933: 125) followed Bodenheimer (1930a: 174) who did not separate *O. atrella* and closely related *O. immaculatella*. Hence, we removed *O. atrella* from the list of Gelechiidae of Israel until its occurrence here can be confirmed.

Isophrictis anthemidella (Wocke, 1871)

Records. Bodenheimer 1930a: 174; Amsel 1935c: 265; Bodenheimer 1937: 101 (*striatella anthemidella*).

Remarks. We were not able to find material on which the above records were based, but we suppose that they refer to *I. magna*. We remove *I. anthemidella* from the list of Israeli Gelechiidae until its occurrence there can be confirmed.

Metzneria metzneriella (Stainton, 1851)

Records. Amsel, 1933: 124; Amsel, 1935c: 262; Bodenheimer 1937: 100.

Remarks. We did not examine material of this species from Israel, and we do not know the identity of the specimens referred to above by Amsel and Bodenheimer. At that time the genus *Metzneria* was unrevised, and several species have since been described from the Mediterranean area and from the Middle East. We therefore consider the occurrence of *M. metzneriella* as doubtful and remove this species from the list of Israeli Gelechiidae until it can be confirmed.

Scrobipalpa nitentella (Fuchs, 1902)

Records. Amsel 1935b: 211; Bodenheimer 1937: 100.

Remarks. We did not examine material of this species from Israel, and we do not know the identity of the specimens referred to above by Amsel and Bodenheimer. At that time the genus *Scrobipalpa* was unrevised, and

several species (some of them very similar to *S. nitentella*) have since been detected or described from the Mediterranean area and from the Middle East. We therefore consider the occurrence of *S. nitentella* as doubtful until it can be confirmed.

***Scrobipalpa otregata* Povolný, 1972**

Records. Povolný (1972: 189)

Remarks. Povolný (1972: 189) described *Scrobipalpa otregata* from three females: the holotype from the Elburs Mts in Iran, and two paratypes from Palestine. The paratypes were stated in the original description (Povolný 1972: 191) to be in poor condition: “Das eine Weibchen ist stark abgeflogen, ... Die beide Falter sind stark from Hyphen eines Myceliums bewachsen” [One of the females is strongly worn, ... Both moths are strongly overgrown by fungal hyphae].

Parts of the female genitalia of all three type specimens were figured in the original description. From these it is clear that the paratypes from Palestine (op cit.: 193, fig. 9, 196, fig. 20) are not conspecific with the holotype from Iran (op cit.: 196, fig. 16), which would also have been surprising as the latter was found at an altitude of 1650 m, whereas Jericho is situated at low elevation. Instead they belong *Scrobipalpa superstes*, a species mainly distributed in western Mediterranean countries, but also found further to the east (see under that species).

***Scrobipalpa remota* Povolný, 1972**

Records. Povolný 1972: 191.

Distribution. Algeria, Israel, North Iran (Povolný 1972: 191; Huemer & Karsholt 2010: 311).

Remarks. *Scrobipalpa remota* was described from five specimens of both sexes (including the holotype) from Iran, and two males from Algeria and Palestine, respectively (Povolný 1972: 192). As shown by Huemer & Karsholt (2010: 115) the male genitalia of *S. remota* are indistinguishable from those of *S. superstes* Povolný, 1977, and the occurrence of the former species in Israel/Palestine requires confirmation.

***Scrobipalpa salinella* (Zeller, 1847)**

Records. Bodenheimer 1937: 100.

Remarks. We did not examine material of this species from Israel, and we do not know the identity of the specimens referred to above by Bodenheimer. At that time the genus *Scrobipalpa* was unrevised, and several species (some of them very similar to *S. salinella*) have since been detected or described from the Mediterranean area and from the Middle East. We therefore consider the occurrence of *S. salinella* as doubtful and remove this species from the list of Israeli Gelechiidae.

***Gnorimoschema epithymella* (Staudinger, 1859)**

Records. Bodenheimer 1930a: 174; Bodenheimer 1930b : 323; Amsel & Hering 1931: 147; Bodenheimer 1937: 100 ; Bytinski-Salz 1966: 34 (all as *Lita epithymella* Stgr., misidentification).

Remarks. The record of *Lita epithymella* (Staudinger, 1859) from Israel (Bodenheimer, 1937: 100) and indication of its larvae feeding on leaves of aubergine (Bodenheimer, 1930a: 174; 1930b: 323; Amsel & Hering, 1931: 147; Bytinski-Salz, 1966: 34) should be referred to *Scrobipalpa ergasima*.

Ephysteris diminutella (Zeller, 1847)

Records. Amsel 1933: 125; Bodenheimer 1937: 100.

Remarks. We did not examine material of this species from Israel, and we do not know the identity of the specimens referred to above by Amsel and Bodenheimer. At that time the genus *Ephysteris* was unrevised, and several species have since been described from the Mediterranean area and from the Middle East. We therefore consider the occurrence of *E. diminutella* as doubtful and remove this species from the list of Israeli Gelechiidae until its occurrence there can be confirmed.

Phthorimaea suasoria Meyrick, 1918

Records. Amsel 1933: 125 (*suasoria*); Amsel 1935c: 263 (*ochrodeta* & *suasoria*); Bodenheimer 1937: 101 (*ochrodeta* & *suasoria*).

Remarks. Amsel (1933: 125) recorded the Indian *Phthorimaea suasoria* Meyrick, 1918 as “*Teleia suasoria* Meyr.” from “Palästine, Gebirge” (repeated by Bodenheimer 1937: 101). *P. suasoria* is similar to *Ochrodia subdiminutella* (Stainton, 1867) and perhaps a synonym of that species, and we presume that the record of *suasoria* from Israel/Palestine is based on misidentification and belongs to *O. subdiminutella*.

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